

APPLICATION
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TITLE: HUMANIZED ANTI-CCR2 ANTIBODIES AND
METHODS OF USE THEREFOR

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HUMANIZED ANTI-CCR2 ANTIBODIES AND METHODS OF USE THEREFOR

This application is a continuation-in-part of U.S. Patent Application No. 10/272,899, filed October 17, 2002, which claims priority to U.S. provisional application number 60/392,364 filed on June 26, 2002, and 60/350,166 filed on October 19, 2001, the contents of all of which are incorporated herein by reference

Background of the Invention

Over the past several years a growing family of leukocyte chemoattractant/activating factors, termed chemokines, has been described (Oppenheim, J.J. et al., Annu. Rev. Immunol., 9:617-648 (1991); Schall and Bacon, Curr. Opin. Immunol., 6:865-873 (1994); Baggiolini, M., et al., Adv. Immunol., 55:97-179 (1994)). Members of this family are produced and secreted by many cell types in response to early inflammatory mediators such as IL-1 β or TNF α . The chemokine superfamily comprises two main branches: the α -chemokines (or CXC chemokines) and the β -chemokines (CC chemokines). The α -chemokine branch includes proteins such as IL-8, neutrophil activating peptide-2 (NAP-2), melanoma growth stimulatory activity (MGSA/gro or GRO α), and ENA-78, each of which have attracting and activating effects predominantly on neutrophils. The members of the β -chemokine branch affect other cell types such as monocytes, lymphocytes, basophils, and eosinophils (Oppenheim, J.J. et al., Annu. Rev. Immunol., 9:617-648 (1991); Baggiolini, M., et al., Adv. Immunol., 55:97-179 (1994); Miller and Krangel, Crit. Rev. Immunol., 12:17-46 (1992); Jose, P.J., et al., J. Exp. Med., 179:881-118 (1994); Ponath, P.D., et al., J. Clin. Invest., 97:604-612 (1996)), and include proteins such as monocyte chemotactic proteins 1-4 (MCP-1, MCP-2, MCP-3, MCP-4 and MCP-5), RANTES, and macrophage inflammatory proteins (MIP-1 α , MIP-1 β). Recently a new class of membrane-bound chemokines designated CX3C chemokines has been identified (Bazan, J.F., et al., Nature 385:640-644 (1997)). Chemokines can mediate a range of pro-inflammatory effects on leukocytes, such as triggering of chemotaxis, degranulation, synthesis of lipid mediators, and integrin activation (Oppenheim, J.J. et al., Annu. Rev. Immunol., 9:617-648 (1991); Baggiolini, M., et al., Adv. Immunol., 55:97-179 (1994); Miller, M.D. and Krangel, M.S., Crit. Rev. Immunol., 12:17-46 (1992)). Lately, certain β -chemokines have been shown to

suppress HIV-1 infection of human T cell lines in vitro (Cocchi, F., et al., Science (Wash. DC), 270:1811-1815 (1995)).

Chemokines bind to 7 transmembrane spanning (7TMS) G protein-coupled receptors (Murphy, P.M., Annu. Rev. Immunol., 12:593-633 (1994)). Some known receptors for the CC or β chemokines include CCR1, which binds MIP-1 α and RANTES (Neote, K., et al., Cell, 72:415-425 (1993); Gao, J.L., J. Exp. Med., 177:1421-1427 (1993)); CCR2, which binds chemokines including MCP-1, MCP-2, MCP-3 and MCP-4 (Charo, I.F., et al., Proc. Natl. Acad. Sci. USA, 91:2752-2756 (1994); Myers, S.J., et al., J. Biol. Chem., 270:5786-5792 (1995); Gong et al., J. Biol. Chem. 272:11682-11685 (1997); Garcia-Zepeda et al., J. Immunol. 157:5613-5626 (1996)); CCR3, which binds chemokines including eotaxin, RANTES and MCP-3 (Ponath, P.D., et al., J. Exp. Med., 183:2437-2448 (1996)); CCR4, which has been found to signal in response to MCP-1, MIP-1 α , and RANTES (Power, C.A., et al., J. Biol. Chem., 270:19495-19500 (1995)); and CCR5, which has been shown to signal in response to MIP-1 α , MIP-1 β and RANTES (Boring, L., et al., J. Biol. Chem., 271 (13):7551-7558 (1996); Raport, C.J., J. Biol. Chem., 271:17161-17166 (1996); and Samson, M. et al., Biochemistry, 35:3362-3367 (1996)).

CCR2 is expressed on the surface of several leukocyte subsets, and appears to be expressed in two slightly different forms (CCR2a and CCR2b) due to alternative splicing of the mRNA encoding the carboxy-terminal region (Charo et al., Proc. Natl. Acad. Sci. USA 91:2752-2756 (1994)). MCP-1 acts upon monocytes, lymphocytes and basophils, inducing chemotaxis, granule release, respiratory burst and histamine and cytokine release. Studies have suggested that MCP-1 is implicated in the pathology of diseases such as rheumatoid arthritis, atherosclerosis, granulomatous diseases and multiple sclerosis (Koch, J. Clin. Invest. 90:772-79 (1992); Hosaka et al., Clin. Exp. Immunol. 97:451-457 (1994); Schwartz et al., Am. J. Cardiol. 71(6):9B-14B (1993); Schimmer et al., J. Immunol. 160:1466-1471 (1998); Flory et al., Lab. Invest. 69:396-404 (1993); Gong et al., J. Exp. Med. 186:131-137 (1997)). Additionally, CCR2 can act as a co-receptor for HIV (Connor et al., J. Exp. Med. 185:621-628 (1997)). Thus, CCR2 receptor antagonists may represent a new class of important therapeutic agents.

Summary of the Invention

The present invention relates to an antibody or functional fragment thereof (e.g., an antigen-binding fragment) which binds to a mammalian CC-chemokine receptor 2 (also referred to as CCR2, CKR-2, MCP-1RA or MCP-1RB) or portion of the receptor (anti-CCR2). In one embodiment, the antibody of the present invention or fragment thereof has specificity for human or rhesus CCR2 or a portion thereof. In another embodiment, the antibody or fragment of the invention blocks binding of a ligand (e.g., MCP-1, MCP-2, MCP-3, MCP-4, MCP-5, eotaxin) to the receptor and inhibits function associated with binding of the ligand to the receptor (e.g., leukocyte trafficking). For example, as described herein, antibodies and fragments thereof of the present invention which bind human or rhesus CCR2 or a portion thereof, can block binding of a chemokine (e.g., MCP-1, MCP-2, MCP-3, MCP-4, MCP-5, eotaxin) to the receptor and inhibit function associated with binding of the chemokine to the receptor. In one embodiment, the antibody is monoclonal antibody (mAb) LS132.1D9 (1D9) or an antibody which can compete with 1D9 for binding to human CCR2 or a portion of human CCR2. Functional fragments of the foregoing antibodies are also envisioned.

In another embodiment, the antibody or functional fragment of the present invention binds human CCR2 or a portion thereof, and inhibits human immunodeficiency virus (HIV) binding to the receptor, thereby inhibiting function associated with binding of HIV to the receptor (e.g., HIV antigen release and infectivity). In one embodiment, the antibody is monoclonal antibody 1D9 or an antibody which can compete with 1D9 for binding to human CCR2 or a portion of human CCR2.

The present invention also relates to an antibody or functional fragment thereof (e.g., an antigen-binding fragment) which binds to a mammalian CCR2 or portion of the receptor and provides increased fluorescent staining intensity of CCR2 or compositions comprising CCR2 relative to other anti-CCR2 antibodies. In one embodiment, the antibody is monoclonal antibody 1D9 or LS132.8G2 (8G2) or an antibody which can compete with 1D9 or 8G2 for binding to human CCR2 or a portion of human CCR2.

The present invention also relates to a humanized immunoglobulin or antigen-binding fragment thereof having binding specificity for CCR2, said immunoglobulin comprising an antigen binding region of nonhuman origin (e.g., rodent) and at least a portion

of an immunoglobulin of human origin (e.g., a human framework region, a human constant region of the gamma type). In one embodiment, the humanized immunoglobulin or fragment thereof described herein can compete with 1D9 for binding to CCR2. In a preferred embodiment, the antigen binding region of the humanized immunoglobulin is derived from monoclonal antibody 1D9 (e.g., an immunoglobulin comprising the variable regions of the light and heavy chains as shown in Figure 7 (SEQ ID NO: 9) and Figure 8 (SEQ ID NO: 10), respectively). In some embodiments, the humanized immunoglobulin or antigen binding fragment thereof can further include all or a portion of a constant region of human origin, e.g., all or a portion of a human heavy chain constant region and/or a human light chain constant region. In some embodiments, the humanized immunoglobulin or antigen binding portion thereof can include all or a portion of human constant region having one or more mutations, e.g., one or more mutations that reduce binding to Fc receptors and/or the ability to fix complement. For example, the human constant region or portion thereof can be a heavy chain constant region, e.g., a gamma heavy chain constant region, or portion thereof that includes a mutation at residues 235 and 237 of the human gamma constant region. The mutation at residue 235 can be from a leucine to an alanine. The mutation at residue 237 can be from a glycine to an alanine. In one embodiment, the humanized immunoglobulin includes all or a portion of the heavy chain constant region of SEQ ID NO:110 and/or a light chain constant region of SEQ ID NO:112.

The humanized immunoglobulin or antigen-binding fragment thereof can comprise an antigen binding region comprising at least one complementarity determining region (CDR) of nonhuman origin, and a framework region (FR) derived from a human framework region. In one aspect, the humanized immunoglobulin having binding specificity for CCR2 comprises a light chain comprising at least one CDR derived from an antibody of nonhuman origin which binds CCR2 and a FR derived from a light chain of human origin (e.g., from HF-21/28), and a heavy chain comprising a CDR derived from an antibody of nonhuman origin which binds CCR2 and a FR derived from a heavy chain of human origin (e.g., from 4B4'CL). In another aspect, the light chain comprises three CDRs derived from the light chain of the 1D9 antibody, and the heavy chain comprises three CDRs derived from the heavy chain of the 1D9 antibody. In some embodiments, the humanized immunoglobulin

further includes all or a portion of the heavy chain constant region of SEQ ID NO:110 and/or a light chain constant region of SEQ ID NO:112.

The present invention also relates to humanized immunoglobulin light chains and antigen-binding fragments thereof (e.g., comprising CDR1, CDR2 and CDR3 of the light chain of the 1D9 antibody, and a human light chain FR), and to humanized immunoglobulin heavy chains and antigen-binding fragments thereof (e.g., comprising CDR1, CDR2 and CDR3 of the heavy chain of the 1D9 antibody, and a human heavy chain FR). In a preferred embodiment, the invention relates to humanized heavy and light chains described herein (e.g., a humanized light chain comprising the variable region of the light chain shown in Figure 7 (SEQ ID NO: 9), a humanized heavy chain comprising the variable region of the heavy chain shown in Figure 8 (SEQ ID NO: 10). Also encompassed are humanized immunoglobulins comprising one or more humanized light and/or heavy chains.

The subject invention also relates to a humanized immunoglobulin or antigen-binding fragment thereof having binding specificity for CCR2 comprising a heavy chain and a light chain, wherein said light chain comprises at least one complementarity determining region derived from murine monoclonal antibody 1D9 and a framework region derived from the light chain of human antibody HF-21/28, and wherein said heavy chain comprises at least one complementarity determining region derived from murine monoclonal antibody 1D9 and a framework region derived from the heavy chain of human antibody 4B4'CL. In one embodiment, the light chain comprises three complementarity determining regions derived from the light chain of the 1D9 antibody, and the heavy chain comprises three complementarity determining regions derived from the heavy chain of the 1D9 antibody. In another embodiment, the complementarity determining regions derived from the light chain of 1D9 are amino acids 24-39 of SEQ ID NO: 9, amino acids 55-61 of SEQ ID NO: 9 and amino acids 94-102 of SEQ ID NO: 9, and the complementarity determining regions derived from the heavy chain of 1D9 are amino acids 31-35 of SEQ ID NO: 10, amino acids 50-68 of SEQ ID NO: 10 and amino acids 101-106 of SEQ ID NO: 10. In some embodiments, the humanized immunoglobulin further includes all or a portion of the heavy chain constant region of SEQ ID NO:110 and/or a light chain constant region of SEQ ID NO:112.

The invention further relates to a humanized immunoglobulin or antigen-binding fragment thereof having binding specificity for CCR2 comprising a light chain and a

complementary heavy chain, wherein said light chain comprises a variable region comprising SEQ ID NO: 12. In some embodiments, the light chain can further include all or a portion of a light chain constant region of human origin, e.g., a kappa light chain constant region of human origin. Preferably, the light chain constant region includes all or a portion of the light chain constant region of SEQ ID NO:112. The invention also relates to a humanized immunoglobulin or antigen-binding fragment thereof having binding specificity for CCR2 comprising a heavy chain and a complementary light chain, wherein said heavy chain comprises a variable region comprising SEQ ID NO: 17. In some embodiments, the heavy chain can further include all or a portion of a heavy chain constant region of human origin, e.g., a gamma heavy chain constant region of human origin. Preferably, the heavy chain constant region includes all or a portion of the heavy chain constant region of SEQ ID NO:110. In a preferred embodiment, the invention relates to a humanized immunoglobulin or antigen-binding fragment thereof having binding specificity for CCR2 comprising a heavy chain and a light chain, wherein said light chain comprises a variable region comprising SEQ ID NO: 12, and wherein said heavy chain comprises a variable region comprising SEQ ID NO: 17. In other preferred embodiments, the invention relates to a humanized immunoglobulin or antigen binding fragment thereof having a light and heavy chain, wherein the light chain includes a variable region comprising SEQ ID NO: 12 and a constant region comprising all or a portion of SEQ ID NO: 112, and wherein the heavy chain includes a variable region comprising SEQ ID NO:17 and a constant region comprising all or a portion of SEQ ID NO:110. In one embodiment, the humanized immunoglobulin or antigen-binding fragment can compete with murine antibody 1D9 for binding to CCR2. In a further embodiment, the humanized immunoglobulin or antigen-binding fragment inhibits binding of a ligand to CCR2. In some embodiments, the humanized immunoglobuli or antigen binding fragment thereof includes at least two light chains and at least two heavy chains.

The invention further relates to isolated nucleic acid molecules comprising a nucleic acid sequence which encodes a humanized immunoglobulin of the present invention (e.g., a single chain antibody), as well as to isolated nucleic acid molecules comprising a sequence which encodes a humanized immunoglobulin light chain (e.g., a sequence encoding a light chain variable region comprising SEQ ID NO: 109 or SEQ ID NO: 98 and/or a sequence encoding a light chain constant region comprising SEQ ID NO: 113, SEQ ID

NO:117, or portions thereof) or heavy chain (e.g., a sequence encoding a light chain variable region comprising SEQ ID NO: 108 or SEQ ID NO:97 and/or a sequence encoding a heavy chain constant region of SEQ ID NO: 111, SEQ ID NO:115, or portions thereof) of the present invention. For example, the present invention provides a nucleic acid (e.g., a fused nucleic acid) encoding a humanized immunoglobulin light or heavy chain comprising a first nucleic acid sequence encoding an antigen binding region derived from murine 1D9 monoclonal antibody; and a second nucleic acid sequence encoding at least a portion of a constant region of an immunoglobulin of human origin. In one embodiment, the invention provides a nucleic acid (e.g., a fused nucleic acid) encoding a humanized light chain that includes a first nucleic acid comprising SEQ ID NO:109 and a second nucleic acid comprising SEQ ID NO:113 or a portion thereof. In another embodiment, the invention provides a nucleic acid (e.g., a fused nucleic acid) encoding a humanized heavy chain that includes a first nucleic acid comprising SEQ ID NO:108 and a second nucleic acid comprising SEQ ID NO:111 or a portion thereof.

The invention also relates to isolated nucleic acid molecules comprising a nucleic acid sequence which encodes a humanized immunoglobulin of the present invention (e.g., a single chain antibody), as well as to isolated nucleic acid molecules comprising a sequence which encodes a humanized immunoglobulin light chain (e.g., a sequence encoding an amino acid sequence of SEQ ID NO:12, 13, 14, or 15 and/or a sequence encoding an amino acid of SEQ ID NO: 111, 116 or portions thereof) or heavy chain (e.g., a sequence encoding an amino acid sequence of SEQ ID NO: 17, 18, 19 or 20 and/or a sequence encoding an amino acid sequence of SEQ ID NO: 110, 114 or portions thereof).

The present invention further relates to a construct comprising a nucleic acid molecule encoding a humanized immunoglobulin having binding specificity for CCR2 or a chain of such an immunoglobulin. For example, an expression vector comprising a gene (e.g., a fused gene) encoding a humanized immunoglobulin light chain, comprising a nucleotide sequence encoding a CDR derived from a light chain of a nonhuman antibody having binding specificity for CCR2, and a framework region derived from a light chain of human origin, is provided. An expression vector comprising a gene encoding a humanized immunoglobulin heavy chain, comprising a nucleotide sequence encoding a CDR derived from a heavy chain of a nonhuman antibody having binding specificity for CCR2, and a

framework region derived from a heavy chain of human origin is another example of such a construct. In one embodiment, the expression vector can include a nucleic acid encoding a light chain that includes a first nucleic acid sequence encoding a light chain variable region, e.g., SEQ ID NO:109, and a second nucleic acid sequence encoding a light chain constant region, e.g., SEQ ID NO:113 or a portion thereof. In another embodiment, the expression vector can include a nucleic acid encoding a heavy chain that includes a first nucleic acid sequence encoding a heavy chain variable region, e.g., SEQ ID NO:108, and a second nucleic acid sequence encoding a heavy chain constant region, e.g., SEQ ID NO:111 or a portion thereof. In some embodiments, the expression vector can include a nucleic acid encoding a light chain as described herein and a nucleic acid encoding a heavy chain as described herein. In some embodiments, the expression vector will express a light chain (e.g., a light chain variable region of SEQ ID NO: 12 and/or a heavy chain constant region of SEQ ID NO:112 or a portion thereof) and/or a heavy chain (e.g., a heavy chain variable region of SEQ ID NO:17 and/or a heavy chain constant region of SEQ ID NO:110 or a portion thereof). The nucleic acid encoding one or more of the light chain variable region, the light chain constant region, the heavy chain variable region and the heavy chain constant region can further include additional sequence on one or more of the 5' or 3' ends (e.g., a restriction enzyme site and/or flanking sequence) which, when properly inserted into the vector, does not encode amino acids expressed as part of the polypeptide. For example, the nucleotide sequence of SEQ ID Nos: 98, 117, 97 and 115 include additional sequences that are not expressed as part of the polypeptide.

The present invention also relates to a host cell comprising a nucleic acid molecule of the present invention, including one or more constructs comprising a nucleic acid molecule of the present invention. In one embodiment, the invention relates to a host cell comprising a first recombinant nucleic acid encoding a humanized immunoglobulin light chain, and a second recombinant nucleic acid encoding a humanized immunoglobulin heavy chain, said first nucleic acid comprising a nucleotide sequence encoding a CDR derived from the light chain of murine 1D9 antibody and a framework region derived from a light chain of human origin; and said second nucleic acid comprising a nucleotide sequence encoding a CDR derived from the heavy chain of murine 1D9 antibody and a framework region derived from a heavy chain of human origin. In one embodiment, the host cell can include a nucleic

acid encoding a light chain that includes a first sequence encoding a light chain variable region, e.g., SEQ ID NO:109, and a second sequence encoding a light chain constant region, e.g., SEQ ID NO:113 or a portion thereof. In another embodiment, the host cell can include a nucleic acid encoding a heavy chain that includes a first sequence encoding a heavy chain variable region, e.g., SEQ ID NO:108, and a second sequence encoding a heavy chain constant region, e.g., SEQ ID NO:111 or a portion thereof. In preferred embodiments, the host cell includes a nucleic acid encoding a light chain described herein and a nucleic acid encoding a heavy chain described herein.

The present invention also provides a method of preparing a humanized immunoglobulin, or antigen binding fragment thereof, comprising maintaining a host cell of the present invention under conditions appropriate for expression of a humanized immunoglobulin, whereby a humanized immunoglobulin chain(s) is expressed and a humanized immunoglobulin or antigen binding fragment thereof is produced. The method can further comprise the step of isolating the humanized immunoglobulin.

The humanized immunoglobulins, and antigen binding fragments thereof, of the present invention can be less immunogenic than their murine or other nonhuman counterparts. Thus, the humanized immunoglobulins, and antigen binding fragments thereof, described herein can be used as therapeutic agents in humans, for example to control lymphocyte homing to mucosal lymphoid tissue, thereby, reducing inflammatory responses.

The invention further relates to a humanized immunoglobulin of the present invention for use in diagnosis or therapy (including prophylaxis). In one embodiment, the invention relates to a humanized immunoglobulin, or antigen binding fragment thereof, of the present invention for use in the treatment of diseases associated with leukocyte infiltration of tissues, for example, in the treatment of inflammatory diseases, autoimmune diseases, graft rejection, HIV infection and monocyte-mediated disorders such as atherosclerosis.

In another aspect, the invention relates to use of a humanized immunoglobulin, or antigen binding fragment thereof, of the present invention for the manufacture of a medicament for the treatment of diseases associated with leukocyte infiltration of tissues, for example, in the treatment of inflammatory diseases, autoimmune diseases, monocyte-mediated disorders such as atherosclerosis, graft rejection, or HIV infection.

The present invention further relates to a method of inhibiting the interaction of a cell bearing mammalian (e.g., human, non-human primate or murine) CCR2 with a ligand thereof, comprising contacting the cell with an effective amount of an antibody or functional fragment thereof which binds to a mammalian CCR2 or a portion of CCR2, e.g., an antibody or antigen binding fragment thereof described herein. Suitable cells include granulocytes, leukocytes, such as monocytes, macrophages, basophils and eosinophils, mast cells, and lymphocytes including T cells (e.g., CD8+ cells, CD4+ cells, CD25+ cells, CD45RO+ cells), and other cells expressing CCR2 such as a recombinant cell expressing CCR2 (e.g., transfected cells). In a particular embodiment, the antibody is 1D9 or an antibody which can compete with 1D9 for binding to human CCR2 or a portion of human CCR2.

Another embodiment of the invention relates to a method of inhibiting the interaction of a cell bearing mammalian CCR2 with a chemokine, comprising contacting said cell with an effective amount of an antibody or functional fragment thereof which binds to CCR2 or a portion of said receptor, e.g., an antibody or functional fragment thereof described herein. In one embodiment of the method, the antibody or functional fragment thereof is any one or more of 1D9, an antigen-binding fragment of 1D9 or an antibody or fragment thereof having an epitopic specificity which is the same as or similar to that of 1D9. Furthermore, the invention relates to a method of inhibiting a function associated with binding of a chemokine to CCR2, comprising administering an effective amount of an antibody or functional fragment thereof which binds to a mammalian CCR2 protein or a portion of said receptor, e.g., an antibody or functional fragment thereof described herein. In one aspect of the method, the antibody or functional fragment thereof is any one or more of 1D9, an antigen-binding fragment of 1D9 or an antibody or fragment thereof having an epitopic specificity which is the same as or similar to that of 1D9.

Another aspect of the invention is a method of identifying expression of a mammalian CCR2 or portion of the receptor by a cell. According to the method, a composition comprising a cell or fraction thereof (e.g., a membrane fraction) is contacted with an antibody or functional fragment thereof (e.g., 1D9 or 8G2) which binds to a mammalian CCR2 protein or portion of the receptor, e.g., an antibody or functional fragment thereof described herein, under conditions appropriate for binding of the antibody thereto,

and the formation of a complex between said antibody or fragment and said protein or portion thereof is detected. Detection of the complex, directly or indirectly, indicates the presence of the receptor on the cell. The present invention also relates to a kit for use in detecting the presence of CCR2 or a portion thereof in a biological sample, comprising an antibody or functional fragment thereof which binds to a mammalian CC-chemokine receptor 2 or a portion of said receptor, e.g., an antibody or functional fragment thereof described herein, and one or more ancillary reagents suitable for detecting the presence of a complex between said antibody or fragment and said protein or portion thereof.

Also encompassed by the present invention are methods of identifying additional ligands or other substances which bind a mammalian CCR2 protein, including inhibitors and/or promoters of mammalian CCR2 function. For example, agents having the same or a similar binding specificity as that of an antibody of the present invention or functional fragment thereof can be identified by a competition assay with said antibody or fragment. Thus, the present invention also encompasses methods of identifying ligands or other substances which bind the CCR2 receptor, including inhibitors (e.g., antagonists) or promoters (e.g., agonists) of receptor function. In one embodiment, cells which naturally express CCR2 receptor protein or suitable host cells which have been engineered to express a CCR2 receptor or variant encoded by a nucleic acid introduced into said cells are used in an assay to identify and assess the efficacy of ligands, inhibitors or promoters of receptor function. Such cells are also useful in assessing the function of the expressed receptor protein or polypeptide.

Thus, the invention also relates to a method of detecting or identifying an agent which binds a mammalian CCR2 or ligand binding variant thereof, comprising combining an agent to be tested, an antibody or antigen-binding fragment of the present invention (e.g., monoclonal antibody 1D9, an antibody having an epitopic specificity which is the same as or similar to that of 1D9, antigen-binding fragments of 1D9, monoclonal antibody 8G2, an antibody having an epitopic specificity which is the same as or similar to that of 8G2, and antigen-binding fragments of 8G2) and a composition comprising a mammalian CCR2 protein or a ligand binding variant thereof. The foregoing components can be combined under conditions suitable for binding of the antibody or antigen-binding fragment to mammalian CCR2 protein or a ligand binding variant thereof, and binding of the

antibody or fragment to the mammalian CCR2 protein or ligand binding variant is detected or measured, either directly or indirectly, according to methods described herein or other suitable methods. A decrease in the amount of complex formed relative to a suitable control (e.g., in the absence of the agent to be tested) is indicative that the agent binds said receptor or variant. The composition comprising a mammalian CCR2 protein or a ligand binding variant thereof can be a membrane fraction of a cell bearing recombinant CCR2 protein or ligand binding variant thereof. The antibody or fragment thereof can be labeled with a label such as a radioisotope, spin label, antigen label, enzyme label, fluorescent group and chemiluminescent group. These and similar assays can be used to detect agents, including ligands (e.g., chemokines which interact with CCR2) or other substances, including inhibitors or promoters of receptor function, which can bind CCR2 and compete with the antibodies described herein for binding to the receptor.

According to the present invention, ligands, inhibitors or promoters of receptor function can be identified in a suitable assay, and further assessed for therapeutic effect. Inhibitors of receptor function can be used to inhibit (reduce or prevent) receptor activity, and ligands and/or promoters can be used to induce (trigger or enhance) normal receptor function where indicated. The present invention also provides a method of treating inflammatory diseases, autoimmune diseases, atherosclerosis, and graft rejection, or HIV infection, comprising administering an inhibitor of receptor function (e.g., chemokine binding or HIV binding) to an individual (e.g., a mammal, such as a human). The present invention further provides a method of stimulating receptor function by administering a novel ligand or promoter to an individual, providing a new approach to selective stimulation of leukocyte function, which is useful, for example, in the treatment of infectious diseases and cancer.

Another aspect of the invention relates to a method of inhibiting HIV infection of a cell which expresses a mammalian CCR2 or portion thereof, comprising contacting the cell with an effective amount of an antibody or functional fragment thereof which binds to a mammalian CCR2 or portion of the receptor (e.g., an antibody or functional fragment thereof described herein) and inhibits HIV binding and infection. In a particular embodiment of the invention, the antibody or functional fragment thereof is any of 1D9, an antibody having an

epitopic specificity which is the same as or similar to that of 1D9, an antibody which can compete with 1D9 for binding to human CCR2, and antigen-binding fragments thereof.

Also encompassed by the present invention is a method of inhibiting (e.g., treating) HIV in a patient, comprising administering to the patient an effective amount of an antibody or functional fragment thereof which binds to a mammalian CCR2 or a portion of said receptor (e.g., an antibody or functional fragment thereof described herein) and inhibits HIV binding to the CCR2 receptor. The anti-CCR2 antibody or fragment can be administered alone or in combination with one or more additional therapeutic agents, e.g., one or more antibodies which bind a co-receptor for HIV infection and inhibit binding to said co-receptor, such as an anti-CCR3, anti-CCR5, and/or anti-CXCR4 antibody.

Another aspect of the invention also relates to a method of preventing or inhibiting HIV infection in an individual, comprising administering to the individual an effective amount of an antibody or functional fragment thereof which binds to CCR2 (e.g., an antibody or functional fragment thereof described herein) and inhibits HIV binding to CCR2. According to the method, preventing HIV infection includes treatment in order to prevent (reduce or eliminate) infection of new cells in an infected individual or in order to prevent infection in an individual who may be, may have been or has been exposed to HIV. For example, individuals such as an HIV infected individual, a fetus of an HIV infected female, or a health care worker can be treated according to the method of the present invention.

The present invention also encompasses a method of inhibiting leukocyte trafficking in a patient, comprising administering to the patient an effective amount of an antibody or functional fragment thereof which binds to a mammalian CCR2 or portion of said receptor (e.g., an antibody or functional fragment thereof described herein) and inhibits function associated with binding of a ligand to the receptor.

The present invention also relates to a method of inhibiting or treating CCR2-mediated disorders, such as inflammatory disorders, comprising administering to a patient an effective amount of an antibody or functional fragment thereof which binds to a mammalian CCR2 or portion of said receptor (e.g., an antibody or functional fragment thereof described herein) and inhibits CCR2-mediated function. For example, the invention relates to a method of inhibiting or treating stenosis or restenosis of the vasculature comprising administering to

a patient an effective amount of an antibody or functional fragment thereof which binds to a mammalian CCR2 or portion of said receptor and inhibits CCR2-mediated function.

The present invention further relates to an antibody or fragment thereof as described herein (e.g., humanized monoclonal antibody 1D9 or an antigen-binding fragment thereof) for use in therapy (including prophylaxis) or diagnosis, and to the use of such an antibody or fragment for the manufacture of a medicament for the treatment of a CCR2-mediated disorder, or other disease or inflammatory condition as described herein.

Also included in the invention are immunoglobulin light or heavy chains or fragments thereof of any antibody or immunoglobulin described herein. This can also be included for nucleic acids encoding such antibodies or immunoglobulins or fragments thereof, vectors, and cells transformed with the vectors.

Brief Description of the Figures

Figures 1A-1O are a fluorescence activated cell scanning (FACS) histogram profile illustrating that mAbs 1D9 and 8G2 stain CCR2 transfectants but not CCR5 or CCR1 transfectants. L1/2 (also referred to herein as L1.2) murine pre-B lymphoma host cells were transfected with CCR2, CCR5 and CCR1 as indicated, and stained with antibodies with different receptor specificities. Staining was analyzed by flow cytometry.

Figures 2A-2L are a FACS dot plot showing expression of CCR2 on most monocytes, a subpopulation of lymphocytes and a small subset of granulocytes. Whole blood cells were stained with one of three anti-CCR2 mAbs (5A11, generated using a peptide consisting of the first 32 amino acids of the CCR2 amino-terminus as an immunogen, and 1D9 and 8G2 generated as described herein using CCR2b L1/2 cell transfectants as the immunogen). Staining was analyzed by flow cytometry, and the lymphocyte, granulocyte and monocyte populations were gated using the forward and side light scatter. The X-axis represents forward light scatter (a measure of cell size), and the Y-axis fluorescence intensity of staining for CCR2. The level of negative control staining is indicated by a line.

Figures 3A-3I are a FACS dot plot showing that mAb 1D9 stains an IgE positive population in peripheral blood (basophils) using two-color staining for IgE and CCR2. Whole blood cells were first stained with either a negative control antibody (anti-

Flag), anti-CCR2 antibody 1D9, or an anti-CXCR1 antibody, as indicated, and detected by an anti-mouse-FITC conjugate. A second staining was done using either PBS or a biotinylated antibody specific for IgE or CD16, as indicated, and detected with a streptavidin-phycoerythrin. Staining was analyzed by flow cytometry.

Figure 4 illustrates that mAb 1D9 inhibits [¹²⁵I]MCP-1 binding to THP-1 cell membranes. 3.0 µg of THP-1 membrane protein was incubated with 0.1 nM [¹²⁵I]MCP-1 in the presence of various concentrations of 1D9 or the isotype-matched anti-CXCR3 antibody 1C6. The amount of bound tracer was determined by separation of free from bound by filtration and scintillation counting. The data was analyzed to determine the IC₅₀ value by non-linear regression using a 4-parameter logistic equation with KaleidaGraph software.

Figure 5 illustrates that mAb 1D9 inhibits [¹²⁵I]MCP-1 binding to fresh human PBMC. Freshly isolated peripheral blood mononuclear cells (500,000) were incubated with 0.1 nM [¹²⁵I]MCP-1 in the presence of various concentrations of 1D9 or the isotype-matched anti-CXCR3 antibody 1C6. The amount of bound tracer was determined by separation of free from bound by filtration and scintillation counting. The data was analyzed to determine the IC₅₀ value as for Figure 4.

Figures 6A and 6B are graphs demonstrating that mAb 1D9 inhibits MCP-1-induced chemotaxis, but not RANTES-induced chemotaxis, of fresh PBMC. Figure 6A shows the results of chemotaxis assays of PBMC to 10 nM MCP-1 with no antibody, or 0.1 or 10 µg/ml of 1D9 or nonspecific murine IgG2a. The spontaneous nonspecific migration is also indicated. Figure 6B shows the results of chemotaxis assays of PBMC to 10 nM RANTES with no antibody, 10 µg/ml 1D9 or 10 µg/ml nonspecific murine IgG2a. The spontaneous nonspecific migration in the absence of RANTES is also indicated.

Figure 7 shows the amino acid sequence (SEQ ID NO: 9) of the kappa light chain variable region of the murine 1D9 antibody. The CDRs are highlighted in bold.

Figure 8 shows the amino acid sequence (SEQ ID NO: 10) of the heavy chain variable region of the murine 1D9 antibody. The CDRs are highlighted in bold.

Figure 9 illustrates the canonical classes of CDRs in the murine 1D9 VK region. "Chothia Canonical Classes" indicates where the canonical classes as defined by Chothia and his colleagues (Chothia and Lesk, J. Mol. Biol. 197:901 (1987); Chothia et al., Nature 34:877 (1989); Tramontano et al., J. Mol. Biol. 215:175 (1990); and Chothia et al., J.

Mol. Biol. 227:799 (1992)) were used, while “Martin Canonical Classes” signifies where the canonical classes defined by Martin and Thornton (Martin and Thornton, J. Mol. Biol. 263:800 (1996)) were used. FR residues are highlighted in bold.

Figure 10 illustrates the canonical classes of CDRs in the murine 1D9 VH region. “Chothia Canonical Classes” indicates where the canonical classes as defined by Chothia and his colleagues (Chothia and Lesk, J. Mol. Biol. 197:901 (1987); Chothia et al., Nature 34:877 (1989); Tramontano et al., J. Mol. Biol. 215:175 (1990); and Chothia et al., J. Mol. Biol. 227:799 (1992)) were used, while “Martin Canonical Classes” signifies where the canonical classes defined by Martin and Thornton (Martin and Thornton, J. Mol. Biol. 263:800 (1996)) were used. FR residues are highlighted in bold.

Figure 11 shows the amino acid sequences of various versions of the humanized 1D9 VK region (SEQ ID NOS: 12-15 and 107, respectively). Where the 1D9 VK region residues (SEQ ID NO: 9) and the human HF-21/28 VK region (SEQ ID NO: 11) sequences match a dot [.] is shown. Where no amino acid is present at a specific residue position a dash [-] is shown. Where an amino acid in the HF-21/28 FRs is changed in the humanized 1D9 VK region, it is highlighted in bold. The CDRs are described by the use of nomenclature [=L1=]. The numbering used is according to Kabat et al., Sequences of proteins of immunological interest, Fifth edition, U.S. Department of Health and Human Services, U.S. Government Printing Office (1991).

Figure 12 shows the amino acid sequences of various versions of the humanized 1D9 VH region (SEQ ID NOS: 17-20, respectively). Where the 1D9 VH region residues (SEQ ID NO: 10) and the human 4B4'CL VH region sequences (SEQ ID NO: 16) match a dot [.] is shown. Where no amino acid is present at a specific residue position a dash [-] is shown. Where an amino acid in the 4B4'CL is changed in the humanized 1D9 VH region, it is highlighted in bold. The CDRs are described by the use of nomenclature [=H1=], while [-----] denotes part of the H1 structure loop. The numbering used is according to Kabat et al., Sequences of proteins of immunological interest, Fifth edition, U.S. Department of Health and Human Services, U.S. Government Printing Office (1991).

Figure 13 shows a comparison of a portion of the murine 1D9 VK region (SEQ ID NO: 21) with mouse germline VK gene sequences (SEQ ID NOS: 22-33, respectively). “Identical residues” represents the number of identical residues in a mouse

germline VK region to the murine 1D9 VK region. Where the 1D9 VK region sequence and the mouse germline VK region sequences match a dot [.] is shown. Where no amino acid is present at a specific residue position a dash [-] is shown.

Figure 14 shows a comparison of a portion of the murine 1D9 VH region (SEQ ID NO: 34) with mouse germline VH gene sequences (SEQ ID NOS: 35-53, respectively). "Identical residues" represents the number of identical residues in a mouse germline VH region to the murine 1D9 VH region. Where the 1D9 VH region sequence and the mouse germline VH region sequences match a dot [.] is shown. Where no amino acid is present at a specific residue position a dash [-] is shown.

Figure 15 shows a comparison of the murine 1D9 VK region (SEQ ID NO: 9) with the most homologous seventeen human VK amino acid sequences (SEQ ID NOS: 54-70, respectively). "ID" represents the percentage identity of the human VK sequences to the murine 1D9 VK region. Where the 1D9 VK region residues and the human VK region sequences match a dot [.] is shown. Where no amino acid is present at a specific residue position a dash [-] is shown. "S" indicates amino acid positions on the surface of the FV domain. "C" indicates residues located within the core of the FV domain. Residues within 5Å of a CDR are defined using capital letters, while those located further away are described with a lower case letter. The CDRs themselves are described by the use of the nomenclature == L1 ==. "v" denotes the Vernier residues (Foote and Winter, J. Mol. Biol. 224:487 (1992)) located in the FRs. Those residues in the human VK region sequences which are underlined differ from their closest human VK germline gene. The numbering used is as according to Kabat et al., Sequences of proteins of immunological interest, Fifth edition, U.S. Department of Health and Human Services, U.S. Government Printing Office (1991).

Figure 16 shows a comparison of the murine 1D9 VK region with the most homologous seventeen human VK amino acid sequences. "ID" indicates the percentage identity of the human VK sequence to the murine 1D9 VK region. "Surface" indicates the number of identical residues on the surface. "Core" indicates the number of identical residues within the core of the FV domain. "CDR" indicates the number of identical residues within the CDRs. "FR" indicates the number of identical residues within the FRs. "FR Surface" indicates the number of identical residues which are surface exposed. "FR Core" indicates the number of identical residues which are located within the core of the FV

domain. "FR Near CDR" represents the number of identical residues amongst the FR amino acids within 5Å of a CDR. "Vernier" indicates the number of identical residues amongst the 14 Vernier amino acids (Foote and Winter, J. Mol. Biol. 224:487 (1992)). "VK" indicates the number of identical residues within the VK gene. "J Chain" indicates the number of identical residues within the J chain gene. "L1 Len" to "L3 Len" defines the number of residues in each CDR, while "L1 Class" to "L3 Class" describes the canonical class of the CDR according to Martin and Thornton (Martin and Thornton, J. Mol. Biol. 263:800 (1996)).

Figures 17A-17B show a comparison of the murine 1D9 VH region (SEQ ID NO: 10) with the most homologous 24 human VH amino acid sequences (SEQ ID NOS: 71-94, respectively). "ID" represents the percentage identity of the human VH sequences to the murine 1D9 VH region. Where the 1D9 VH region residues and the human VH region sequences match a dot [.] is shown. Where no amino acid is present at a specific residue position a dash [-] is shown. "S" indicates amino acid positions on the surface of the FV domain. "C" indicates residues located within the core of the FV domain. Residues within 5Å of a CDR are defined using capital letters, while those located farther away are described with a lower case letter. The CDRs themselves are described by the use of the nomenclature == H1= . "v" denotes the Vernier residues (Foote and Winter, J. Mol. Biol. 224:487 (1992)) located in the FRs. Those residues in the human VH region sequences which are underlined differ from their closest human VH germline gene. The numbering used is as according to Kabat et al., Sequences of proteins of immunological interest, Fifth edition, U.S. Department of Health and Human Services, U.S. Government Printing Office (1991).

Figures 18A-18B show a comparison of the murine 1D9 VH region with the most homologous 24 human VH amino acid sequences. "ID" indicates percentage identity of the human VH sequence to the murine 1D9 VH region. "All" indicates the number of identical residues in the whole of the human VH region when compared to the whole of the murine 1D9 VH region. "Surface" indicates the number of identical residues on the surface. "Core" indicates the number of identical residues within the core of the FV domain. "CDR" indicates the number of identical residues within the CDRs. "FR" indicates the number of identical residues within the FRs. "FR Surface" indicates the number of identical residues which are surface exposed. "FR Core" indicates the number of identical residues which are located within the core of the FV domain. "FR Near CDR" represents the number of

identical residues amongst the FR amino acids within 5Å of a CDR. “Vernier” indicates the number of identical residues amongst the 14 Vernier amino acids (Foote and Winter, J. Mol. Biol. 224:487 (1992)). “VH” indicates the number of identical residues within the VH gene. “J Chain” indicates the number of identical residues within the J chain gene. “H1 Size” to
 5 “H3 Size” define the number of residues in each CDR, while “H1 Class” and “H2 Class” describe the canonical class of the CDR according to Martin and Thornton, (J. Mol. Biol. 263:800 (1996)).

Figures 19A-19C show the alignment of amino acid sequences leading to the design of the first (1D9RKA) and second (1D9RKB) humanized versions of the 1D9
 10 antibody kappa light chain variable region. Amino acids identical to the mouse 1D9 at a particular residue position in column 7 are not shown; “-” indicates no amino acid is located at this residue position. Boldface type indicates positions in FRs and CDRs where the human amino acid residue was replaced by the corresponding mouse residue. “Δ” indicates the numbering of changes in the human FRs of 1D9RKA. “Mouse 1D9 VK” indicates the amino
 15 acid sequence of the VK region from the murine 1D9 kappa light chain variable region. “Mouse 6-II” indicates the consensus sequence of mouse VK regions from Kabat subgroup 6-II. “Human 6-II” indicates the consensus sequence of human VK regions from Kabat subgroup 6-II. “HF-21/28” indicates the amino acid sequence of the light chain variable region from the human HF-21/28 antibody (Chastagner et al., Gene 101(2):305-6 (1991)).
 20 The number in parenthesis (005056) is the Kabat database ID number. “Surface or Core” indicates the position of the amino acid in relation to the rest of the residues in both chains of the antibody variable regions. Residues within 5Å of a CDR are defined using capital letters. “1D9RKA” indicates the amino acid sequence of the first version of the humanized 1D9 VK region. “1D9RKB” indicates the amino acid sequence of the second version of the
 25 humanized 1D9 VK region.

Figures 20A-20C show the alignment of amino acid sequences leading to the design of the first (1D9RHA) and second (1D9RKB) humanized human versions of the 1D9
 antibody kappa light chain variable region. Amino acids identical to the mouse 1D9 at a particular residue position in column 7 are not shown. “-” indicates that no amino acid is
 30 located at this residue position. Boldface type indicates positions in the FRs and CDRs where the human amino acid residue was replaced by the corresponding mouse residue. “Δ”

indicates the numbering of changes in the human FRs of 1D9RHA. “Mouse 1D9 VH” indicates the amino acid sequence of the VH region from the murine 1D9 heavy chain variable region. “Mouse IIIc” indicates the consensus sequence of mouse VH regions from Kabat subgroup IIIc. “Human III” indicates the consensus sequence of human VH regions from Kabat subgroup III. “4B4'CL” indicates the amino acid sequence of the heavy chain variable region from the human 4B4'CL antibody (Sanz et al., Journal of Immunology 142:883 (1989)). The number in parenthesis (000490) is the Kabat database ID number. “Surface or Core” indicates the position of the amino acid in relation to the rest of the residues in both chains of the antibody variable regions. Residues within 5Å of a CDR are defined using capital letters. “1D9RHA” indicates the amino acid sequence of the first version of the humanized 1D9 VH region. “1D9RHB” indicates the amino acid sequence of the second version of the humanized 1D9 VH region.

Figure 21 shows the nucleotide sequence, complement and encoded amino acid sequence of the murine antibody 1D9 heavy chain variable region. The leader sequence and a portion of the constant region are also shown. The illustrated nucleotide sequence is SEQ ID NO: 96, the complementary sequence is SEQ ID NO: 99, and the amino acid sequence is SEQ ID NO: 100.

Figure 22 shows the nucleotide sequence, complement and encoded amino acid sequence of the murine antibody 1D9 kappa light chain variable region. The leader sequence and a portion of the constant region are also shown. The illustrated nucleotide sequence is SEQ ID NO: 95, the complementary sequence is SEQ ID NO: 101, and the amino acid sequence is SEQ ID NO: 102.

Figure 23 shows the nucleotide sequence of the humanized heavy chain 1D9RHA. The indicated enzyme sites were added for cloning into the vector pLKTOK41. The vector also has human leader and constant regions. The illustrated nucleotide sequence is SEQ ID NO: 97, the complementary sequence is SEQ ID NO: 103, and the amino acid sequence is SEQ ID NO: 104.

Figure 24 shows the nucleotide sequence of the humanized light chain 1D9RKA. The indicated enzyme sites were added for cloning into the vector pLKTOK41. The vector also has human leader and constant regions. The bracketed Y indicates a residue which changes to aspartate when cloned into the Eco RV cloning site of pLKTOK41. The

illustrated nucleotide sequence is SEQ ID NO: 98, the complementary sequence is SEQ ID NO: 105, and the amino acid sequence is SEQ ID NO: 106.

Figure 25 illustrates a comparison of the abilities of murine mAb 1D9 and a humanized version of mAb 1D9 (heavy chain of 1D9RHAVH, light chain of 1D9RKAVK) to inhibit the binding of [125I]-MCP-1 to whole THP-1 cells. Data points for murine 1D9 (mus-1D9) are shown as closed triangles. Data points for the humanized version of 1D9 (hum-1D9) are shown as closed squares.

Figure 26 depicts construction process for transfer of heavy chain cassettes and light chain cassettes into a single combination immunoglobulin DNA cassette vector. For creation of combined vectors, the heavy chain cassette, including promoter, nVHL, IgG constant region and polyadenylation region, can be cloned as a Bgl II/Bam HI fragment into the Bam HI site of the vector with the light chain cassette. Bgl II and Bam HI have cohesive ends and both sites are lost upon ligation.

Figure 27 depicts the structure of the complete combination heavy and light chain immunoglobulin DNA cassette antibody expression vector.

Figure 28 depicts the cloning process for incorporation of desired variable sequences (VH and VK) into the combination heavy and light chain immunoglobulin DNA cassette antibody expression vector.

Detailed Description of the Invention

Embodiments of the present invention relates to an antibody (anti-CCR2) or functional fragment thereof which binds mammalian CC-chemokine receptor 2 (CCR2, CKR-2, MCP-1RA or MCP-1RB) or a portion of CCR2. In one embodiment, the antibody has specificity for human or rhesus CCR2 or portion thereof. In one embodiment, the antibodies (immunoglobulins) are raised against an isolated and/or recombinant mammalian CCR2 or portion thereof (e.g., peptide) or against a host cell which expresses mammalian CCR2. In a preferred embodiment, the antibodies specifically bind human CCR2 receptor(s) (e.g., CCR2a and/or CCR2b) or a portion thereof, and in a particularly preferred embodiment the antibodies have specificity for a naturally occurring or endogenous human CCR2. As used herein, "CC-chemokine receptor 2" ("CCR2") refers to CC-chemokine receptor 2a and/or CC-chemokine receptor 2b. Antibodies or functional fragments thereof which can

inhibit one or more functions characteristic of a mammalian CCR2, such as a binding activity (e.g., ligand, inhibitor and/or promoter binding), a signaling activity (e.g., activation of a mammalian G protein, induction of a rapid and transient increase in the concentration of cytosolic free calcium $[Ca^{2+}]_i$), and/or stimulation of a cellular response (e.g., stimulation of chemotaxis, exocytosis or inflammatory mediator release by leukocytes, integrin activation) are also encompassed by the present invention, such as an antibody which can inhibit binding of a ligand (i.e., one or more ligands) to CCR2 and/or one or more functions mediated by CCR2 in response to a ligand. For example, in one aspect, the antibodies or functional fragments thereof can inhibit (reduce or prevent) the interaction of receptor with a natural ligand, such as MCP-1, MCP-2, MCP-3, MCP-4, MCP-5 and/or eotaxin. In another aspect, an antibody or functional fragment thereof that binds to CCR2 can inhibit binding of MCP-1, MCP-2, MCP-3, MCP-4, MCP-5 and/or eotaxin and/or HIV to mammalian CCR2 (e.g., human CCR2, non-human primate CCR2, murine CCR2). The antibodies or functional fragments thereof of the present invention can inhibit functions mediated by human CCR2, including leukocyte trafficking, HIV entry into a cell, T cell activation, inflammatory mediator release and/or leukocyte degranulation. Preferably, the antibodies or fragments can bind CCR2 with an affinity of at least about 0.1×10^{-8} M, preferably at least about 1×10^{-9} M, and more preferably at least about 3×10^{-9} M. In a particular embodiment, antibodies or functional fragments thereof demonstrate inhibition of chemokine-induced (e.g., MCP-1-induced) chemotaxis of cells (e.g., PBMC) at less than about 150 $\mu\text{g/ml}$, preferably less than about 100 $\mu\text{g/ml}$, more preferably less than about 50 $\mu\text{g/ml}$, and even more preferably less than about 20 $\mu\text{g/ml}$.

In a further embodiment of the invention, the antibodies or functional fragments thereof of the invention can inhibit binding of a CCR2 ligand (e.g., a chemokine) to CCR2 with an IC_{50} of less than about 1.0 $\mu\text{g/ml}$, preferably less than about 0.05 $\mu\text{g/ml}$, and more preferably less than about 0.005 $\mu\text{g/ml}$.

Murine monoclonal antibodies specific for CCR2, designated 1D9 and 8G2, were produced as described herein. In a preferred embodiment, the antibodies of the present invention bind human CCR2, and have an epitopic specificity which is the same as or similar to that of murine 1D9 or 8G2 antibody described herein. Antibodies with an epitopic specificity which is the same as or similar to that of murine 1D9 monoclonal antibody can be

identified by their ability to compete with murine 1D9 monoclonal antibody for binding to human CCR2 (e.g., to cells bearing human CCR2, such as transfectants bearing CCR2, CD8+ cells, CD4+ cells, CDR45RO+ cells, CD25+ cells, monocytes, dendritic cells, macrophages and basophils). Similarly, antibodies with an epitopic specificity which is the same as or similar to that of murine 8G2 monoclonal antibody can be identified by their ability to compete with murine 8G2 monoclonal antibody for binding to human CCR2. Using receptor chimeras (Rucker et al., Cell 87:437-446 (1996)), the binding site of mAbs 1D9 and 8G2 has been mapped to the amino-terminal domain of human CC-chemokine receptor 2, specifically to an epitope comprising from about amino acid 1 to about amino acid 30 of the protein. Using these or other suitable techniques, antibodies having an epitopic specificity which is the same as or similar to that of an antibody of the present invention can be identified. mAbs 1D9 and 8G2 have epitopic specificity for the amino-terminal domain of the CCR2 receptor, e.g., from about amino acid number 1 to about amino acid number 30 of the receptor protein. Thus, the invention pertains to an antibody or functional portion thereof which binds to the amino-terminal domain or portion thereof of mammalian CC-chemokine receptor 2, and particularly to an epitope comprising from about amino acid 1 to about amino acid 30 of mammalian CC-chemokine receptor 2.

The invention also relates to a bispecific antibody, or functional fragment thereof (e.g., F(ab')₂), which has the same or similar epitopic specificity as at least two of the antibodies described herein (see, e.g., U.S. Patent No. 5,141,736 (Iwasa et al.), U.S. Patent Nos. 4,444,878, 5,292,668, 5,523,210 (all to Paulus et al.) and U.S. Patent No. 5,496,549 (Yamazaki et al.). For example, a bispecific antibody of the present invention can have the same or similar epitopic specificity as mAb 1D9 and 8G2, e.g., binds the amino terminal domain, or portion thereof, of mammalian CCR2 protein.

Hybridoma cell lines producing antibodies according to the present invention were deposited on July 17, 1998, on behalf of LeukoSite, Inc., 215 First Street, Cambridge, MA 02142, U.S.A. (now Millennium Pharmaceuticals, Inc., 75 Sidney Street, Cambridge, MA 02139, U.S.A.), at the American Type Culture Collection, 10801 University Boulevard, Manassas, Virginia 20110, U.S.A., under Accession Nos. HB-12549 (1D9) and HB-12550 (8G2). The present invention also pertains to the hybridoma cell lines deposited under ATCC Accession No. HB-12549 and ATCC Accession No. HB-12550, as well as to the

monoclonal antibodies produced by the hybridoma cell lines deposited under ATCC Accession Nos. HB-12549 and HB-12550.

The antibodies of the present invention can be polyclonal or monoclonal, and the term "antibody" is intended to encompass both polyclonal and monoclonal antibodies.

Furthermore, it is understood that methods described herein which utilize 8G2 can also utilize functional fragments (e.g., antigen-binding fragments) of 8G2, antibodies which have the same or similar epitopic specificity as 8G2, and combinations thereof, optionally in combination with antibodies or fragments having an epitopic specificity which is not the same as or similar to 8G2; similarly, methods described as utilizing 1D9 can also utilize functional fragments of 1D9, antibodies which have the same or similar epitopic specificity as 1D9, and combinations thereof, optionally in combination with antibodies or fragments having an epitopic specificity which is not the same as or similar to 1D9. Antibodies of the present invention can be raised against an appropriate immunogen, such as isolated and/or recombinant mammalian CCR2 protein or portion thereof, or synthetic molecules, such as synthetic peptides. In a preferred embodiment, cells which express receptor, such as transfected cells, can be used as immunogens or in a screen for antibody which binds receptor.

The antibodies of the present invention, and fragments thereof, are useful in therapeutic, diagnostic and research applications as described herein. The present invention encompasses an antibody or functional portion thereof of the present invention (e.g., mAb 1D9 or 8G2, or antigen-binding fragments thereof or other CCR-2 binding antibodies or fragments thereof as described herein) for use in therapy (including prophylaxis) or diagnosis (e.g., of particular diseases or conditions as described herein), and use of such antibodies or functional portions thereof for the manufacture of a medicament for use in treatment of diseases or conditions as described herein.

Preparation of immunizing antigen, and polyclonal and monoclonal antibody production can be performed as described herein, or using other suitable techniques. A variety of methods have been described (see e.g., Kohler et al., *Nature*, 256: 495-497 (1975) and *Eur. J. Immunol.* 6: 511-519 (1976); Milstein et al., *Nature* 266: 550-552 (1977); Koprowski et al., U.S. Patent No. 4,172,124; Harlow, E. and D. Lane, 1988, *Antibodies: A Laboratory Manual*, (Cold Spring Harbor Laboratory: Cold Spring Harbor, NY); Current

Protocols In Molecular Biology, Vol. 2 (Supplement 27, Summer '94), Ausubel, F.M. et al., Eds., (John Wiley & Sons: New York, NY), Chapter 11, (1991)). Generally, a hybridoma can be produced by fusing a suitable immortal cell line (e.g., a myeloma cell line such as SP2/0) with antibody producing cells. The antibody producing cell, preferably those of the spleen or lymph nodes, are obtained from animals immunized with the antigen of interest. The fused cells (hybridomas) can be isolated using selective culture conditions, and cloned by limiting dilution. Cells which produce antibodies with the desired binding properties can be selected by a suitable assay (e.g., ELISA).

Other suitable methods of producing or isolating antibodies which bind CCR2, including human or artificial antibodies, can be used, including, for example, methods which select recombinant antibody (e.g., single chain Fv or Fab) from a library, or which rely upon immunization of transgenic animals (e.g., mice) capable of producing a repertoire of human antibodies (see e.g., Jakobovits et al., Proc. Natl. Acad. Sci. USA, 90: 2551-2555 (1993); Jakobovits et al., Nature, 362: 255-258 (1993); Lonberg et al., U.S. Patent No. 5,545,806; Surani et al., U.S. Patent No. 5,545,807).

Single chain antibodies, and chimeric, humanized or primatized (CDR-grafted) antibodies, as well as chimeric or CDR-grafted single chain antibodies, and the like, comprising portions derived from different species, are also encompassed by the present invention and the term "antibody". The various portions of these antibodies can be joined together chemically by conventional techniques, or can be prepared as a contiguous protein using genetic engineering techniques. For example, nucleic acids encoding a chimeric or humanized chain can be expressed to produce a contiguous protein. See, e.g., Cabilly et al., U.S. Patent No. 4,816,567; Cabilly et al., European Patent No. 0,125,023 B1; Boss et al., U.S. Patent No. 4,816,397; Boss et al., European Patent No. 0,120,694 B1; Neuberger, M.S. et al., WO 86/01533; Neuberger, M.S. et al., European Patent No. 0,194,276 B1; Winter, U.S. Patent No. 5,225,539; Winter, European Patent No. 0,239,400 B1; and Queen et al., U.S. Patent Nos. 5,585,089, 5,698,761 and 5,698,762. See also, Newman, R. et al., BioTechnology, 10: 1455-1460 (1992), regarding primatized antibody, and Ladner et al., U.S. Patent No. 4,946,778 and Bird, R.E. et al., Science, 242: 423-426 (1988)) regarding single chain antibodies.

In addition, functional fragments of antibodies, including fragments of chimeric, humanized, primatized or single chain antibodies, can also be produced.

Functional fragments of the foregoing antibodies retain at least one binding function and/or modulation function of the full-length antibody from which they are derived. Preferred functional fragments retain an antigen-binding function of a corresponding full-length antibody (e.g., the ability to bind a mammalian CCR2). Particularly preferred functional fragments retain the ability to inhibit one or more functions characteristic of a mammalian CCR2, such as a binding activity, a signaling activity, and/or stimulation of a cellular response. For example, in one embodiment, a functional fragment can inhibit the interaction of CCR2 with one or more of its ligands (e.g., MCP-1, MCP-2, MCP-3, MCP-4, MCP-5 and/or eotaxin) and/or can inhibit one or more receptor-mediated functions, such as leukocyte trafficking, HIV entry into cells, T cell activation, inflammatory mediator release and/or leukocyte degranulation.

For example, antibody fragments capable of binding to a mammalian CCR2 receptor or portion thereof, including, but not limited to, Fv, Fab, Fab' and F(ab')₂ fragments are encompassed by the invention. Such fragments can be produced by enzymatic cleavage or by recombinant techniques, for example. For instance, papain or pepsin cleavage can generate Fab or F(ab')₂ fragments, respectively. Antibodies can also be produced in a variety of truncated forms using antibody genes in which one or more stop codons has been introduced upstream of the natural stop site. For example, a chimeric gene encoding a F(ab')₂ heavy chain portion can be designed to include DNA sequences encoding the CH1 domain and hinge region of the heavy chain.

The present invention relates to a humanized immunoglobulin or antigen-binding fragment thereof having binding specificity for CCR2, comprising an antigen binding region of nonhuman origin (e.g., rodent) and at least a portion of an immunoglobulin of human origin (e.g., a human framework region, a human constant region or portion thereof). In one embodiment, the humanized immunoglobulin includes an antigen binding region of nonhuman origin which binds CCR2 and a constant region derived from a human constant region. In another embodiment, the humanized immunoglobulin which binds CCR2 comprises a complementarity determining region of nonhuman origin and a variable framework region of human origin, and optionally, a constant region of human origin.

Preferably, the humanized immunoglobulin that binds CCR2 includes a complementarity determining region of nonhuman origin, a variable framework region of human origin and at least a portion of a constant region of human origin, e.g., at least a portion of a gamma constant region of human origin and/or at least a portion of a kappa constant region of human origin. The constant region can include one or more mutations described herein. In some embodiments, the humanized immunoglobulin can comprise a heavy chain and a light chain, wherein the light chain comprises a complementarity determining region derived from an antibody of nonhuman origin which binds CCR2 and a framework region derived from a light chain of human origin, and the heavy chain comprises a complementarity determining region derived from an antibody of nonhuman origin which binds CCR2 and a framework region derived from a heavy chain of human origin.

In one embodiment, the humanized immunoglobulin can compete with murine 1D9 or 8G2 monoclonal antibody for binding to human CCR2. In a preferred embodiment, the antigen-binding region of the humanized immunoglobulin (a) is derived from 1D9 monoclonal antibody (e.g., as in a humanized immunoglobulin comprising CDR1, CDR2 and CDR3 of the 1D9 light chain and/or CDR1, CDR2 and CDR3 of the 1D9 heavy chain) or (b) is derived from 8G2 monoclonal antibody (e.g., as in a humanized immunoglobulin comprising CDR1, CDR2 and CDR3 of the 8G2 light chain and/or CDR1, CDR2 and CDR3 of the 8G2 heavy chain). Chimeric or CDR-grafted single chain antibodies are also encompassed by the term humanized immunoglobulin.

The present invention also relates to a humanized immunoglobulin light chain or antigen-binding fragment thereof or a humanized immunoglobulin heavy chain or antigen-binding fragment thereof. In one embodiment, the invention relates to a humanized light chain comprising a light chain CDR (i.e., one or more CDRs) of nonhuman origin and a human light chain framework region. In some embodiments, the humanized immunoglobulin light chain can further include at least a portion of a constant region of human origin, e.g., a kappa light chain constant region of human origin. In another embodiment, the present invention relates to a humanized immunoglobulin heavy chain comprising a heavy chain CDR (i.e., one or more CDRs) of nonhuman origin and a human heavy chain framework region. In some embodiments, the humanized immunoglobulin heavy chain can further include at least a portion of a constant region of human origin, e.g., a

gamma heavy chain constant region of human origin. The constant region can include one or more mutations described herein. The CDRs can be derived from a nonhuman immunoglobulin.

Naturally occurring immunoglobulins have a common core structure in which two identical light chains (about 24 kD) and two identical heavy chains (about 55 or 70 kD) form a tetramer. The amino-terminal portion of each chain is known as the variable (V) region and can be distinguished from the more conserved constant (C) regions of the remainder of each chain. Within the variable region of the light chain is a C-terminal portion known as the J region. Within the variable region of the heavy chain, there is a D region in addition to the J region. Most of the amino acid sequence variation in immunoglobulins is confined to three separate locations in the V regions known as hypervariable regions or complementarity determining regions (CDRs) which are directly involved in antigen binding. Proceeding from the amino-terminus, these regions are designated CDR1, CDR2 and CDR3, respectively. The CDRs are held in place by more conserved framework regions (FRs). Proceeding from the amino-terminus, these regions are designated FR1, FR2, FR3, and FR4, respectively. The locations of CDR and FR regions and a numbering system have been defined by Kabat et al. (Kabat et al., Sequences of Proteins of Immunological Interest, Fifth Edition, U.S. Department of Health and Human Services, U.S. Government Printing Office (1991)).

Human immunoglobulins can be divided into classes and subclasses, depending on the isotype of the heavy chain. The classes include IgG, IgM, IgA, IgD and IgE, in which the heavy chains are of the gamma (γ), mu (μ), alpha (α), delta (δ) or epsilon (ϵ) type, respectively. Subclasses include IgG1, IgG2, IgG3, IgG4, IgA1 and IgA2, in which the heavy chains are of the γ 1, γ 2, γ 3, γ 4, α 1 and α 2 type, respectively. Human immunoglobulin molecules of a selected class or subclass may contain either a kappa (κ) or lambda (λ) light chain. See e.g., Cellular and Molecular Immunology, Wonsiewicz, M.J., Ed., Chapter 45, pp. 41-50, W. B. Saunders Co, Philadelphia, PA (1991); Nisonoff, A., Introduction to Molecular Immunology, 2nd Ed., Chapter 4, pp. 45-65, Sinauer Associates, Inc., Sunderland, MA (1984).

The term "immunoglobulin" as used herein includes whole antibodies and biologically functional fragments thereof. Such biologically functional fragments retain at

least one antigen-binding function of a corresponding full-length antibody (e.g., specificity for CCR2 of antibody 1D9), and preferably, retain the ability to inhibit the interaction of CCR2 with one or more of its ligands (e.g., HIV, MCP-1, MCP-2, MCP-3, MCP-4, MCP-5, eotaxin). Examples of biologically functional antibody fragments which can be used include fragments capable of binding to CCR2, such as single chain antibodies, Fv, Fab, Fab' and F(ab')₂ fragments. Such fragments can be produced by enzymatic cleavage or by recombinant techniques. For instance, papain or pepsin cleavage can be used to generate Fab or F(ab')₂ fragments, respectively. Antibodies can also be produced in a variety of truncated forms using antibody genes in which one or more stop codons have been introduced upstream of the natural stop site. For example, a chimeric gene encoding the heavy chain of an F(ab')₂ fragment can be designed to include DNA sequences encoding the CH1 domain and hinge region of the heavy chain. As used herein, an antigen-binding fragment of a humanized immunoglobulin heavy or light chain is intended to mean a fragment which binds to an antigen when paired with a complementary chain. That is, an antigen-binding fragment of a humanized light chain will bind to an antigen when paired with a heavy chain (e.g., murine, chimeric, humanized) comprising a variable region, and an antigen-binding fragment of a humanized heavy chain will bind to an antigen when paired with a light chain (e.g., murine, chimeric, humanized) comprising a variable region.

The term "humanized immunoglobulin" as used herein refers to an immunoglobulin comprising portions of immunoglobulins of different origin, wherein at least one portion is of human origin. For example, the humanized antibody can comprise portions derived from an immunoglobulin of nonhuman origin with the requisite specificity, such as a mouse, and from immunoglobulin sequences of human origin (e.g., chimeric immunoglobulin), joined together chemically by conventional techniques (e.g., synthetic) or prepared as a contiguous polypeptide using genetic engineering techniques (e.g., DNA encoding the protein portions of the chimeric antibody can be expressed to produce a contiguous polypeptide chain). Another example of a humanized immunoglobulin of the present invention is an immunoglobulin containing one or more immunoglobulin chains comprising a CDR derived from an antibody of nonhuman origin and a framework region derived from a light and/or heavy chain of human origin (e.g., CDR-grafted antibodies with or without framework changes). Chimeric or CDR-grafted single chain antibodies are also

encompassed by the term humanized immunoglobulin. See, e.g., Cabilly et al., U.S. Patent No. 4,816,567; Cabilly et al., European Patent No. 0,125,023 B1; Boss et al., U.S. Patent No. 4,816,397; Boss et al., European Patent No. 0,120,694 B1; Neuberger, M.S. et al., WO 86/01533; Neuberger, M.S. et al., European Patent No. 0,194,276 B1; Winter, U.S. Patent
 5 No. 5,225,539; Winter, European Patent No. 0,239,400 B1; Padlan, E.A. et al., European Patent Application No. 0,519,596 A1. See also, Ladner et al., U.S. Patent No. 4,946,778; Huston, U.S. Patent No. 5,476,786; and Bird, R.E. et al., *Science*, 242: 423-426 (1988)), regarding single chain antibodies.

For example, humanized immunoglobulins can be produced using synthetic
 10 and/or recombinant nucleic acids to prepare genes (e.g., cDNA) encoding the desired humanized chain. For example, nucleic acid (e.g., DNA) sequences coding for humanized variable regions can be constructed using PCR mutagenesis methods to alter DNA sequences encoding a human or humanized chain, such as a DNA template from a previously humanized variable region (see e.g., Kamman, M., et al., *Nucl. Acids Res.*, 17: 5404 (1989));
 15 Sato, K., et al., *Cancer Research*, 53: 851-856 (1993); Daugherty, B.L. et al., *Nucleic Acids Res.*, 19(9): 2471-2476 (1991); and Lewis, A.P. and J.S. Crowe, *Gene*, 101: 297-302 (1991)). Using these or other suitable methods, variants can also be readily produced. In one embodiment, cloned variable regions can be mutagenized, and sequences encoding variants with the desired specificity can be selected (e.g., from a phage library; see e.g., Krebber et
 20 al., U.S. 5,514,548; Hoogenboom et al., WO 93/06213, published April 1, 1993)).

The antigen binding region of the humanized immunoglobulin (the nonhuman portion) can be derived from an immunoglobulin of nonhuman origin (referred to as a donor immunoglobulin) having binding specificity for CCR2. For example, a suitable antigen binding region can be derived from the murine monoclonal antibody 1D9. Other sources
 25 include CCR2-specific antibodies obtained from nonhuman sources, such as rodent (e.g., mouse, rat), rabbit, pig goat or non-human primate (e.g., monkey). Additionally, other polyclonal or monoclonal antibodies, such as antibodies which bind to the same or similar epitope as the 1D9 antibody, can be made (e.g., Kohler et al., *Nature*, 256:495-497 (1975); Harlow et al., 1988, *Antibodies: A Laboratory Manual*, (Cold Spring Harbor, NY); and
 30 *Current Protocols in Molecular Biology*, Vol. 2 (Supplement 27, Summer '94), Ausubel et al., Eds. (John Wiley & Sons: New York, NY), Chapter 11 (1991)).

For example, antibodies can be raised against an appropriate immunogen in a suitable mammal (e.g., a mouse, rat, rabbit or sheep). Cells bearing CCR2, membrane fractions containing CCR2, and immunogenic fragments of CCR2 are examples of suitable immunogens. Antibody-producing cells (e.g., a lymphocyte) can be isolated from, for example, the lymph nodes or spleen of an immunized animal. The cells can then be fused to a suitable immortalized cell (e.g., a myeloma cell line), thereby forming a hybridoma. Fused cells can be isolated employing selective culturing techniques. Cells which produce antibodies with the desired specificity can be selected by a suitable assay (e.g., ELISA). Immunoglobulins of nonhuman origin having binding specificity for CCR2 can also be obtained from antibody libraries (e.g., a phage library comprising nonhuman Fab molecules).

In one embodiment, the antigen binding region of the humanized immunoglobulin comprises a CDR of nonhuman origin. In this embodiment, the humanized immunoglobulin having binding specificity for CCR2 comprises at least one CDR of nonhuman origin. For example, CDRs can be derived from the light and heavy chain variable regions of immunoglobulins of nonhuman origin, such that a humanized immunoglobulin includes substantially heavy chain CDR1, CDR2 and/or CDR3, and/or light chain CDR1, CDR2 and/or CDR3, from one or more immunoglobulins of nonhuman origin, and the resulting humanized immunoglobulin has binding specificity for CCR2. Preferably, all three CDRs of a selected chain are substantially the same as the CDRs of the corresponding chain of a donor, and more preferably, all three CDRs of the light and heavy chains are substantially the same as the CDRs of the corresponding donor chain. In one embodiment, the invention relates to an immunoglobulin having binding specificity for CCR2 comprising a humanized light chain or antigen-binding fragment thereof comprising CDR1, CDR2 and CDR3 of the light chain of the 1D9 antibody and a heavy chain, e.g., a human heavy chain. The invention also includes an immunoglobulin having binding specificity for CCR2 comprising a humanized heavy chain or antigen-binding fragment thereof comprising CDR1, CDR2 and CDR3 of the heavy chain of the 1D9 antibody and a light chain, e.g., a human light chain.

The invention also relates to an immunoglobulin having binding specificity for CCR2 comprising a light chain and a heavy chain, wherein the light chain comprises at least 1 CDR of an antibody of non-human origin (e.g., 1D9) and framework (e.g., SEQ ID

NO: 12, SEQ ID NO: 13, SEQ ID NO:14, SEQ ID NO:15 and SEQ ID NO: 107), and constant regions (e.g., SEQ ID NO:112) of human origin and wherein the heavy chain comprises a variable region of non-human origin (e.g., from 1D9) and a constant region of human origin. The invention also provides antigen-binding fragments of these immunoglobulins. The invention also relates to an immunoglobulin having binding specificity for CCR2 comprising a light chain and a heavy chain, wherein the light chain comprises a variable chain of non-human origin (e.g., from 1D9) and a constant region of human origin, and wherein the heavy chain comprises at least 1 CDR of an antibody of non-human origin (e.g., 1D9) and framework (e.g., SEQ ID NO: 17, SEQ ID NO: 18, SEQ ID NO:19 and SEQ ID NO: 20) and constant regions of human origin (e.g., SEQ ID NO:110). The invention also provides antigen-binding fragments of these immunoglobulins.

The portion of the humanized immunoglobulin or immunoglobulin chain which is of human origin (the human portion) can be derived from any suitable human immunoglobulin or immunoglobulin chain. For example, a human constant region or portion thereof, if present, can be derived from the κ or λ light chains, and/or the γ (e.g., $\gamma 1$, $\gamma 2$, $\gamma 3$, $\gamma 4$), μ , α (e.g., $\alpha 1$, $\alpha 2$), δ or ϵ , heavy chains of human antibodies, including allelic variants. A particular constant region (e.g., IgG1), variant or portions thereof can be selected in order to tailor effector function. For example, a mutated constant region (variant) can be incorporated into a fusion protein to minimize binding to Fc receptors and/or ability to fix complement (see e.g., Example 3; see also, Winter et al., GB 2,209,757 B; Morrison et al., WO 89/07142; Morgan et al., WO 94/29351, December 22, 1994). Mutations L235A and G237A inhibit the binding of the constant region to human Fc receptors and inhibit the initiation of ADCC reactions, and have been described previously in US Patent No.: 5,985,279 and International Publication No.: WO98/06248, which are incorporated herein by reference. The numbering of the residues in the immunoglobulin chain is that of the EU index. See Kabat et al., (1991) *J Immunol.* 147:1709-19. A humanized immunoglobulin of the invention can include one or more of: a light chain constant region of human origin (e.g., SEQ ID NO: 112); and a heavy chain constant region of human origin. A heavy chain constant region of the humanized antibody can further include a mutated heavy chain constant region, e.g., a heavy chain constant region having one or more of mutations L235A and G237A, e.g., a heavy chain constant region of SEQ ID NO: 110.

If present, human framework regions (e.g., of the light chain variable region) are preferably derived from a human antibody variable region having sequence similarity to the analogous or equivalent region (e.g., light chain variable region) of the antigen binding region donor. Other sources of framework regions for portions of human origin of a humanized immunoglobulin include human variable consensus sequences (see, e.g., Kettleborough, C.A. et al., Protein Engineering 4:773-783 (1991); Carter et al., WO 94/04679, published March 3, 1994)). For example, the sequence of the antibody or variable region used to obtain the nonhuman portion can be compared to human sequences as described in Kabat et al., Sequences of Proteins of Immunological Interest, Fifth Edition, U.S. Department of Health and Human Services, U.S. Government Printing Office (1991). In a particularly preferred embodiment, the framework regions of a humanized immunoglobulin chain are derived from a human variable region having at least about 60% overall sequence identity, preferably at least about 70% overall sequence identity and more preferably at least about 85% overall sequence identity, with the variable region of the nonhuman donor (e.g., murine antibody 1D9). A human portion can also be derived from a human antibody having at least about 65% sequence identity, and preferably at least about 70% sequence identity, within the particular portion (e.g., FR) being used, when compared to the equivalent portion (e.g., FR) of the nonhuman donor.

In one embodiment, the humanized immunoglobulin comprises at least one of the framework regions (FR) derived from one or more chains of an antibody of human origin. Thus, the FR can include a FR1 and/or FR2 and/or FR3 and/or FR4 derived from one or more antibodies of human origin. Preferably, the human portion of a selected humanized chain includes FR1, FR2, FR3 and FR4 derived from a variable region of human origin (e.g., from a human immunoglobulin chain, from a human consensus sequence).

The immunoglobulin portions of nonhuman and human origin for use in the present invention have sequences identical to immunoglobulins or immunoglobulin portions from which they are derived or to variants thereof. Such variants include mutants differing by the addition, deletion, or substitution of one or more residues. As indicated above, the CDRs which are of nonhuman origin are substantially the same as in the nonhuman donor, and preferably are identical to the CDRs of the nonhuman donor. As described in Example 2, changes in the framework region, such as those which substitute a residue of the

framework region of human origin with a residue from the corresponding position of the donor, can be made. One or more mutations in the framework region can be made, including deletions, insertions and substitutions of one or more amino acids. Several such substitutions are described in the design of humanized 1D9 antibodies in Example 2. For a selected humanized antibody or chain, framework mutations can be designed as described herein. Preferably, the humanized immunoglobulins can bind CCR2 with an affinity similar to or better than that of the nonhuman donor. Variants can be produced by a variety of suitable methods, including mutagenesis of nonhuman donor or acceptor human chains.

The humanized immunoglobulins of the present invention have binding specificity for human CCR2. In a preferred embodiment, the humanized immunoglobulin of the present invention has at least one functional characteristic of murine antibody 1D9, such as binding function (e.g., having specificity for CCR2, having the same or similar epitopic specificity), and/or inhibitory function (e.g., the ability to inhibit CCR2-dependent function in vitro and/or in vivo, such as the ability to inhibit the binding of a cell bearing CCR2 to a ligand thereof (e.g., a chemokine)). Thus, preferred humanized immunoglobulins can have the binding specificity of the murine antibody 1D9, the epitopic specificity of murine antibody 1D9 (e.g., can compete with murine 1D9, a chimeric 1D9 antibody, or humanized 1D9 for binding to CCR2 (e.g., on a cell bearing CCR2)), and/or inhibitory function of murine antibody 1D9.

The binding function of a humanized immunoglobulin having binding specificity for CCR2 can be detected by standard immunological methods, for example using assays which monitor formation of a complex between humanized immunoglobulin and CCR2 (e.g., a membrane fraction comprising CCR2, on a cell bearing CCR2, human cell line or recombinant host cell comprising nucleic acid encoding CCR2 which expresses CCR2). Binding and/or adhesion assays or other suitable methods can also be used in procedures for the identification and/or isolation of humanized immunoglobulins (e.g., from a library) with the requisite specificity (e.g., an assay which monitors adhesion between a cell bearing CCR2 and a ligand thereof (e.g., HIV, MCP-1, MCP-2, MCP-3, MCP-4, MCP-5, eotaxin), or other suitable methods.

The immunoglobulin portions of nonhuman and human origin for use in the present invention include light chains, heavy chains and portions of light and heavy chains.

These immunoglobulin portions can be obtained or derived from immunoglobulins (e.g., by de novo synthesis of a portion), or nucleic acid molecules encoding an immunoglobulin or chain thereof having the desired property (e.g., binding CCR2, sequence similarity) can be produced and expressed. Humanized immunoglobulins comprising the desired portions (e.g., antigen binding region, CDR, FR, C region) of human and nonhuman origin can be produced using synthetic and/or recombinant nucleic acids to prepare genes (e.g., cDNA) encoding the desired humanized chain. To prepare a portion of a chain, one or more stop codons can be introduced at the desired position. For example, nucleic acid (e.g., DNA) sequences coding for newly designed humanized variable regions can be constructed using PCR mutagenesis methods to alter existing DNA sequences (see e.g., Kamman, M., et al., Nucl. Acids Res. 17:5404 (1989)). PCR primers coding for the new CDRs can be hybridized to a DNA template of a previously humanized variable region which is based on the same, or a very similar, human variable region (Sato, K., et al., Cancer Research 53:851-856 (1993)). If a similar DNA sequence is not available for use as a template, a nucleic acid comprising a sequence encoding a variable region sequence can be constructed from synthetic oligonucleotides (see e.g., Kolbinger, F., Protein Engineering 8:971-980 (1993)). A sequence encoding a signal peptide can also be incorporated into the nucleic acid (e.g., on synthesis, upon insertion into a vector). If the natural signal peptide sequence is unavailable, a signal peptide sequence from another antibody can be used (see, e.g., Kettleborough, C.A., Protein Engineering 4:773-783 (1991)). Using these methods, methods described herein or other suitable methods, variants can be readily produced. In one embodiment, cloned variable regions can be mutagenized, and sequences encoding variants with the desired specificity can be selected (e.g., from a phage library; see e.g., Krebber et al., U.S. 5,514,548; Hoogenboom et al., WO 93/06213, published April 1, 1993)).

The invention relates to a humanized immunoglobulin light chain or antigen-binding fragment thereof, said light chain or antigen-binding fragment thereof having an amino acid sequence comprising at least a functional portion of the light chain variable region amino acid sequence of SEQ ID NO: 9. In a preferred embodiment, the amino acid sequence comprises at least one, preferably two, and more preferably three of the CDRs of SEQ ID NO: 9. The invention also relates to a humanized immunoglobulin heavy chain or antigen-binding fragment thereof, said heavy chain or antigen-binding fragment thereof

having an amino acid sequence comprising at least a functional portion of the heavy chain variable region amino acid sequence shown in SEQ ID NO: 10. In a preferred embodiment, the amino acid sequence comprises at least one, preferably two, and more preferably three of the CDRs of SEQ ID NO: 10. It is noted that all murine sequences described herein are
5 derived from *Mus musculus*.

According to one embodiment of the invention, a humanized immunoglobulin light chain or antigen-binding fragment thereof having binding specificity for CCR2 can comprise an amino acid sequence selected from the group consisting of SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 14, SEQ ID NO: 15 and SEQ ID NO: 107. According to
10 another embodiment of the invention, a humanized immunoglobulin heavy chain or antigen-binding fragment thereof having binding specificity for CCR2 can comprise an amino acid sequence selected from the group consisting of SEQ ID NO: 17, SEQ ID NO: 18, SEQ ID NO: 19, and SEQ ID NO: 20. In a particular embodiment, a humanized immunoglobulin of the invention can comprise both a light chain or antigen-binding fragment thereof having
15 binding specificity for CCR2, comprising an amino acid sequence selected from the group consisting of SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 14, SEQ ID NO: 15 and SEQ ID NO: 107, and a heavy chain or antigen-binding fragment thereof having binding specificity for CCR2 comprising an amino acid sequence selected from the group consisting of SEQ ID NO: 17, SEQ ID NO: 18, SEQ ID NO: 19, and SEQ ID NO: 20. Such humanized
20 immunoglobulins can further include a light chain constant region comprising the amino acid sequence of SEQ ID NO: 112, or a portion thereof, and/or a heavy chain constant region comprising the amino acid sequence of SEQ ID NO: 110, or a portion thereof.

In one embodiment, the invention relates to a humanized immunoglobulin having binding specificity for CCR2 comprising a light chain comprising the amino acid
25 sequence of SEQ ID NO: 12 and a complementary heavy chain, or an antigen-binding fragment of said humanized immunoglobulin having binding specificity for CCR2. In another embodiment, the invention relates to a humanized immunoglobulin having binding specificity for CCR2 comprising a heavy chain comprising the amino acid sequence of SEQ ID NO: 17 and a complementary light chain, or an antigen-binding fragment of said
30 humanized immunoglobulin having binding specificity for CCR2. A complementary light or heavy chain is one which is capable of associating with a selected heavy or light,

respectively, chain, resulting in the ability of an immunoglobulin comprising said complementary heavy and light chains to have binding specificity for CCR2. In a preferred embodiment, the invention relates to a humanized immunoglobulin having binding specificity for CCR2 comprising a light chain comprising the amino acid sequence of SEQ ID NO: 12 and a heavy chain comprising the amino acid sequence of SEQ ID NO: 17, or an antigen-binding fragment of said humanized immunoglobulin having binding specificity for CCR2. In another preferred embodiment, the humanized immunoglobulin can further include a light chain constant region comprising the amino acid sequence of SEQ ID NO: 112, or a portion thereof, and/or a heavy chain constant region comprising the amino acid sequence of SEQ ID NO: 110, or a portion thereof.

In an alternative embodiment, a humanized immunoglobulin of the invention comprises both a light chain comprising the amino acid sequence of SEQ ID NO: 12 and a heavy chain comprising an amino acid sequence selected from the group consisting of SEQ ID NO: 18, SEQ ID NO: 19 and SEQ ID NO: 20, or an antigen-binding fragment of said immunoglobulin having binding specificity for CCR2. In a further embodiment, a humanized immunoglobulin of the invention comprises both a light chain comprising the amino acid sequence of SEQ ID NO: 13 and a heavy chain comprising an amino acid sequence selected from the group consisting of SEQ ID NO: 17, SEQ ID NO: 18, SEQ ID NO: 19 and SEQ ID NO: 20, or an antigen-binding fragment of said immunoglobulin having binding specificity for CCR2. In an additional embodiment, a humanized immunoglobulin of the invention comprises both a light chain comprising the amino acid sequence of SEQ ID NO: 14 and a heavy chain comprising an amino acid sequence selected from the group consisting of SEQ ID NO: 17, SEQ ID NO: 18, SEQ ID NO: 19 and SEQ ID NO: 20, or an antigen-binding fragment of said immunoglobulin having binding specificity for CCR2. In another embodiment, a humanized immunoglobulin of the invention comprises both a light chain comprising the amino acid sequence of SEQ ID NO: 15 and a heavy chain comprising an amino acid sequence selected from the group consisting of SEQ ID NO: 17, SEQ ID NO: 18, SEQ ID NO: 19 and SEQ ID NO: 20, or an antigen-binding fragment of said immunoglobulin having binding specificity for CCR2. In an alternative embodiment, a humanized immunoglobulin of the invention comprises both a light chain comprising the amino acid sequence of SEQ ID NO: 107 and a heavy chain comprising an amino acid

sequence selected from the group consisting of SEQ ID NO: 17, SEQ ID NO: 18, SEQ ID NO: 19 and SEQ ID NO: 20, or an antigen-binding fragment of said immunoglobulin having binding specificity for CCR2.

In another embodiment, a humanized immunoglobulin of the invention
 5 comprises both a light chain comprising an amino acid sequence selected from the group consisting of SEQ ID NO: 13, SEQ ID NO: 14, SEQ ID NO: 15 and SEQ ID NO: 107 and a heavy chain comprising the amino acid sequence of SEQ ID NO: 17, or an antigen-binding fragment of said immunoglobulin having binding specificity for CCR2. In an alternate
 10 embodiment, a humanized immunoglobulin of the invention comprises both a light chain comprising an amino acid sequence selected from the group consisting of SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 14, SEQ ID NO: 15 and SEQ ID NO: 107 and a heavy chain comprising the amino acid sequence of SEQ ID NO: 18, or an antigen-binding fragment of said immunoglobulin having binding specificity for CCR2. In a further embodiment, a
 15 humanized immunoglobulin of the invention comprises both a light chain comprising an amino acid sequence selected from the group consisting of SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 14, SEQ ID NO: 15 and SEQ ID NO: 107 and a heavy chain comprising the amino acid sequence of SEQ ID NO: 19, or an antigen-binding fragment of said immunoglobulin having binding specificity for CCR2. In an additional embodiment, a
 20 humanized immunoglobulin of the invention comprises both a light chain comprising an amino acid sequence selected from the group consisting of SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 14, SEQ ID NO: 15 and SEQ ID NO: 107 and a heavy chain comprising the amino acid sequence of SEQ ID NO: 20, or an antigen-binding fragment of said immunoglobulin having binding specificity for CCR2.

In one embodiment, the humanized immunoglobulin light chain or antigen-
 25 binding fragment thereof having binding specificity for CCR2 can be encoded by a nucleic acid molecule comprising SEQ ID NO: 109. The humanized immunoglobulin light chain or antigen binding fragment thereof can further include a constant region encoded by a nucleic acid of SEQ ID NO: 113. Preferably, the humanized immunoglobulin light chain is encoded by a nucleic acid of SEQ ID NO: 109 and linked to a nucleic acid of SEQ ID NO: 113. In
 30 another embodiment, the humanized immunoglobulin heavy chain or antigen-binding fragment thereof having binding specificity for CCR2 can be encoded by a nucleic acid

molecule comprising SEQ ID NO: 108. The humanized immunoglobulin heavy chain or antigen binding fragment thereof can further include a constant region encoded by a nucleic acid of SEQ ID NO:111. Preferably, the humanized immunoglobulin light chain is encoded by a nucleic acid of SEQ ID NO:108 and linked to a nucleic acid of SEQ ID NO:111.

5 The invention also relates to a chimeric immunoglobulin or antigen-binding fragment thereof having binding specificity for CCR2 comprising a light chain variable region of nonhuman origin and a human constant region (e.g., a light chain constant region). The invention further relates to a chimeric immunoglobulin or antigen-binding fragment thereof having binding specificity for CCR2 comprising a heavy chain variable region of nonhuman origin and a human constant region (e.g., a heavy chain constant region). In 10 another embodiment, the chimeric immunoglobulin or antigen-binding fragment thereof having binding specificity for CCR2 comprises a light chain variable chain region of nonhuman origin and a heavy chain variable region of nonhuman origin and further comprises a human constant region (e.g., a human light chain constant region and/or a human 15 heavy chain constant region).

Nucleic Acids and Constructs

The present invention also relates to isolated and/or recombinant (including, e.g., essentially pure) nucleic acid molecules comprising nucleic acid sequences which 20 encode a humanized immunoglobulin or humanized immunoglobulin light or heavy chain of the present invention.

Nucleic acid molecules referred to herein as "isolated" are nucleic acid molecules which have been separated away from the nucleic acids of the genomic DNA or cellular RNA of their source of origin (e.g., as it exists in cells or in a mixture of nucleic 25 acids such as a library), and include nucleic acid molecules obtained by methods described herein or other suitable methods, including essentially pure nucleic acid molecules, nucleic acid molecules produced by chemical synthesis, by combinations of biological and chemical methods, and recombinant nucleic acid molecules which are isolated (see e.g., Daugherty, B.L. et al., Nucleic Acids Res., 19(9): 2471-2476 (1991); Lewis, A.P. and J.S. Crowe, Gene, 30 101: 297-302 (1991)).

Nucleic acid molecules referred to herein as "recombinant" are nucleic acid molecules which have been produced by recombinant DNA methodology, including those nucleic acid molecules that are generated by procedures which rely upon a method of artificial recombination, such as the polymerase chain reaction (PCR) and/or cloning into a vector using restriction enzymes. "Recombinant" nucleic acid molecules are also those that result from recombination events that occur through the natural mechanisms of cells, but are selected for after the introduction to the cells of nucleic acids designed to allow and make probable a desired recombination event.

The present invention also relates more specifically to isolated and/or recombinant nucleic acid molecules comprising a nucleotide sequence which encodes a humanized 1D9 immunoglobulin (i.e., a humanized immunoglobulin of the present invention in which the nonhuman portion is derived from the murine monoclonal antibody 1D9) or chain thereof. In one embodiment, the light chain comprises three complementarity determining regions derived from the light chain of the 1D9 antibody, and the heavy chain comprises three complementarity determining regions derived from the heavy chain of the 1D9 antibody. Such nucleic acid molecules include, for example, (a) a nucleic acid molecule comprising a sequence which encodes a polypeptide comprising the amino acid sequence of the heavy chain variable region of a humanized 1D9 immunoglobulin (e.g., heavy chain variable region of Figures 8 and 21) (e.g., nucleotides 58-411 of SEQ ID NO: 96); (b) a nucleic acid molecule comprising a sequence which encodes a polypeptide comprising the amino acid sequence of the light chain variable region of a humanized 1D9 immunoglobulin (e.g., light chain variable region of Figures 7 and 22) (e.g., nucleotides 52-390 of SEQ ID NO: 95); (c) a nucleic acid molecule comprising a sequence which encodes at least a functional portion of the light or heavy chain variable region of a humanized 1D9 immunoglobulin (e.g., a portion sufficient for antigen binding of a humanized immunoglobulin which comprises said chain). In one embodiment, the nucleic acid comprises the nucleotide sequence of the variable region as set forth or substantially as set forth in Figure 21 or as set forth or substantially as set forth in Figure 22, including double or single-stranded polynucleotides. (Although various figures may illustrate polypeptides which are larger than the variable region (i.e., include a signal peptide coding sequence or a portion of a constant region coding sequence), reference to the variable region of a particular

figure is meant to include the variable region portion of the sequence shown). In another preferred embodiment, the nucleic acid sequence encoding the light chain or antigen binding portion thereof includes the nucleotide sequence of SEQ ID NO:109 and/or a nucleotide sequence of SEQ ID NO:113. In another preferred embodiment, the nucleic acid encoding the heavy chain or antigen binding portion thereof includes the nucleotide sequence of SEQ ID NO:108 and/or a nucleotide sequence of SEQ ID NO:111. Due to the degeneracy of the genetic code, a variety of nucleic acids can be made which encode a selected polypeptide. Accordingly, the invention also features a nucleic acid encoding a light chain that includes a nucleic acid encoding the variable light chain of SEQ ID NO:12 and/or a nucleic acid encoding the constant light chain of SEQ ID NO: 112. The invention also features a nucleic acid encoding a heavy chain that includes a nucleic acid encoding the variable heavy chain of SEQ ID NO:17 and/or a nucleic acid encoding the constant heavy chain of SEQ ID NO: 110. Isolated and/or recombinant nucleic acid molecules meeting these criteria can comprise nucleic acid molecules encoding sequences identical to sequences of humanized 1D9 antibody or variants thereof as discussed above.

Nucleic acid molecules of the present invention can be used in the production of humanized immunoglobulins having binding specificity for CCR2. For example, a nucleic acid molecule (e.g., DNA) encoding a humanized immunoglobulin of the present invention can be incorporated into a suitable construct (e.g., a vector) for further manipulation of sequences or for production of the encoded polypeptide in suitable host cells. Vectors containing such nucleic acids preferably express the humanized immunoglobulins described herein, e.g., express one or more of the variable light chain of SEQ ID NO:12, the constant light chain of SEQ ID NO: 112, the variable heavy chain of SEQ ID NO:17 and the heavy chain constant region of SEQ ID NO: 110. The nucleic acid sequences for insertion into the vector can include, e.g., additional sequences on one or more ends (such as restriction endonuclease sites, restriction endonucleases complementary to sites available in the vector or flanking sequences) such that, when properly inserted into the vectors, the vector will not express amino acids encoded by those additional sequences. Examples of nucleic acid sequences that include such additional sequences are set forth in SEQ ID NO:98, SEQ ID NO: 117, SEQ ID NO: 97 and SEQ ID NO:115.

The vector can include an operable linkage between a leader sequence and a stable immunoglobulin domain (*e.g.*, constant region) that includes sequences amenable to digestion using restriction endonucleases. Preferred restriction endonuclease sites engineered into the operable linkage include EcoRV, PpuMI or MfeI and BlnI or BsiWI, however, additional or alternative sites may be engineered in order to accommodate the particularly desired additional sequences used in conjunction with the DNA cassette. Methods for engineering restriction sites (*e.g.*, site directed mutagenesis) and determination of preferred sites through sequence analyses are well known in the art. Restriction sites engineered in the linkage allow for subsequent incorporation of additional immunoglobulin sequences (*e.g.*, variable region) such that the additional sequences are incorporated in operable linkage with the leader and stable domain (*e.g.*, constant region) to allow production of a nucleic acid encoding a complete operable immunoglobulin molecule (*e.g.*, cloning of the additional variable sequence remains in frame with the leader and constant region to produce nucleic acid encoding an immunoglobulin protein including leader, variable, and constant domains).

An immunoglobulin variable domain nucleic acid sequence can be isolated, wherein the 5' and 3' ends of the insert sequences comprise sequences available for restriction endonucleases complementary to sites available in the DNA cassette. In one embodiment, immunoglobulin variable sequences are isolated using PCR techniques known in the art, wherein primer sequences are engineered to incorporate DNA restriction endonuclease sequences in the proper placement to allow operable linkage upon incorporation into the immunoglobulin DNA cassette.

Targeting Molecules

The invention also relates to targeting molecules which can effectuate the interaction of a CCR2-expressing cell with a target cell. The targeting molecule includes a first binding moiety which can bind mammalian CCR2, and a second binding moiety which can bind a molecule expressed on the surface of a target cell. Preferred target cells include tumor cells and virus infected cells. A variety of molecules which are expressed at higher levels or uniquely on tumor cells (*e.g.*, tumor antigens, such as Lewis Y, HER-2/neu, disialoganglioside G3, carcinoembryonic antigen, CD30) and/or virus infected cells (*e.g.*,

viral antigens, such as influenza virus hemagglutinin, Epstein-Barr virus LMP-1, hepatitis C virus E2 glycoprotein, HIV gp160, HIV gp 120) are known in the art. The targeting molecule can contain any suitable binding second moiety which binds to a molecule expressed on a desired target cell (see, for example Ring, U.S. Patent No. 5,948,647, the entire teachings of which are incorporated herein by reference). Suitable binding moieties include, for example, proteins and peptides (including post-translationally modified forms e.g., glycosylated, phosphorylated, lipidated), sugars, lipids, peptidomimetics, small organic molecules, nucleic acids and other agents which bind mammalian CCR2 or a molecule expressed on the surface of a target cell. Suitable binding moieties can be identified using any suitable method, such as the binding assays described herein.

In a preferred embodiment, the first binding moiety can be, for example, a humanized immunoglobulin of the invention which binds mammalian CCR2 or antigen-binding fragment thereof (e.g., Fab, Fv, Fab', F(ab')₂). The second binding moiety can be, for example, an antibody (e.g., a second humanized immunoglobulin) or antigen-binding fragment thereof which binds to a molecule expressed on the target cell or antigen binding fragment thereof. Where the targeting molecule comprises a first binding moiety which is a humanized anti-CCR2 immunoglobulin or antigen-binding fragment thereof, it is preferred that the humanized anti-CCR2 immunoglobulin does not inhibit binding of ligand to CCR2.

The first binding moiety can be directly or indirectly bonded to the second binding moiety through a variety of suitable linkages. For example, when the first binding moiety and the second binding moiety are both proteins or peptides, the moieties can be part of a contiguous polypeptide (i.e., a fusion protein). Where the targeting molecule is a fusion protein, the first and second binding moieties can be arranged on the polypeptide in any suitable configuration. The first and second binding moieties can be indirectly bonded through a (i.e., one or more) peptide linker, or bonded directly to each other through a peptide bond.

Where the binding moieties are not part of a contiguous polypeptide they can be directly bonded by a chemical bond formed by reaction of a functional group (or activated derivative thereof) on the first moiety with a second functional group (or activated derivative thereof) on the second moiety. For example, two thiols can react to form a disulfide bond and an amine can react with a carboxylic acid or acyl halide to form an amide. A variety of

other suitable reactions which can be used are known in the art (see, for example, Hermanson, G. T., *Bioconjugate Techniques*, Academic Press: San Diego, CA (1996)). The binding moieties can be indirectly bonded through a suitable linker (e.g., a peptide linker). Generally, a linker contains two reactive groups which can react to form bonds with the first binding moiety and/or the second binding moiety. Linkers which contain two different reactive groups (e.g., a heterobifunctional linker) can be used to selectively conjugate the first binding moiety to the second binding moiety. Many linkers which are suitable for forming conjugates between proteins, nucleic acids, peptides, vitamins, sugars, lipids, small organic molecules and other suitable agents are known (see, for example, U.S. Patent Nos. 5,856,571, 5,880,270; Hermanson, G. T., *Bioconjugate Techniques*, Academic Press: San Diego, CA (1996)).

Preferably, the independent activities of the binding moieties (e.g., binding activities, chemoattractant activity) of the targeting molecule are not significantly different from the activities of the binding moieties as separate molecular entities. For example, where the first binding moiety is a humanized immunoglobulin or antigen-binding fragment that binds CCR2, the targeting molecule can bind to CCR2 with an affinity which is within a factor of about 1000, preferably within a factor of 100, more preferably within a factor of 10 or substantially the same as the affinity of the free antibody or antigen-binding fragment. Target molecules with these preferred characteristics can be prepared using any suitable method. The resulting targeting molecule can then be assayed for binding (e.g., by ELISA) and for chemoattractant activity.

In one embodiment, the targeting molecule is a bispecific humanized antibody or bispecific antigen-binding fragment thereof (e.g., $F(ab')_2$) which has specificity for mammalian CCR2 and a molecule expressed on a target cell (e.g., tumor antigen, viral antigen). Bispecific antibodies can be secreted by triomas and hybrid hybridomas. The supernatants of triomas and hybrid hybridomas can be assayed for bispecific antibody using a suitable assay (e.g., ELISA), and bispecific antibodies can be purified using conventional methods. These antibodies can then be humanized according to methods described herein. Thus, the invention provides a targeting molecule which is a humanized bispecific antibody having binding specificity for CCR2 and an antigen expressed on a target cell, or a bivalent antigen-binding fragment of the bispecific antibody. The invention also relates to a method

of effectuating the interaction of a CCR2-bearing cell with a target cell in a patient, comprising administering to the patient an effective amount of a targeting molecule which is a humanized bispecific antibody having binding specificity for CCR2 and an antigen expressed on a target cell, or a bivalent antigen-binding fragment of the bispecific antibody.

5

Method of Producing Humanized Immunoglobulins Having Specificity for CCR2

Another aspect of the invention relates to a method of preparing a humanized immunoglobulin which has binding specificity for CCR2. The humanized immunoglobulin can be obtained, for example, by the expression of one or more recombinant nucleic acids
10 encoding a humanized immunoglobulin having binding specificity for CCR2 in a suitable host cell, for example.

Constructs or expression vectors suitable for the expression of a humanized immunoglobulin having binding specificity for CCR2 are also provided. The constructs can be introduced into a suitable host cell, and cells which express a humanized immunoglobulin
15 of the present invention can be produced and maintained in culture. Suitable host cells can be prokaryotic, including bacterial cells such as *E. coli*, *B. subtilis* and or other suitable bacteria, or eucaryotic, such as fungal or yeast cells (e.g., *Pichia pastoris*, *Aspergillus* species, *Saccharomyces cerevisiae*, *Schizosaccharomyces pombe*, *Neurospora crassa*), or other lower eucaryotic cells, and cells of higher eucaryotes such as those from insects (e.g., Sf9 insect
20 cells (WO 94/26087, O'Connor, published November 24, 1994)) or mammals (e.g., COS cells, such as COS-1 (ATCC Accession No. CRL-1650) and COS-7 (ATCC Accession No. CRL-1651), CHO (e.g., ATCC Accession No. CRL-9096), 293 (ATCC Accession No. CRL-1573), HeLa (ATCC Accession No. CCL-2), CV1 (ATCC Accession No. CCL-70), WOP (Dailey et al., *J. Virol.* 54:739-749 (1985)), 3T3, 293T (Pear et al., *Proc. Natl. Acad. Sci.*
25 U.S.A., 90:8392-8396 (1993)), NSO cells, SP2/0, HuT 78 cells, and the like (see, e.g., Ausubel, F.M. et al., eds. *Current Protocols in Molecular Biology*, Greene Publishing Associates and John Wiley & Sons Inc., (1993)). Preferred mammalian host cells for expressing the antibodies of the invention include Chinese Hamster Ovary (CHO cells) (including dhfr- CHO cells, described in Urlaub and Chasin, (1980) *Proc. Natl. Acad. Sci.*
30 *USA* 77:4216-4220, used with a DHFR selectable marker, e.g., as described in R.J. Kaufman and P.A. Sharp (1982) *Mol. Biol.* 159:601-621).

Host cells which produce a humanized immunoglobulin having binding specificity for CCR2 can be produced as follows. For example, a nucleic acid encoding all or part of the coding sequence for the desired humanized immunoglobulin can be inserted into a nucleic acid vector, e.g., a DNA vector, such as a plasmid, virus or other suitable replicon for expression. A variety of vectors are available, including vectors which are maintained in single copy or multiple copy, or which become integrated into the host cell chromosome.

Suitable expression vectors can contain a number of components, including, but not limited to one or more of the following: an origin of replication; a selectable marker gene; one or more expression control elements, such as a transcriptional control element (e.g., a promoter, an enhancer, terminator), and/or one or more translation signals; a signal sequence or leader sequence for membrane targeting or secretion. In a construct, a signal sequence can be provided by the vector or other source. For example, the transcriptional and/or translational signals of an immunoglobulin can be used to direct expression.

A promoter can be provided for expression in a suitable host cell. Promoters can be constitutive or inducible. For example, a promoter can be operably linked to a nucleic acid encoding a humanized immunoglobulin or immunoglobulin chain, such that it directs expression of the encoded polypeptide. A variety of suitable promoters for procaryotic (e.g., lac, tac, T3, T7 promoters for *E. coli*) and eucaryotic (e.g., yeast alcohol dehydrogenase (ADH1), SV40, CMV, EF-1 α) hosts are available.

In addition, the expression vectors typically comprise a selectable marker for selection of host cells carrying the vector, and, in the case of replicable expression vector, an origin of replication. Genes encoding products which confer antibiotic or drug resistance are common selectable markers and may be used in procaryotic (e.g., β -lactamase gene (ampicillin resistance), Tet gene for tetracycline resistance) and eucaryotic cells (e.g., neomycin (G418 or geneticin), gpt (mycophenolic acid), ampicillin, or hygromycin resistance genes). Dihydrofolate reductase marker genes permit selection with methotrexate in a variety of hosts. Genes encoding the gene product of auxotrophic markers of the host (e.g., LEU2, URA3, HIS3) are often used as selectable markers in yeast. Use of viral (e.g., baculovirus) or phage vectors, and vectors which are capable of integrating into the genome of the host cell, such as retroviral vectors, are also contemplated. In one embodiment, the

vector is pLKTOK38. The present invention also relates to cells carrying these expression vectors.

An expression vector comprising a fused gene encoding a humanized immunoglobulin light chain, said gene comprising a nucleotide sequence encoding a CDR
5 derived from a light chain of a nonhuman antibody having binding specificity for CCR2 and a framework region derived from a light chain of human origin, and, optionally, a constant region, e.g., a constant region described herein.

Thus, the invention includes an expression vector comprising a gene encoding a humanized immunoglobulin light chain, said gene comprising a nucleotide sequence
10 encoding a CDR derived from a light chain of a nonhuman antibody having binding specificity for CCR2 and a framework region derived from a light chain of human origin, and, optionally, a nucleotide sequence encoding all or a portion of a light chain constant region, e.g., a light chain constant region described herein. The invention also relates to an expression vector comprising a gene encoding a humanized immunoglobulin heavy chain,
15 said gene comprising a nucleotide sequence encoding a CDR derived from a heavy chain of a nonhuman antibody having binding specificity for CCR2 and a framework region derived from a heavy chain of human origin, and, optionally, a nucleotide sequence encoding all or a portion of a heavy chain constant region, e.g., a heavy chain constant region described herein. In one embodiment, the nonhuman antibody is murine antibody 1D9. The invention
20 also includes host cells comprising the expression vectors of the invention. The invention also relates to an isolated or recombinant gene encoding a humanized immunoglobulin light or heavy chain comprising a first nucleic acid sequence encoding an antigen binding region derived from murine monoclonal antibody 1D9; and a second nucleic acid sequence encoding at least a portion of a constant region of an immunoglobulin of human origin, or a
25 mutated constant region of an immunoglobulin of human origin as described herein.

The invention also relates to a host cell (e.g., which expresses a humanized immunoglobulin or an antigen binding fragment thereof having specificity for CCR2) comprising a first recombinant nucleic acid molecule encoding a humanized immunoglobulin light chain or fragment thereof and a second recombinant nucleic acid molecule encoding a
30 humanized immunoglobulin heavy chain or fragment thereof, wherein said first nucleic acid molecule comprises a nucleotide sequence encoding a CDR derived from the light chain of

murine antibody 1D9 and a framework region derived from a light chain of human origin, and wherein said second nucleic acid molecule comprises a nucleotide sequence encoding a CDR derived from the heavy chain of murine antibody 1D9 and a framework region derived from a heavy chain of human origin. The first nucleic acid molecule can further comprise a nucleic acid molecule encoding all or a portion of a light chain constant region described herein and/or the second nucleic acid molecule can further comprise a nucleic acid molecule encoding all or a portion of a heavy chain constant region described herein. The invention also includes a method of preparing a humanized immunoglobulin or antigen-binding fragment thereof comprising maintaining a host cell of the invention under conditions appropriate for expression of a humanized immunoglobulin, whereby humanized immunoglobulin chains are expressed and a humanized immunoglobulin or antigen-binding fragment thereof having specificity for CCR2 is produced. The method can further comprise the step of isolating the humanized immunoglobulin or fragment thereof.

For example, a nucleic acid molecule (i.e., one or more nucleic acid molecules) encoding the heavy and light chains of a humanized immunoglobulin having binding specificity for CCR2, or a construct (i.e., one or more constructs) comprising such nucleic acid molecule(s), can be introduced into a suitable host cell by a method appropriate to the host cell selected (e.g., transformation, transfection, electroporation, infection), such that the nucleic acid molecule(s) are operably linked to one or more expression control elements (e.g., in a vector, in a construct created by processes in the cell, integrated into the host cell genome). Host cells can be maintained under conditions suitable for expression (e.g., in the presence of inducer, suitable media supplemented with appropriate salts, growth factors, antibiotic, nutritional supplements, etc.), whereby the encoded polypeptide(s) are produced. If desired, the encoded protein (e.g., humanized 1D9 antibody) can be isolated from, e.g., the host cells, medium, milk. This process encompasses expression in a host cell of a transgenic animal (see e.g., WO 92/03918, GenPharm International, published March 19, 1992).

Fusion proteins can be produced in which a humanized immunoglobulin or immunoglobulin chain is linked to a non-immunoglobulin moiety (i.e., a moiety which does not occur in immunoglobulins as found in nature) in an N-terminal location, C-terminal location or internal to the fusion protein. For example, some embodiments can be produced

by the insertion of a nucleic acid encoding immunoglobulin sequences into a suitable expression vector, such as a pET vector (e.g., pET-15b, Novagen), a phage vector (e.g., pCANTAB 5 E, Pharmacia), or other vector (e.g., pRIT2T Protein A fusion vector, Pharmacia). The resulting construct can be introduced into a suitable host cell for expression. Upon expression, some fusion proteins can be isolated or purified from a cell lysate by means of a suitable affinity matrix (see e.g., Current Protocols in Molecular Biology (Ausubel, F.M. et al., eds., Vol. 2, Suppl. 26, pp. 16.4.1-16.7.8 (1991))).

Therapeutic Methods and Compositions

The present invention provides humanized immunoglobulins which (1) can bind CCR2 *in vitro* and/or *in vivo*; and/or (2) can modulate an activity or function of CCR2, such as (a) binding function (e.g., the ability of CCR2 to bind to a ligand) and/or (b) leukocyte trafficking, including recruitment and/or accumulation of leukocytes in tissues. Preferably the humanized immunoglobulins are capable of selectively binding CCR2 *in vitro* and/or *in vivo*, and inhibiting CCR2-mediated interactions. In one embodiment, a humanized immunoglobulin can bind CCR2, and can inhibit binding of CCR2 to one or more of its ligands (e.g., HIV, MCP-1, MCP-2, MCP-3, MCP-4, MCP-5, eotaxin).

The humanized immunoglobulins of the present invention are useful in a variety of processes with applications in research, diagnosis and therapy. For instance, they can be used to detect, isolate, and/or purify CCR2 or variants thereof (e.g., by affinity purification or other suitable methods), and to study CCR2 structure (e.g., conformation) and function. The humanized immunoglobulins of the present invention can also be used in diagnostic applications (e.g., *in vitro*, *ex vivo*) or to modulate CCR2 function in therapeutic (including prophylactic) applications.

For example, the humanized immunoglobulins of the present invention can be used to detect and/or measure the level of CCR2 in a sample (e.g., tissues or body fluids, such as an inflammatory exudate, blood, serum, bowel fluid, on cells bearing CCR2). For example, a sample (e.g., tissue and/or body fluid) can be obtained from an individual and a suitable immunological method can be used to detect and/or measure CCR2 expression, including methods such as enzyme-linked immunosorbent assays (ELISA), including chemiluminescence assays, radioimmunoassay, and immunohistology. In one embodiment, a

method of detecting a selected CCR2 in a sample is provided, comprising contacting a sample with a humanized immunoglobulin of the present invention under conditions suitable for specific binding of the humanized immunoglobulin to CCR2 and detecting antibody-CCR2 complexes which are formed. In an application of the method, humanized immunoglobulins can be used to analyze normal versus inflamed tissues (e.g., from a human) for CCR2 reactivity and/or expression (e.g., immunohistologically), to detect associations between particular conditions and increased expression of CCR2 (e.g., in affected tissues). The humanized immunoglobulins of the present invention permit immunological methods of assessment of the presence of CCR2 in normal versus inflamed tissues, through which the presence of disease, disease progress and/or the efficacy of anti-CCR2 integrin therapy in inflammatory disease can be assessed. In addition, the humanized immunoglobulins of the invention provide methods for assessing the presence of CCR2 though the progression of unwanted cellular proliferation associated with CCR2 expression, e.g., a cancer that expresses CCR2.

The humanized immunoglobulins of the present invention can also be used to modulate (e.g., inhibit (reduce or prevent)) binding function and/or leukocyte (e.g., lymphocyte, monocyte) trafficking modulated by CCR2. For example, humanized immunoglobulins which inhibit the binding of CCR2 to a ligand (i.e., one or more ligands) can be administered according to the method in the treatment of diseases associated with leukocyte (e.g., lymphocyte, monocyte) infiltration of tissues. Additionally, humanized immunoglobulins which inhibit the binding of CCR2 to a ligand (i.e., one or more ligands) can be administered according to the method in the treatment of HIV. An effective amount of a humanized immunoglobulin of the present invention (i.e., one or more) is administered to an individual (e.g., a mammal, such as a human or other primate) in order to treat such a disease.

The humanized immunoglobulin is administered in an effective amount which inhibits binding of CCR2 to a ligand thereof. For therapy, an effective amount will be sufficient to achieve the desired therapeutic (including prophylactic) effect (such as an amount sufficient to reduce or prevent CCR2-mediated binding and/or signaling). The humanized immunoglobulin can be administered in a single dose or multiple doses. The dosage can be determined by methods known in the art and can be dependent, for example,

upon the individual's age, sensitivity, tolerance and overall well-being. Suitable dosages for antibodies can be from about 0.1 mg/kg body weight to about 10.0 mg/kg body weight per treatment. For example, the dose can be about 1 mg/kg body weight to about 7 mg/kg body weight, about 3 mg/kg body weight to about 5 mg/kg body weight.

5 According to the method, the humanized immunoglobulin can be administered to an individual (e.g., a human) alone or in conjunction with another agent. A humanized immunoglobulin can be administered before, along with or subsequent to administration of the additional agent. Thus, the invention includes pharmaceutical compositions comprising a humanized immunoglobulin or antigen-binding fragment thereof of the invention and a
10 suitable carrier. In one embodiment, more than one humanized immunoglobulin which inhibits the binding of CCR2 to its ligands is administered. In another embodiment an additional monoclonal antibody is administered in addition to a humanized immunoglobulin of the present invention. In yet another embodiment, an additional pharmacologically active ingredient (e.g., an antiinflammatory compound, such as sulfasalazine, another non-steroidal
15 antiinflammatory compound, or a steroidal antiinflammatory compound) can be administered in conjunction with a humanized immunoglobulin of the present invention.

 A variety of routes of administration are possible, including, but not necessarily limited to, parenteral (e.g., intravenous, intraarterial, intramuscular, subcutaneous injection), oral (e.g., dietary), topical, inhalation (e.g., intrabronchial, intranasal or oral
20 inhalation, intranasal drops), or rectal, depending on the disease or condition to be treated. Parenteral administration is a preferred mode of administration.

 Formulation will vary according to the route of administration selected (e.g., solution, emulsion). An appropriate composition comprising the humanized antibody to be administered can be prepared in a physiologically acceptable vehicle or carrier. For solutions
25 or emulsions, suitable carriers include, for example, aqueous or alcoholic/aqueous solutions, emulsions or suspensions, including saline and buffered media. Parenteral vehicles can include sodium chloride solution, Ringer's dextrose, dextrose and sodium chloride, lactated Ringer's or fixed oils. Intravenous vehicles can include various additives, preservatives, or fluid, nutrient or electrolyte replenishers (See, generally, Remington's Pharmaceutical
30 Sciences, 17th Edition, Mack Publishing Co., PA, 1985). For inhalation, the compound can

be solubilized and loaded into a suitable dispenser for administration (e.g., an atomizer, nebulizer or pressurized aerosol dispenser).

Thus, the invention includes a method of inhibiting HIV infection of a cell, comprising contacting a cell with an effective amount of a composition comprising a humanized immunoglobulin or antigen-binding fragment thereof of the invention. The invention also relates to a method of treating HIV or inhibiting HIV infection in a patient comprising administering to the patient a composition comprising an effective amount of a humanized immunoglobulin or antigen-binding fragment thereof of the invention.

The invention also relates to a method of inhibiting a function associated with binding of a chemokine to mammalian CCR2 or a functional portion of CCR2, comprising contacting a composition comprising CCR2 or portion thereof with an effective amount of a humanized immunoglobulin or antigen-binding fragment thereof of the invention, wherein said humanized immunoglobulin inhibits binding of the chemokine to mammalian CCR2 and inhibits one or more functions associated with binding of the chemokine to CCR2. For example, the chemokine can be selected from the group consisting of MCP-1, MCP-2, MCP-3, MCP-4, MCP-5, eotaxin and combinations thereof.

The invention also relates to a method of inhibiting leukocyte trafficking in a patient, comprising administering to the patient a composition comprising an effective amount of a humanized immunoglobulin or antigen-binding fragment thereof of the invention which binds to mammalian CCR2 and inhibits binding of a ligand to the receptor. For example, the ligand can be a chemokine (e.g., MCP-1, MCP-2, MCP-3, MCP-4, MCP-5, eotaxin) or HIV.

The invention also relates to a method of inhibiting the interaction of a first cell expressing CCR2 with a ligand (e.g., on a second cell expressing a ligand of CCR2), comprising contacting the first cell with an effective amount of a humanized immunoglobulin or antigen-binding fragment thereof of the invention, particularly wherein said immunoglobulin or fragment inhibits the binding of ligand to CCR2. For example, the cell can be selected from the group consisting of lymphocytes, monocytes, granulocytes, T cells, basophils, and cells comprising a recombinant nucleic acid encoding CCR2 or a portion thereof. In one embodiment, the ligand is a chemokine (e.g., MCP-1, MCP-2, MCP-3, MCP-4, MCP-5, eotaxin). In another embodiment, the ligand is HIV.

The invention also includes a method of treating a CCR2-mediated disorder in a patient, comprising administering to the patient an effective amount of a humanized immunoglobulin or antigen-binding fragment thereof of the invention which binds to mammalian CCR2. The disorder can include, but is not limited to, allergy, atherogenesis, anaphylaxis, malignancy, chronic and acute inflammatory disorders, histamine and IgE-mediated allergic reactions, shock, and rheumatoid arthritis, atherosclerosis, multiple sclerosis, stenosis, restenosis, allograft rejection, fibrotic disease, asthma, scleroderma, inflammatory glomerulopathies, and immune-related diseases.

In a particular embodiment, the invention relates to a method of inhibiting restenosis in a patient, comprising administering to the patient an effective amount of a humanized immunoglobulin or antigen-binding fragment thereof of the invention which binds to mammalian CCR2. The invention also includes a humanized immunoglobulin or antigen-binding fragment thereof of the invention for use in therapy or diagnosis or for use in treating a CCR2-mediated disease or disorder. The invention also includes the use of a humanized immunoglobulin or antigen-binding fragment thereof of the invention for the manufacture of a medicament for treating a CCR2-mediated disease.

Anti-idiotypic antibodies are also provided. Anti-idiotypic antibodies recognize antigenic determinants associated with the antigen-binding site of another antibody. Anti-idiotypic antibodies can be prepared against second antibody by immunizing an animal of the same species, and preferably of the same strain, as the animal used to produce the second antibody. See e.g., U.S. Patent No. 4,699,880.

The present invention also pertains to the hybridoma cell lines deposited under ATCC Accession Nos. HB-12549 and HB-12550, as well as to the monoclonal antibodies produced by the hybridoma cell lines deposited under ATCC Accession Nos. HB-12549 and HB-12550. The cell lines of the present invention have uses other than for the production of the monoclonal antibodies. For example, the cell lines of the present invention can be fused with other cells (such as suitably drug-marked human myeloma, mouse myeloma, human-mouse heteromyeloma or human lymphoblastoid cells) to produce additional hybridomas, and thus provide for the transfer of the genes encoding the monoclonal antibodies. In addition, the cell lines can be used as a source of nucleic acids encoding the anti-CCR2 immunoglobulin chains, which can be isolated and expressed (e.g.,

upon transfer to other cells using any suitable technique (see e.g., Cabilly et al., U.S. Patent No. 4,816,567; Winter, U.S. Patent No. 5,225,539)). For instance, clones comprising a rearranged anti-CCR2 light or heavy chain can be isolated (e.g., by PCR) or cDNA libraries can be prepared from mRNA isolated from the cell lines, and cDNA clones encoding an anti-CCR2 immunoglobulin chain can be isolated. Thus, nucleic acids encoding the heavy and/or light chains of the antibodies or portions thereof can be obtained and used in accordance with recombinant DNA techniques for the production of the specific immunoglobulin, immunoglobulin chain, or variants thereof (e.g., humanized immunoglobulins) in a variety of host cells or in an in vitro translation system. For example, the nucleic acids, including cDNAs, or derivatives thereof encoding variants such as a humanized immunoglobulin or immunoglobulin chain, can be placed into suitable prokaryotic or eukaryotic vectors (e.g., expression vectors) and introduced into a suitable host cell by an appropriate method (e.g., transformation, transfection, electroporation, infection), such that the nucleic acid is operably linked to one or more expression control elements (e.g., in the vector or integrated into the host cell genome). For production, host cells can be maintained under conditions suitable for expression (e.g., in the presence of inducer, suitable media supplemented with appropriate salts, growth factors, antibiotic, nutritional supplements, etc.), whereby the encoded polypeptide is produced. If desired, the encoded protein can be recovered and/or isolated (e.g., from the host cells, medium, milk). It will be appreciated that the method of production encompasses expression in a host cell of a transgenic animal (see e.g., WO 92/03918, GenPharm International, published March 19, 1992).

As described herein, antibodies and functional fragments thereof of the present invention can block (inhibit) binding of a ligand to CCR2 and/or inhibit function associated with binding of the ligand to the CCR2. As discussed below various methods can be used to assess inhibition of binding of a ligand to CCR2 and/or function associated with binding of the ligand to the receptor.

Binding Assays

As used herein "mammalian CCR2 protein" refers to naturally occurring or endogenous mammalian CCR2 proteins and to proteins having an amino acid sequence which is the same as that of a naturally occurring or endogenous corresponding mammalian

CCR2 protein (e.g., recombinant proteins). Accordingly, as defined herein, the term includes mature receptor protein, polymorphic or allelic variants, and other isoforms of a mammalian CCR2 (e.g., produced by alternative splicing or other cellular processes), and modified or unmodified forms of the foregoing (e.g., glycosylated, unglycosylated). Mammalian CCR2 proteins can be isolated and/or recombinant proteins (including synthetically produced proteins). Naturally occurring or endogenous mammalian CCR2 proteins include wild type proteins such as mature CCR2, polymorphic or allelic variants and other isoforms which occur naturally in mammals (e.g., humans, non-human primates), such as the CCR2a and CCR2b forms of the receptor protein which are produced by alternative splicing of the carboxy-terminus of the protein. Such proteins can be recovered or isolated from a source which naturally produces mammalian CCR2, for example. These proteins and mammalian CCR2 proteins having the same amino acid sequence as a naturally occurring or endogenous corresponding mammalian CCR2, are referred to by the name of the corresponding mammal. For example, where the corresponding mammal is a human, the protein is designated as a human CCR2 protein (e.g., a recombinant human CCR2 produced in a suitable host cell).

"Functional variants" of mammalian CCR2 proteins include functional fragments, functional mutant proteins, and/or functional fusion proteins (e.g., produced via mutagenesis and/or recombinant techniques). Generally, fragments or portions of mammalian CCR2 proteins include those having a deletion (i.e., one or more deletions) of an amino acid (i.e., one or more amino acids) relative to the mature mammalian CCR2 protein (such as N-terminal, C-terminal or internal deletions). Fragments or portions in which only contiguous amino acids have been deleted or in which non-contiguous amino acids have been deleted relative to mature mammalian CCR2 protein are also envisioned.

Generally, mutants of mammalian CCR2 proteins include natural or artificial variants of a mammalian CCR2 protein differing by the addition, deletion and/or substitution of one or more contiguous or non-contiguous amino acid residues (e.g., receptor chimeras). Such mutations can be in a conserved region or nonconserved region (compared to other CXC and/or CC chemokine receptors), extracellular, cytoplasmic, or transmembrane region, for example.

Generally, fusion proteins encompass polypeptides comprising a mammalian CCR2 (e.g., human CCR2) as a first moiety, linked via a peptide bond to a second moiety not

occurring in the mammalian CCR2 as found in nature. Thus, the second moiety can be an amino acid, oligopeptide or polypeptide. The first moiety can be in an N-terminal location, C-terminal location or internal to the fusion protein. In one embodiment, the fusion protein comprises an affinity ligand (e.g., an enzyme, an antigen, epitope tag) as the first moiety, and a second moiety comprising a linker sequence and human CCR2 or a portion thereof.

A "functional fragment or portion", "functional mutant" and/or "functional fusion protein" of a mammalian CCR2 protein refers to an isolated and/or recombinant protein or polypeptide which has at least one function characteristic of a mammalian CCR2 protein as described herein, such as a binding activity, a signaling activity and/or ability to stimulate a cellular response. Preferred functional variants can bind a ligand (i.e., one or more ligands such as MCP-1, MCP-2, MCP-3, MCP-4, MCP-5 and/or eotaxin), and are referred to herein as "ligand binding variants".

In one embodiment, a functional variant of mammalian CCR2 shares at least about 85% sequence identity with said mammalian CCR2, preferably at least about 90% sequence identity, and more preferably at least about 95% sequence identity with said mammalian CCR2. The nucleic acid and amino acid sequences of human CCR2a and CCR2b are described in U.S. Patent No. 5,707,815. Sequence identity can be determined using a suitable program, such as the Blastx program (Version 1.4), using appropriate parameters, such as default parameters. In one embodiment, parameters for Blastx search are scoring matrix BLOSUM62, W=3. In another embodiment, a functional variant comprises a nucleic acid sequence which is different from the naturally-occurring nucleic acid molecule but which, due to the degeneracy of the genetic code, encodes mammalian CCR2 or a portion thereof.

A composition comprising an isolated and/or recombinant mammalian CCR2 or functional variant thereof can be maintained under conditions suitable for binding, the mammalian CCR2 or variant is contacted with an antibody or fragment to be tested, and binding is detected or measured directly or indirectly. In one embodiment, cells which naturally express CCR2 or cells comprising a recombinant nucleic acid sequence which encodes a mammalian CCR2 or variant thereof are used. The cells are maintained under conditions appropriate for expression of receptor. The cells are contacted with an antibody or fragment under conditions suitable for binding (e.g., in a suitable binding buffer), and

binding is detected by standard techniques. To determine binding, the extent of binding can be determined relative to a suitable control (e.g., compared with background determined in the absence of antibody, compared with binding of a second antibody (i.e., a standard), compared with binding of antibody to untransfected cells). A cellular fraction, such as a
5 membrane fraction, containing receptor or liposomes comprising receptor can be used in lieu of whole cells.

In one embodiment, the antibody is labeled with a suitable label (e.g., fluorescent label, isotope label, antigen or epitope label, enzyme label), and binding is determined by detection of the label. In another embodiment, bound antibody can be
10 detected by labeled second antibody. Specificity of binding can be assessed by competition or displacement, for example, using unlabeled antibody or a ligand as competitor.

Binding inhibition assays can also be used to identify antibodies or fragments thereof which bind CCR2 and inhibit binding of another compound such as a ligand (e.g., MCP-1, MCP-2, MCP-3, MCP-4, MCP-5 and/or eotaxin) to CCR2 or a functional variant.
15 For example, a binding assay can be conducted in which a reduction in the binding of a ligand of CCR2 (in the presence of an antibody), as compared to binding of the ligand in the absence of the antibody, is detected or measured. A composition comprising an isolated and/or recombinant mammalian CCR2 or functional variant thereof can be contacted with the ligand and antibody simultaneously, or one after the other, in either order. A reduction in the
20 extent of binding of the ligand in the presence of the antibody, is indicative of inhibition of binding by the antibody. For example, binding of the ligand could be decreased or abolished.

In one embodiment, direct inhibition of the binding of a ligand (e.g., a chemokine such as MCP-1) to a mammalian CCR2 or variant thereof by an antibody or fragment is monitored. For example, the ability of an antibody to inhibit the binding of 125I-
25 labeled MCP-1, 125I-labeled MCP-2, 125I-labeled MCP-3, 125I-labeled MCP-4, 125I-labeled MCP-5, or 125I-labeled eotaxin to mammalian CCR2 can be monitored. Such an assay can be conducted using suitable cells bearing CCR2 or a functional variant thereof, such as isolated blood cells (e.g., T cells, PBMC) or a suitable cell line naturally expressing CCR2, or a cell line containing nucleic acid encoding a mammalian CCR2, or a membrane
30 fraction from said cells, for instance.

Other methods of identifying the presence of an antibody which binds CCR2 are available, such as other suitable binding assays, or methods which monitor events which are triggered by receptor binding, including signaling function and/or stimulation of a cellular response (e.g., leukocyte trafficking).

5 It will be understood that the inhibitory effect of antibodies of the present invention can be assessed in a binding inhibition assay. Competition between antibodies for receptor binding can also be assessed in the method. Antibodies which are identified in this manner can be further assessed to determine whether, subsequent to binding, they act to inhibit other functions of CCR2 and/or to assess their therapeutic utility.

10 Signaling Assays

The binding of a ligand or promoter, such as an agonist, to CCR2 can result in signaling by this G protein-coupled receptor, and the activity of G proteins as well as other intracellular signaling molecules is stimulated. The induction of signaling function by a
15 compound (e.g., an antibody or fragment thereof) can be monitored using any suitable method. Such an assay can be used to identify antibody agonists of CCR2. The inhibitory activity of an antibody or functional fragment thereof can be determined using a ligand or promoter in the assay, and assessing the ability of the antibody to inhibit the activity induced by ligand or promoter.

20 G protein activity, such as hydrolysis of GTP to GDP, or later signaling events triggered by receptor binding, such as induction of rapid and transient increase in the concentration of intracellular (cytosolic) free calcium $[Ca^{2+}]_i$, can be assayed by methods known in the art or other suitable methods (see e.g., Neote, K. et al., Cell, 72: 415-425 1993); Van Riper et al., J. Exp. Med., 177: 851-856 (1993); Dahinden, C.A. et al., J. Exp. Med.,
25 179: 751-756 (1994)).

For example, the functional assay of Sledziewski et al. using hybrid G protein coupled receptors can be used to monitor the ability a ligand or promoter to bind receptor and activate a G protein (Sledziewski et al., U.S. Patent No. 5,284,746, the teachings of which are incorporated herein by reference).

30 Such assays can be performed in the presence of the antibody or fragment thereof to be assessed, and the ability of the antibody or fragment to inhibit the activity

induced by the ligand or promoter is determined using known methods and/or methods described herein.

Chemotaxis and Assays of Cellular Stimulation

5 Chemotaxis assays can also be used to assess the ability of an antibody or functional fragment thereof to block binding of a ligand to mammalian CCR2 or functional variant thereof and/or inhibit function associated with binding of the ligand to the receptor. These assays are based on the functional migration of cells in vitro or in vivo induced by a compound. Chemotaxis can be assessed as described in the Examples, e.g., in an assay
10 utilizing a 96-well chemotaxis plate, or using other art-recognized methods for assessing chemotaxis. For example, the use of an in vitro transendothelial chemotaxis assay is described by Springer et al. (Springer et al., WO 94/20142, published September 15, 1994, the teachings of which are incorporated herein by reference; see also Berman et al., Immunol. Invest. 17: 625-677 (1988)). Migration across endothelium into collagen gels has also been
15 described (Kavanaugh et al., J. Immunol., 146: 4149-4156 (1991)). Stable transfectants of mouse L1-2 pre-B cells or of other suitable host cells capable of chemotaxis can be used in chemotaxis assays, for example.

Generally, chemotaxis assays monitor the directional movement or migration of a suitable cell (such as a leukocyte (e.g., lymphocyte, eosinophil, basophil)) into or
20 through a barrier (e.g., endothelium, a filter), toward increased levels of a compound, from a first surface of the barrier toward an opposite second surface. Membranes or filters provide convenient barriers, such that the directional movement or migration of a suitable cell into or through a filter, toward increased levels of a compound, from a first surface of the filter toward an opposite second surface of the filter, is monitored. In some assays, the membrane
25 is coated with a substance to facilitate adhesion, such as ICAM-1, fibronectin or collagen. Such assays provide an in vitro approximation of leukocyte "homing".

For example, one can detect or measure inhibition of the migration of cells in a suitable container (a containing means), from a first chamber into or through a microporous membrane into a second chamber which contains an antibody to be tested, and which is
30 divided from the first chamber by the membrane. A suitable membrane, having a suitable pore size for monitoring specific migration in response to compound, including, for example,

nitrocellulose, polycarbonate, is selected. For example, pore sizes of about 3-8 microns, and preferably about 5-8 microns can be used. Pore size can be uniform on a filter or within a range of suitable pore sizes.

To assess migration and inhibition of migration, the distance of migration into the filter, the number of cells crossing the filter that remain adherent to the second surface of the filter, and/or the number of cells that accumulate in the second chamber can be determined using standard techniques (e.g., microscopy). In one embodiment, the cells are labeled with a detectable label (e.g., radioisotope, fluorescent label, antigen or epitope label), and migration can be assessed in the presence and absence of the antibody or fragment by determining the presence of the label adherent to the membrane and/or present in the second chamber using an appropriate method (e.g., by detecting radioactivity, fluorescence, immunoassay). The extent of migration induced by an antibody agonist can be determined relative to a suitable control (e.g., compared to background migration determined in the absence of the antibody, compared to the extent of migration induced by a second compound (i.e., a standard), compared with migration of untransfected cells induced by the antibody).

In one embodiment, particularly for T cells, monocytes or cells expressing a mammalian CCR2, transendothelial migration can be monitored. In this embodiment, transmigration through an endothelial cell layer is assessed. To prepare the cell layer, endothelial cells can be cultured on a microporous filter or membrane, optionally coated with a substance such as collagen, fibronectin, or other extracellular matrix proteins, to facilitate the attachment of endothelial cells. Preferably, endothelial cells are cultured until a confluent monolayer is formed. A variety of mammalian endothelial cells can be available for monolayer formation, including for example, vein, artery or microvascular endothelium, such as human umbilical vein endothelial cells (Clonetics Corp, San Diego, CA). To assay chemotaxis in response to a particular mammalian receptor, endothelial cells of the same mammal are preferred; however endothelial cells from a heterologous mammalian species or genus can also be used.

Generally, the assay is performed by detecting the directional migration of cells into or through a membrane or filter, in a direction toward increased levels of a compound, from a first surface of the filter toward an opposite second surface of the filter, wherein the filter contains an endothelial cell layer on a first surface. Directional migration

occurs from the area adjacent to the first surface, into or through the membrane, towards a compound situated on the opposite side of the filter. The concentration of compound present in the area adjacent to the second surface, is greater than that in the area adjacent to the first surface.

5 In one embodiment used to test for an antibody inhibitor, a composition comprising cells capable of migration and expressing a mammalian CCR2 receptor can be placed in the first chamber. A composition comprising one or more ligands or promoters capable of inducing chemotaxis of the cells in the first chamber (having chemoattractant function) is placed in the second chamber. Preferably shortly before the cells are placed in
10 the first chamber, or simultaneously with the cells, a composition comprising the antibody to be tested is placed, preferably, in the first chamber. Antibodies or functional fragments thereof which can bind receptor and inhibit the induction of chemotaxis, by a ligand or promoter, of the cells expressing a mammalian CCR2 in this assay are inhibitors of receptor function (e.g., inhibitors of stimulatory function). A reduction in the extent of migration
15 induced by the ligand or promoter in the presence of the antibody or fragment is indicative of inhibitory activity. Separate binding studies (see above) could be performed to determine whether inhibition is a result of binding of the antibody to receptor or occurs via a different mechanism.

In vivo assays which monitor leukocyte infiltration of a tissue, in response to
20 injection of a compound (e.g., chemokine or antibody) in the tissue, are described below (see Models of Inflammation). These models of *in vivo* homing measure the ability of cells to respond to a ligand or promoter by emigration and chemotaxis to a site of inflammation and to assess the ability of an antibody or fragment thereof to block this emigration.

In addition to the methods described, the effects of an antibody or fragment on
25 the stimulatory function of CCR2 can be assessed by monitoring cellular responses induced by active receptor, using suitable host cells containing receptor.

Identification of Additional Ligands, Inhibitors and/or Promoters of Mammalian CCR2 Function

30 The assays described above, which can be used to assess binding and function of the antibodies and fragments of the present invention, can be adapted to identify additional

ligands or other substances which bind a mammalian CCR2 or functional variant thereof, as well as inhibitors and/or promoters of mammalian CCR2 function. For example, agents having the same or a similar binding specificity as that of an antibody of the present invention or functional portion thereof can be identified by a competition assay with said antibody or portion thereof. Thus, the present invention also encompasses methods of identifying ligands of the receptor or other substances which bind a mammalian CCR2 protein, as well as inhibitors (e.g., antagonists) or promoters (e.g., agonists) of receptor function. In one embodiment, cells bearing a mammalian CCR2 protein or functional variant thereof (e.g., leukocytes, cell lines or suitable host cells which have been engineered to express a mammalian CCR2 protein or functional variant encoded by a nucleic acid introduced into said cells) are used in an assay to identify and assess the efficacy of ligands or other substances which bind receptor, including inhibitors or promoters of receptor function. Such cells are also useful in assessing the function of the expressed receptor protein or polypeptide.

According to the present invention, ligands and other substances which bind receptor, inhibitors and promoters of receptor function can be identified in a suitable assay, and further assessed for therapeutic effect. Inhibitors of receptor function can be used to inhibit (reduce or prevent) receptor activity, and ligands and/or promoters can be used to induce (trigger or enhance) normal receptor function where indicated. Thus, the present invention provides a method of treating inflammatory diseases, including autoimmune disease and graft rejection, comprising administering an inhibitor of receptor function to an individual (e.g., a mammal). The present invention further provides a method of stimulating receptor function by administering a novel ligand or promoter of receptor function to an individual, providing a new approach to selective stimulation of leukocyte function, which is useful, for example, in the treatment of infectious diseases and cancer.

As used herein, a "ligand" of a mammalian CCR2 protein refers to a particular class of substances which bind to a mammalian CCR2 protein, including natural ligands and synthetic and/or recombinant forms of natural ligands. Infectious agents having a tropism for mammalian CCR2-positive cells (e.g., viruses such as HIV) can also bind to a mammalian CCR2 protein. A natural ligand of a selected mammalian receptor is of a mammalian origin which is the same as that of the mammalian CCR2 protein (e.g., a chemokine such as MCP-

1, MCP-2, MCP-3, MCP-4, MCP-5 and/or eotaxin). In a preferred embodiment, ligand binding of a mammalian CCR2 protein occurs with high affinity.

As used herein, an "inhibitor" is a substance which inhibits (decreases or prevents) at least one function characteristic of a mammalian CCR2 protein (e.g., a human CCR2), such as a binding activity (e.g., ligand binding, promoter binding, antibody binding), a signaling activity (e.g., activation of a mammalian G protein, induction of rapid and transient increase in the concentration of cytosolic free calcium $[Ca^{2+}]_i$), and/or cellular response function (e.g., stimulation of chemotaxis, exocytosis or inflammatory mediator release by leukocytes). An inhibitor is also a substance which inhibits HIV entry into a cell. The term inhibitor refers to substances including antagonists which bind receptor (e.g., an antibody, a mutant of a natural ligand, small molecular weight organic molecules, other competitive inhibitors of ligand binding), and substances which inhibit receptor function without binding thereto (e.g., an anti-idiotypic antibody).

As used herein, a "promoter" is a substance which promotes (induces, causes, enhances or increases) at least one function characteristic of a mammalian CCR2 protein (e.g., a human CCR2), such as a binding activity (e.g., ligand, inhibitor and/or promoter binding), a signaling activity (e.g., activation of a mammalian G protein, induction of rapid and transient increase in the concentration of cytosolic free calcium $[Ca^{2+}]_i$), and/or a cellular response function (e.g., stimulation of chemotaxis, exocytosis or inflammatory mediator release by leukocytes). The term promoter refers to substances including agonists which bind receptor (e.g., an antibody, a homolog of a natural ligand from another species), and substances which promote receptor function without binding thereto (e.g., by activating an associated protein). In a preferred embodiment, the agonist is other than a homolog of a natural ligand.

Thus, the invention also relates to a method of detecting or identifying an agent which binds a mammalian CC-chemokine receptor 2 or ligand binding variant thereof, including ligands, inhibitors, promoters, and other substances which bind a mammalian CCR2 receptor or functional variant. According to the method, an agent to be tested, an antibody or antigen-binding fragment of the present invention (e.g., 8G2, 1D9, an antibody having an epitopic specificity which is the same as or similar to that of 8G2 or 1D9, and antigen-binding fragments thereof) and a composition comprising a mammalian CC-

chemokine receptor 2 or a ligand binding variant thereof can be combined. The foregoing components are combined under conditions suitable for binding of the antibody or antigen-binding fragment to mammalian CC-chemokine receptor 2 or a ligand binding variant thereof, and binding of the antibody or fragment to the mammalian CC-chemokine receptor 2 or ligand binding variant is detected or measured, either directly or indirectly, according to methods described herein or other suitable methods. A decrease in the amount of complex formed relative to a suitable control (e.g., in the absence of the agent to be tested) is indicative that the agent binds said receptor or variant. The composition comprising a mammalian CC-chemokine receptor 2 or a ligand binding variant thereof can be a membrane fraction of a cell bearing recombinant chemokine receptor 2 protein or ligand binding variant thereof. The antibody or fragment thereof can be labeled with a label such as a radioisotope, spin label, antigen or epitope label, enzyme label, fluorescent group and chemiluminescent group.

In one embodiment, the invention relates to a method of detecting or identifying an agent which binds a mammalian CC-chemokine receptor 2 or a ligand binding variant thereof, comprising combining an agent to be tested, an antibody or antigen-binding fragment of the present invention (e.g., 1D9, 8G2, an antibody having an epitopic specificity which is the same as or similar to that of 1D9 or 8G2, or antigen-binding fragments thereof) and a cell bearing a mammalian CC-chemokine receptor 2 or a ligand binding variant thereof. The foregoing components are combined under conditions suitable for binding of the antibody or antigen-binding fragment to the CCR2 protein or ligand binding variant thereof, and binding of the antibody or fragment to the mammalian CC-chemokine receptor 2 or variant is detected or measured, either directly or indirectly, by methods described herein and or other suitable methods. A decrease in the amount of complex formed relative to a suitable control is indicative that the agent binds the receptor or variant. The antibody or fragment thereof can be labeled with a label selected from the group consisting of a radioisotope, spin label, antigen or epitope label, enzyme label, fluorescent group and chemiluminescent group. These and similar assays can be used to detect agents, including ligands (e.g., chemokines or strains of HIV which interact with CCR2) or other substances, including inhibitors or promoters of receptor function, which can bind CCR2 and compete with the antibodies described herein for binding to the receptor.

The assays described above can be used, alone or in combination with each other or other suitable methods, to identify ligands or other substances which bind a mammalian CCR2 protein, and inhibitors or promoters of a mammalian CCR2 protein or variant. The in vitro methods of the present invention can be adapted for high-throughput screening in which large numbers of samples are processed (e.g., a 96-well format). Cells expressing mammalian CCR2 (e.g., human CCR2) at levels suitable for high-throughput screening can be used, and thus, are particularly valuable in the identification and/or isolation of ligands or other substances which bind receptor, and inhibitors or promoters of mammalian CCR2 proteins. Expression of receptor can be monitored in a variety of ways. For instance, expression can be monitored using antibodies of the present invention which bind receptor or a portion thereof. Also, commercially available antibodies can be used to detect expression of an antigen- or epitope-tagged fusion protein comprising a receptor protein or polypeptide (e.g., FLAG tagged receptors), and cells expressing the desired level can be selected.

Nucleic acid encoding a mammalian CCR2 protein or functional variant thereof can be incorporated into an expression system to produce a receptor protein or polypeptide. An isolated and/or recombinant mammalian CCR2 protein or variant, such as a receptor expressed in cells stably or transiently transfected with a construct comprising a recombinant nucleic acid encoding a mammalian CCR2 protein or variant, or in a cell fraction containing receptor (e.g., a membrane fraction from transfected cells, liposomes incorporating receptor), can be used in tests for receptor function. The receptor can be further purified if desired. Testing of receptor function can be carried out in vitro or in vivo.

An isolated and/or recombinant mammalian CCR2 protein or functional variant thereof, such as a human CCR2, can be used in the present method, in which the effect of a compound is assessed by monitoring receptor function as described herein or using other suitable techniques. For example, stable or transient transfectants (e.g., baculovirus infected Sf9 cells, stable transfectants of mouse L1/2 pre-B cells), can be used in binding assays. Stable transfectants of Jurkat cells or of other suitable cells capable of chemotaxis can be used (e.g., mouse L1/2 pre-B cells) in chemotaxis assays, for example.

According to the method of the present invention, compounds can be individually screened or one or more compounds can be tested simultaneously according to

the methods herein. Where a mixture of compounds is tested, the compounds selected by the processes described can be separated (as appropriate) and identified by suitable methods (e.g., PCR, sequencing, chromatography, mass spectroscopy). The presence of one or more compounds (e.g., a ligand, inhibitor, promoter) in a test sample can also be determined according to these methods.

Large combinatorial libraries of compounds (e.g., organic compounds, recombinant or synthetic peptides, "peptoids", nucleic acids) produced by combinatorial chemical synthesis or other methods can be tested (see e.g., Zuckerman, R.N. et al., J. Med. Chem., 37: 2678-2685 (1994) and references cited therein; see also, Ohlmeyer, M.H.J. et al., Proc. Natl. Acad. Sci. USA 90:10922-10926 (1993) and DeWitt, S.H. et al., Proc. Natl. Acad. Sci. USA 90:6909-6913 (1993), relating to tagged compounds; Rutter, W.J. et al. U.S. Patent No. 5,010,175; Huebner, V.D. et al., U.S. Patent No. 5,182,366; and Geysen, H.M., U.S. Patent No. 4,833,092). Where compounds selected from a combinatorial library by the present method carry unique tags, identification of individual compounds by chromatographic methods is possible.

In one embodiment, phage display methodology is used. For example, a mammalian CCR2 protein or functional variant, an antibody or functional portion thereof of the present invention, and a phage (e.g., a phage or collection of phage such as a library) displaying a polypeptide, can be combined under conditions appropriate for binding of the antibody or portion thereof to the mammalian CCR2 protein or variant (e.g., in a suitable binding buffer). Phage which can compete with the antibody or portion thereof and bind to the mammalian CCR2 protein or variant can be detected or selected using standard techniques or other suitable methods. Bound phage can be separated from receptor using a suitable elution buffer. For example, a change in the ionic strength or pH can lead to a release of phage. Alternatively, the elution buffer can comprise a release component or components designed to disrupt binding of compounds (e.g., one or more compounds which can disrupt binding of the displayed peptide to the receptor, such as a ligand, inhibitor, and/or promoter which competitively inhibits binding). Optionally, the selection process can be repeated or another selection step can be used to further enrich for phage which bind receptor. The displayed polypeptide can be characterized (e.g., by sequencing phage DNA). The polypeptides identified can be produced and further tested for binding, and for inhibitor

or promoter function. Analogs of such peptides can be produced which will have increased stability or other desirable properties.

In one embodiment, phage expressing and displaying fusion proteins comprising a coat protein with an N-terminal peptide encoded by random sequence nucleic acids can be produced. Suitable host cells expressing a mammalian CCR2 protein or variant and an anti-CCR2 antibody or functional portion thereof, are combined with the phage, bound phage are selected, recovered and characterized. (See e.g., Doorbar, J. and G. Winter, J. Mol. Biol., 244: 361 (1994) discussing a phage display procedure used with a G protein-coupled receptor).

Other sources of potential ligands or other substances which bind to, or inhibitors and/or promoters of, mammalian CCR2 proteins include, but are not limited to, variants of CCR2 ligands, including naturally occurring, synthetic or recombinant variants of MCP-1, MCP-2, MCP-3, MCP-4, MCP-5 and/or eotaxin, substances such as other chemoattractants or chemokines, variants thereof, low molecular weight organic molecules, other inhibitors and/or promoters (e.g., anti-CCR2 antibodies, antagonists, agonists), other G protein-coupled receptor ligands, inhibitors and/or promoters (e.g., antagonists or agonists), and soluble portions of a mammalian CCR2 receptor, such as a suitable receptor peptide or analog which can inhibit receptor function (see e.g., Murphy, R.B., WO 94/05695).

Models of Inflammation

In vivo models of inflammation are available which can be used to assess the effects of antibodies and fragments of the invention *in vivo* as therapeutic agents. For example, leukocyte infiltration upon intradermal injection of a chemokine and an antibody or fragment thereof reactive with mammalian CCR2 into a suitable animal, such as rabbit, mouse, rat, guinea pig or rhesus macaque can be monitored (see e.g., Van Damme, J. et al., J. Exp. Med., 176: 59-65 (1992); Zachariae, C.O.C. et al., J. Exp. Med. 171: 2177-2182 (1990); Jose, P.J. et al., J. Exp. Med. 179: 881-887 (1994)). In one embodiment, skin biopsies are assessed histologically for infiltration of leukocytes (e.g., eosinophils, granulocytes). In another embodiment, labeled cells (e.g., stably transfected cells expressing a mammalian CCR2, labeled with ¹¹¹In for example) capable of chemotaxis and extravasation are

administered to the animal. For example, an antibody or fragment to be assessed can be administered, either before, simultaneously with or after ligand or agonist is administered to the test animal. A decrease of the extent of infiltration in the presence of antibody as compared with the extent of infiltration in the absence of inhibitor is indicative of inhibition.

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Diagnostic and Therapeutic Applications

The antibodies and fragments of the present invention are useful in a variety of applications, including research, diagnostic and therapeutic applications. In one embodiment, the antibodies are labeled with a suitable label (e.g., fluorescent label, chemiluminescent label, isotope label, antigen or epitope label or enzyme label). For instance, they can be used to isolate and/or purify receptor or portions thereof, and to study receptor structure (e.g., conformation) and function.

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In addition, the various antibodies of the present invention can be used to detect CCR2 or to measure the expression of receptor, for example, on T cells (e.g., CD8+ cells, CD45RO+ cells), monocytes and/or on cells transfected with a receptor gene. Thus, they also have utility in applications such as cell sorting (e.g., flow cytometry, fluorescence activated cell sorting), for diagnostic or research purposes.

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The anti-CCR2 antibodies of the present invention have value in diagnostic applications. An anti-CCR2 antibody or fragment thereof can be used to monitor expression of this receptor in HIV infected individuals, similar to the way anti-CD4 has been used as a diagnostic indicator of disease stage.

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Typically, diagnostic assays entail detecting the formation of a complex resulting from the binding of an antibody or fragment thereof to CCR2. For diagnostic purposes, the antibodies or antigen-binding fragments can be labeled or unlabeled. The antibodies or fragments can be directly labeled. A variety of labels can be employed, including, but not limited to, radionuclides, fluorescers, enzymes, enzyme substrates, enzyme cofactors, enzyme inhibitors and ligands (e.g., biotin, haptens). Numerous appropriate immunoassays are known to the skilled artisan (see, for example, U.S. Patent Nos. 3,817,827; 3,850,752; 3,901,654 and 4,098,876). When unlabeled, the antibodies or fragments can be detected using suitable means, as in agglutination assays, for example. Unlabeled antibodies or fragments can also be used in combination with another (i.e., one or more) suitable reagent

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which can be used to detect antibody, such as a labeled antibody (e.g., a second antibody) reactive with the first antibody (e.g., anti-idiotypic antibodies or other antibodies that are specific for the unlabeled immunoglobulin) or other suitable reagent (e.g., labeled protein A).

In one embodiment, the antibodies or fragments of the present invention can be utilized in enzyme immunoassays, wherein the subject antibody or fragment, or second antibodies, are conjugated to an enzyme. When a biological sample comprising a mammalian CCR2 protein is combined with the subject antibodies, binding occurs between the antibodies and CCR2 protein. In one embodiment, a sample containing cells expressing a mammalian CCR2 protein, such as human blood, is combined with the subject antibodies, and binding occurs between the antibodies and cells bearing a human CCR2 protein comprising an epitope recognized by the antibody. These bound cells can be separated from unbound reagents and the presence of the antibody-enzyme conjugate specifically bound to the cells can be determined, for example, by contacting the sample with a substrate of the enzyme which produces a color or other detectable change when acted on by the enzyme. In another embodiment, the subject antibodies can be unlabeled, and a second, labeled antibody can be added which recognizes the subject antibody.

Kits for use in detecting the presence of a mammalian CCR2 protein in a biological sample can also be prepared. Such kits will include an antibody or functional fragment thereof which binds to a mammalian CC-chemokine receptor 2 or portion of said receptor, as well as one or more ancillary reagents suitable for detecting the presence of a complex between the antibody or fragment and CCR2 or portion thereof. The antibody compositions of the present invention can be provided in lyophilized form, either alone or in combination with additional antibodies specific for other epitopes. The antibodies, which can be labeled or unlabeled, can be included in the kits with adjunct ingredients (e.g., buffers, such as Tris, phosphate and carbonate, stabilizers, excipients, biocides and/or inert proteins, e.g., bovine serum albumin). For example, the antibodies can be provided as a lyophilized mixture with the adjunct ingredients, or the adjunct ingredients can be separately provided for combination by the user. Generally these adjunct materials will be present in less than about 5% weight based on the amount of active antibody, and usually will be present in a total amount of at least about 0.001% weight based on antibody concentration. Where a second antibody capable of binding to the monoclonal antibody is employed, such antibody

can be provided in the kit, for instance in a separate vial or container. The second antibody, if present, is typically labeled, and can be formulated in an analogous manner with the antibody formulations described above.

Similarly, the present invention also relates to a method of detecting and/or
5 quantitating expression of a mammalian CCR2 or a portion of the receptor by a cell, in which a composition comprising a cell or fraction thereof (e.g., membrane fraction) is contacted with an antibody or functional fragment thereof (e.g., 1D9 and/or 8G2) which binds to a mammalian CCR2 or portion of the receptor under conditions appropriate for binding of the antibody or fragment thereto, and binding is monitored. Detection of the antibody, indicative
10 of the formation of a complex between antibody and CCR2 or a portion thereof, indicates the presence of the receptor. Binding of antibody to the cell can be determined as described above under the heading "Binding Assays", for example. The method can be used to detect expression of CCR2 on cells from an individual (e.g., in a sample, such as a body fluid, such as blood, saliva or other suitable sample). The level of expression of CCR2 on the surface of
15 T cells or monocytes can also be determined, for instance, by flow cytometry, and the level of expression (e.g., staining intensity) can be correlated with disease susceptibility, progression or risk.

Chemokine receptors function in the migration of leukocytes throughout the body, particularly to inflammatory sites. Inflammatory cell emigration from the vasculature
20 is regulated by a three-step process involving interactions of leukocyte and endothelial cell adhesion proteins and cell specific chemoattractants and activating factors (Springer, T.A., Cell, 76:301-314 (1994); Butcher, E.C., Cell, 67:1033-1036 (1991); Butcher, E.C. and Picker, L.J., Science (Wash. DC), 272:60-66 (1996)). These are: (a) a low affinity interaction between leukocyte selectins and endothelial cell carbohydrates; (b) a high-affinity
25 interaction between leukocyte chemoattractant receptors and chemoattractant/activating factors; and (c) a tight-binding between leukocyte integrins and endothelial cell adhesion proteins of the immunoglobulin superfamily. Different leukocyte subsets express different repertoires of selectins, chemoattractant receptors and integrins. Additionally, inflammation alters the expression of endothelial adhesion proteins and the expression of chemoattractant
30 and leukocyte activating factors. As a consequence, there is a great deal of diversity for regulating the selectivity of leukocyte recruitment to extravascular sites. The second step is

crucial in that the activation of the leukocyte chemoattractant receptors is thought to cause the transition from the selectin-mediated cell rolling to the integrin-mediated tight binding. This results in the leukocyte being ready to transmigrate to perivascular sites. The chemoattractant/chemoattractant receptor interaction is also crucial for transendothelial migration and localization within a tissue (Campbell, J.J., et al., J. Cell Biol., 134:255-266 (1996); Carr, M.W., et al., Immunity, 4:179-187 (1996)). This migration is directed by a concentration gradient of chemoattractant leading towards the inflammatory focus.

CCR2 has an important role in leukocyte trafficking. It is likely that CCR2 is a key chemokine receptor for T cell or T cell subset or monocyte migration to certain inflammatory sites, and so anti-CCR2 mAbs can be used to inhibit (reduce or prevent) T cell or monocyte migration, particularly that associated with T cell dysfunction, such as autoimmune disease, or allergic reactions or with monocyte-mediated disorders such as atherosclerosis. Accordingly, the antibodies and fragments thereof of the present invention can also be used to modulate receptor function in research and therapeutic applications. For instance, the antibodies and functional fragments described herein can act as inhibitors to inhibit (reduce or prevent) (a) binding (e.g., of a ligand, an inhibitor or a promoter) to the receptor, (b) a receptor signaling function, and/or (c) a stimulatory function. Antibodies which act as inhibitors of receptor function can block ligand or promoter binding directly or indirectly (e.g., by causing a conformational change). For example, antibodies can inhibit receptor function by inhibiting binding of a ligand, or by desensitization (with or without inhibition of binding of a ligand). Antibodies which bind receptor can also act as agonists of receptor function, triggering or stimulating a receptor function, such as a signaling and/or a stimulatory function of a receptor (e.g., leukocyte trafficking) upon binding to receptor.

Thus, the present invention provides a method of inhibiting leukocyte trafficking in a mammal (e.g., a human patient), comprising administering to the mammal an effective amount of an antibody or functional fragment of the present invention. Administration of an antibody or fragment of the present invention can result in amelioration or elimination of the disease state.

The antibody of the present invention, or a functional fragment thereof, can also be used to treat disorders in which activation of the CCR2 receptor by binding of

chemokines is implicated. For example, the antibodies or functional fragments thereof (e.g., 1D9 and/or 8G2 or functional fragments thereof) can be used to treat allergy, atherogenesis, anaphylaxis, malignancy, chronic and acute inflammation, histamine and IgE-mediated allergic reactions, shock, and rheumatoid arthritis, atherosclerosis, multiple sclerosis, stenosis, restenosis, allograft rejection, fibrotic disease, asthma, and inflammatory glomerulopathies.

Diseases or conditions of humans or other species which can be treated with inhibitors of CCR2 receptor function (including antibodies or suitable fragments thereof), include, but are not limited to:

- inflammatory or allergic diseases and conditions, including respiratory allergic diseases such as asthma, allergic rhinitis, hypersensitivity lung diseases, hypersensitivity pneumonitis, interstitial lung diseases (ILD) (e.g., idiopathic pulmonary fibrosis, or ILD associated with rheumatoid arthritis, systemic lupus erythematosus, ankylosing spondylitis, systemic sclerosis, Sjogren's syndrome, polymyositis or dermatomyositis); chronic obstructive pulmonary disease, e.g., emphysema and chronic bronchitis; anaphylaxis or hypersensitivity responses, drug allergies (e.g., to penicillin, cephalosporins), insect sting allergies; inflammatory bowel diseases, such as Crohn's disease and ulcerative colitis; spondyloarthropathies; scleroderma (e.g., systemic scleroderma) and mixed connective tissue disorders; psoriasis and inflammatory dermatoses such as dermatitis, eczema, atopic dermatitis, allergic contact dermatitis, urticaria; vasculitis (e.g., necrotizing, cutaneous, and hypersensitivity vasculitis);

- autoimmune diseases, such as arthritis (e.g., rheumatoid arthritis, juvenile rheumatoid arthritis, psoriatic arthritis, arthritis associated with gastrointestinal disease, and reactive arthritis), multiple sclerosis and other demyelinating diseases, systemic lupus erythematosus (SLE), myasthenia gravis, juvenile onset diabetes, ankylosing spondylitis, undifferentiated spondyloarthropathies and juvenile onset, spondyloarthropathies (including juvenile rheumatoid arthritis), nephritides such as glomerulonephritis, autoimmune thyroiditis, Behcet's disease;

- graft rejection (e.g., in transplantation), including allograft rejection, xenograft rejection or graft-versus-host disease, and organ transplant-associated arteriosclerosis;

- atherosclerosis;

- cancers with leukocyte infiltration of the skin or organs, CCR2 expressing cancers, e.g., solid tumors (e.g., breast cancer) or liquid tumors, CCR2 ligand expressing cancers, e.g., solid or liquid tumors, and cancers that have aberrant or abnormal expression of CCR2;

- stenosis or restenosis of the vasculature, particularly of the arteries, e.g., the coronary artery, such as stenosis or restenosis which results from vascular intervention (e.g., surgical, therapeutic or mechanical intervention), as well as neointimal hyperplasia. For example, restenosis, which typically produces a narrowing of the luminal opening of the vessel, can result from vascular injury including, but not limited to, that produced by vascular graft procedures, angioplasty, including angioplasty performed by balloon, atherectomy, laser or other suitable method (e.g., percutaneous transluminal coronary angioplasty (PTCA)), stent placement (e.g., mechanical or biological endovascular stent placement), vascular bypass procedures or combinations thereof, as well as other procedures used to treat stenotic or occluded blood vessels;

- fibrotic disorders such as hepatic fibrosis (e.g., hepatic fibrosis associated with alcohol induced liver disease, other toxins and drugs, chronic hepatitis, acute hepatitis, environmental insults and autoimmune hepatitis), fibrotic disorders of the lung, kidney or heart, and inflammatory spondyloarthropathies, e.g., ankylosing spondylitis;

- ischemic disorders, e.g., ischemic heart disease (e.g., myocardial infarction, unstable angina, angina, ischemic cardiomyopathies, asymptomatic (silent) angina);

- cerebrovascular disease including ischemic CNS diseases, ischemic stroke, transient ischemic attacks (TAI);

- ischemia reperfusion injury, e.g., of organ systems, surgical conditions such as vascular bypass grafting (e.g., CABG, peripheral bypass grafting), ischemic renal disease, renal artery stenosis with or without subsequent ischemia and atheroembolic disease of the renal artery(ies), and ischemic colitis.

- other diseases or conditions (including CCR2-mediated diseases or conditions), in which undesirable inflammatory responses are to be inhibited can be treated, including, but not limited to, reperfusion injury such as ischemia reperfusion, certain

hematologic malignancies, cytokine-induced toxicity (e.g., septic shock, endotoxic shock), polymyositis, dermatomyositis, and granulomatous diseases including sarcoidosis.

Diseases or conditions of humans or other species which can be treated with promoters of CCR2 receptor function (including antibodies or fragments thereof), include, but are not limited to:

- immunosuppression, such as that in individuals with immunodeficiency syndromes such as AIDS, individuals undergoing radiation therapy, chemotherapy, therapy for autoimmune disease or other drug therapy (e.g., corticosteroid therapy), which causes immunosuppression; and immunosuppression due congenital deficiency in receptor function or other causes.

Anti-CCR2 antibodies of the present invention can block the binding of one or more chemokines, thereby blocking the downstream cascade of one or more events leading to the above disorders.

Antibodies and functional fragments thereof which are antagonists of CCR2 can be used as therapeutics for AIDS, as well as certain inflammatory diseases. HIV-1 and HIV-2 are the etiologic agents of acquired immunodeficiency syndrome (AIDS) in humans. AIDS results in part from the depletion of CD4⁺ T lymphocytes in HIV infected individuals. HIV-1 infects primarily T lymphocytes, monocytes/macrophages, dendritic cells and, in the central nervous system, microglia. All of these cells express the CD4 glycoprotein, which serves as a receptor for HIV-1 and HIV-2. Efficient entry of HIV into target cells is dependent upon binding of the viral exterior envelope glycoprotein, gp120, to the amino-terminal CD4 domain. After virus binding, the HIV-1 envelope glycoproteins mediate the fusion of viral and host cell membranes to complete the entry process. Membrane fusion directed by HIV-1 envelope glycoproteins expressed on the infected cell surface leads to cell-cell fusion, resulting in syncytia.

Recently, host cell factors in addition to CD4 have been suggested to determine the efficiency of HIV-1 envelope glycoprotein-mediated membrane fusion. The 7 transmembrane receptor (7TMR) termed HUMSTR, LESTR, or "fusin" has been shown to allow a range of CD4-expressing cells to support infection and cell fusion mediated by laboratory-adapted HIV-1 envelope glycoproteins (Feng, Y., et al., Science (Wash. DC), 272:872-877 (1996)). Antibodies to HUMSTR blocked cell fusion and infection by

laboratory-adapted HIV-1 isolates but not by macrophage-tropic primary viruses in vitro (Feng, Y., et al., Science (Wash. DC), 272:872-877 (1996)).

The ability of chemokine receptors and related molecules to facilitate the infection of primary clinical HIV-1 isolates has been reported recently by several groups (see e.g., Bates, P., Cell, 86:1-3 (1996); Choe, H., et al., Cell, 85:1135-1148 (1996); Doranz et al., Cell 85:1149-1158 (1996)). These studies indicated that involvement of various members of the chemokine receptor family in the early stages of HIV-1 infection helps to explain viral tropism and β -chemokine inhibition of primary HIV-1 isolates.

The present invention also provides a method of inhibiting HIV infection of a cell (e.g., new infection and/or syncytium formation) which expresses a mammalian CCR2 or portion thereof, comprising contacting the cell with a composition comprising an effective amount of an antibody or functional fragment thereof which binds to a mammalian CCR2 or portion of said receptor. The composition can also comprise one or more additional agents effective against HIV, including, but not limited to, anti-CCR3 antibodies, anti-CCR5 antibodies, and anti-fusin antibodies.

Various methods can be used to assess binding of HIV to a cell and/or infection of a cell by HIV in the presence of the antibodies of the present invention. For example, assays which assess binding of gp120 or a portion thereof to the receptor, HIV infection and syncytium formation can be used (see, for example, Choe, H., et al., Cell, 85:1135-1148 (1996)). The ability of the antibody of the present invention to inhibit these processes can be assessed using these or other suitable methods.

In addition, the present invention provides a method of treating HIV in a patient, comprising administering to the patient a composition comprising an effective amount of an antibody or functional fragment thereof which binds to a mammalian CCR2 or portion of said receptor. Again, the composition can also comprise one or more additional agents effective against HIV, including, but not limited to, anti-CCR3 antibodies, anti-CCR5 antibodies, and anti-fusin antibodies. Therapeutic use of antibody to treat HIV includes prophylactic use (e.g., for treatment of a patient who may be or who may have been exposed to HIV). For example, health care providers who may be exposed or who have been exposed to HIV (e.g., by needle-stick) can be treated according to the method. Another example is

the treatment of a patient exposed to virus after unprotected sexual contact or failure of protection.

In AIDS, multiple drug treatment appears the most promising. An anti-chemokine receptor antagonist that inhibits HIV infection can be added to the drug treatment regimen, in particular by blocking virus infection of new cells. Thus, administration of an antibody or fragment of the present invention in combination with one or more other therapeutic agents such as nucleoside analogues (e.g., AZT, 3TC, ddI) and/or protease inhibitors is envisioned, and provides an important addition to an HIV treatment regimen. In one embodiment, a humanized anti-CCR2 mAb is used in combination with a (i.e., one or more) therapeutic agent to reduce viral load from patients, by preventing fusion and/or infection of new cells. Such an antibody can also be useful in preventing perinatal infection.

Another aspect of the invention relates to a method of preventing HIV infection in an individual, comprising administering to the individual an effective amount of an antibody or functional fragment thereof which binds to CCR2. According to the method, preventing HIV infection includes treatment in order to prevent (reduce or eliminate) infection of new cells in an infected individual or in order to prevent infection in an individual who may be, may have been, or has been, exposed to HIV. For example, individuals such as an HIV infected individual, a fetus of an HIV infected female, or a health care worker may be treated according to the method of the present invention.

Modes of Administration

One or more antibodies or fragments of the present invention can be administered to an individual by an appropriate route, either alone or in combination with (before, simultaneous with, or after) another drug or agent, or before, simultaneous with or after surgical, mechanical or therapeutic intervention. For example, the antibodies of the present invention can also be used in combination with other monoclonal or polyclonal antibodies (e.g., in combination with antibodies which bind other chemokine receptors, including, but not limited to, CCR3 and CCR5) or with existing blood plasma products, such as commercially available gamma globulin and immune globulin products used in prophylactic or therapeutic treatments. The antibodies or fragments of the present invention

can be used as separately administered compositions given in conjunction with antibiotics and/or antimicrobial agents.

An effective amount of an antibody or fragment (i.e., one or more antibodies or fragments) is administered. An effective amount is an amount sufficient to achieve the desired therapeutic (including prophylactic) effect, under the conditions of administration, such as an amount sufficient for inhibition of a CCR2 function, and thereby, inhibition of an inflammatory response or HIV infection, or an amount sufficient for promotion of a CCR2 function, as indicated. The antibody or fragment can be administered in a single dose or multiple doses. The dosage can be determined by methods known in the art and is dependent, for example, upon the antibody or fragment chosen, the subject's age, sensitivity and tolerance to drugs, and overall well-being. Antibodies and antigen-binding fragments thereof, such as human, humanized and chimeric antibodies and antigen-binding fragments can often be administered with less frequency than other types of therapeutics. For example, an effective amount of an antibody can range from about 0.01 mg/kg to about 5 or 10 mg/kg administered daily, weekly, biweekly or monthly.

A variety of routes of administration are possible including, but not necessarily limited to, oral, dietary, topical, parenteral (e.g., intravenous, intraarterial, intramuscular, subcutaneous injection or infusion), inhalation (e.g., intrabronchial, intraocular, intranasal or oral inhalation, intranasal drops), depending on the disease or condition to be treated. Other suitable methods of administration can also include rechargeable or biodegradable devices and slow release polymeric devices. The pharmaceutical compositions of this invention can also be administered as part of a combinatorial therapy with other agents.

Formulation of an antibody or fragment to be administered will vary according to the route of administration and formulation (e.g., solution, emulsion, capsule) selected. An appropriate pharmaceutical composition comprising an antibody or functional fragment thereof to be administered can be prepared in a physiologically acceptable vehicle or carrier. A mixture of antibodies and/or fragments can also be used. For solutions or emulsions, suitable carriers include, for example, aqueous or alcoholic/aqueous solutions, emulsions or suspensions, including saline and buffered media. Parenteral vehicles can include sodium chloride solution, Ringer's dextrose, dextrose and sodium chloride, lactated

Ringer's or fixed oils. A variety of appropriate aqueous carriers are known to the skilled artisan, including water, buffered water, buffered saline, polyols (e.g., glycerol, propylene glycol, liquid polyethylene glycol), dextrose solution and glycine. Intravenous vehicles can include various additives, preservatives, or fluid, nutrient or electrolyte replenishers (See, 5 generally, Remington's Pharmaceutical Science, 16th Edition, Mack, Ed. 1980). The compositions can optionally contain pharmaceutically acceptable auxiliary substances as required to approximate physiological conditions such as pH adjusting and buffering agents and toxicity adjusting agents, for example, sodium acetate, sodium chloride, potassium chloride, calcium chloride and sodium lactate. The antibodies and fragments of this 10 invention can be lyophilized for storage and reconstituted in a suitable carrier prior to use according to art-known lyophilization and reconstitution techniques. The optimum concentration of the active ingredient(s) in the chosen medium can be determined empirically, according to procedures well known to the skilled artisan, and will depend on the ultimate pharmaceutical formulation desired. For inhalation, the antibody or fragment 15 can be solubilized and loaded into a suitable dispenser for administration (e.g., an atomizer, nebulizer or pressurized aerosol dispenser).

The present invention will now be illustrated by the following Examples, which are not intended to be limiting in any way. The teachings of all references cited herein are incorporated herein by reference.

EXAMPLES

Example 1

Materials:

The following materials were obtained from the indicated sources:

25 PE-conjugated anti-CD16, PE-conjugated streptavidin, and biotinylated anti-human IgE were from Pharmingen (San Diego, CA). FITC-conjugated goat anti-mouse IgG was from Jackson ImmunoResearch Laboratories (West Grove, PA).

FACS Lysing Buffer was from Becton Dickenson (Mountain View, CA) and [125I]-MCP-1 was from NEN (Boston, MA).

Cells, Cell Lines, and Tissue Culture

The murine pre-B lymphoma cell line L1/2 was maintained in RPMI-1640 supplemented with 10% Fetal Clone I (Gibco BRL, Gaithersburg, MD) 50 Units/mL penicillin (Gibco BRL), 50 µg/mL streptomycin (Gibco BRL), 2 mM L-Glutamine (Gibco BRL), and 55 µM β-mercaptoethanol (Gibco BRL). Other cell lines included transfectants of L1/2 cells expressing either CCR1 (Campbell, J. et al. (1996) J. Cell Bio., 134:255-266), CCR5 (Wu et al., Nature 384:179-183 (1996)) grown in the above culture medium supplemented with 800 µg/ml active G418. THP-1 cells (ATCC No. TIB202) were grown in accordance with ATCC instructions. PBMC were purified from heparinized blood as described in Ponath et al., J. Clin. Invest., 97:604-612 (1996).

Preparation of CCR2b Expression Construct and Stable Transfectants

The coding region for the human CCR2b (Charo et al. (1994) Proc. Natl. Acad. Sci. USA, 91:2752) was obtained by RT-PCR amplification as described (Qin, S. et al. (1996) Eur. J. Immunol., 26:640-647). cDNA was made using oligo(dT)-priming, and amplification of the CCR2b coding region was achieved by nested PCR with the following sets of primers which correspond to the positions of the CCR2b sequence (GenBank Accession No. U03905; Charo et al., Proc. Natl. Acad. Sci. USA 91:2752-2756 (1994)) as indicated:

1) 5' primer: 5'-TGAGACAAGCCACAAGCTGAAC-3' (nucleotides 11 to 32; SEQ ID NO: 1);

3' Primer: 5'-TCTGTATTAGTACACACAGCCC-3' (nucleotides 1301 to 1280; SEQ ID NO: 2);

2) 5' Primer: 5'-ATGCTGTCCACATCTCGTTCTCGG-3' (nucleotides 81 to 104; SEQ ID NO: 3);

3' Primer: 5'-TTATAAACCAGCCGAGACTTCCTGCTC-3' (nucleotides 1164 to 1137; SEQ ID NO: 4).

The CCR2B cDNA coding region was modified to contain the CD5 signal peptide leader sequence (Aruffo et al., Cell 61:1303-1313 (1990)). The predicted amino acid sequence of this peptide is:

5 NH₂-Met-Pro-Met-Gly-Ser-Leu-Gln-Pro-Leu-Ala-Thr-Leu-Tyr-Leu-Leu-Gly-Met-Leu-Val-Ala-Ser-Val-Leu-Ala... (SEQ ID NO: 5)

10 Using PCR with the CCR2b cDNA as template and two overlapping 5' primers that contain a BamHI restriction site, encode the CD5 signal peptide sequence and the amino terminal sequence of CCR2b, and a 3' primer located internally in the CCR2b coding region.

5' CD5 Seq1 primer

5'-

15 GGGGATCCAGAAACCATGCCCATGGGGTCTCTGCAACCGCTGGCCACCTT
GTACCTGCTG-3' (SEQ ID NO: 6)

5' CD5 Seq2 primer

5'-

20 GCCACCTTGTACCTGCTGGGGATGCTGGTCGCTTCCGTGCTAGCGATGCTGTCCA
CATCTCGTTC-3' (SEQ ID NO: 7)

3' CCR2AB2 primer - 5' - GACGACCAGCATGTTGCC-3' (SEQ ID
NO: 8; U03905 nucleotides 272 to 255)

25

30 The 278 base pair amplified fragment was digested with BamHI and Apal and the resulting 209 base pair fragment was inserted at the Apal site at position 206 of the CCR2b cDNA (GenBank Accession No. U03905) to replace the endogenous 5' base pair fragment of CCR2. The resulting sequence that encodes a CCR2b with the CD5 signal peptide leader sequence immediately preceding the receptor initiator methionine was inserted into the BamHI and XhoI sites of pcDNA3 (Invitrogen, San Diego, CA) to create the

mammalian expression plasmid pCD5MCPRB. The CD5-CCR2b fragment was subcloned into the BamH I-Not I site of pCDEF3 (Goldman et al., (1996) Biotechniques 21:1013-1015), and this construct was designated CCR2bDEF3. In this expression vector, the expression of the inserted gene is driven by the EF-1 α promoter.

5 Fifty milliliters of L1/2 cells were seeded at 4×10^5 cells/mL the day before the electroporation. On the day of the electroporation, the cells, which had grown up to a density of 1×10^6 /mL, were centrifuged out of their medium and resuspended in 800 μ L room temperature electroporation buffer (Zajac et al., DNA 7:509-513). 120 mM L-Glutamic Acid (Sigma), 7 mM Mg Acetate (EM Science), 4.3 mM Glucose (Sigma), 17 mM K Pipes, pH 10 6.9 (Sigma), 1 mM EGTA (Sigma), 5 mM ATP, pH 7.0 (Sigma). Twenty-five micrograms Sca I linearized, phenol/chloroform/isoamyl alcohol extracted and isopropanol precipitated CCR2bDEF3 plasmid DNA was placed in an 0.4 cm gap electroporation curvette. The resuspended cells were added to the curvette, and a single pulse applied at 450 volts, 960 μ Fd. The cells were then transferred from the curvette to a T-75 flask containing 15 mL L1/2 15 growth medium (described above, and grown for three days, at which time the cells were centrifuged out of their medium and resuspended in L1/2 growth medium additionally supplemented with 1 mM sodium pyruvate (Gibco BRL) and 0.8 mg/mL active G418 (Gibco BRL).

20 Selection of Cells Expressing CCR2b by Chemotaxis

The transfected cells were allowed to grow for eleven days, at which point they were split 1:20 into fresh growth medium. On the sixteenth day, the cells were selected by chemotaxis. 600 μ L 1 nM MCP-1 in RPMI 1640 supplemented with 0.5% BSA (RPMI/BSA) was placed in the lower chamber and 1×10^6 CCR2bDEF3 cells in 100 μ L of 25 RPMI/BSA were placed in the upper chamber of a 3.0 micron pore 24-well chemotaxis plate (Becton Dickinson). The cells were allowed to chemotax for four hours and twenty minutes in a 37° C, 5% CO₂, humidified incubator, at which time the upper chamber was removed. This incubation time was chosen at the time of the experiment because it was sufficiently long for cells responding to the MCP-1 to chemotax, but short enough to keep the 30 background low.

Secondary Selection of CCR2b Expressing-Cells by FACS Sorting

The cells which had chemotaxed through the membrane and into the lower chamber were grown up, and further purified by sterile FACS sorting. Ten million CCR2bDEF3 cells were centrifuged out of their medium, resuspended in 2.5 mL PBS(+Ca, Mg) supplemented with 1% heat-inactivated Fetal Calf Serum ("HI FCS") (Gibco BRL) and 2.5 mL sterile filtered anti-CCR2b amino-terminal peptide antibody supernatant 5A11. The cells and the antibody were mixed and allowed to incubate on ice for thirty minutes. The cells were then washed twice with PBS (+) (Gibco BRL), and resuspended in 5 mL of a sterile filtered, 1:250 dilution of FITC-conjugated, affinity-purified F(ab1)2 goat anti-mouse IgG (Jackson ImmunoResearch Laboratories) in PBS (+) supplemented with 1% HI FCS. The cells were incubated for thirty minutes on ice in the dark, and then washed twice with PBS(+) (GIBCO BRL). The cells were sorted on the FACSCalibur® and the brightest 4% of cells were collected. (FL1 > 3 x 10²).

The sorted cells were allowed to grow up, and they were resorted using the same protocol as above. The brightest 1% of cells were collected. (FL1 > 3 x 10³).

Monoclonal Antibody Production

To produce mAbs to CCR2b, transfectants were continually monitored to ensure that levels of expression did not drift downward. FACS staining was performed periodically to ascertain receptor expression on the transfectants using the anti CCR2b antibody supernatant 5A11 with goat anti-mouse IgG FITC as the secondary antibody.

Twenty million CCR2bDEF3.L1/2 cells were washed in RPMI 1640 (Gibco BRL) and incubated in RPMI 1640 plus 0.2 mg/mL Mitomycin C for 30 minutes at 37°C. The cells were then washed twice with PBS (+) and 2 x 10⁷ cells in 0.5 mL PBS (+) were injected intraperitoneally into a C57 BL/6 female mouse. This was repeated two more times at two week intervals. The fourth time, 2 x 10⁷ cells were resuspended in 0.25 mL and injected intravenously. Three days after the intravenous injection, the mouse was sacrificed and the spleen removed and the cells fused with the SP2/0 cell line as described (Current Protocols in Immunology, John Wiley and Sons, New York, 1992).

This set of mice had previously been immunized many times with 2 different cell lines as well as a synthetic peptide, but no antibodies that stained CCR2 positive cells

were generated from several fusions. The above four immunizations with the CCR2bDEF3.L1/2 cell line expressing high levels of CCR2b were critical to obtain the described antibody.

5 Selecting Single Cell Clone of CCR2 Transfectants by Limiting Dilution

After the mouse received the last injection, the twice sorted cells were allowed to grow up again, and then they were further purified by limiting dilution. The cells were plated at 1 and 0.5 cell per well in 96 well plates. Subcloned cells from the 0.5 cell per well dilution were grown up and tested for CCR2b expression by indirect immunofluorescent
10 FACS analysis using the anti-CCR2b antibody supernatant 5A11 with goat anti-mouse IgG FITC as the secondary antibody. The procedure was the same as described above, except that the staining volume was 100 μ l. Four positives were selected and frozen down.

Identification of Positive Monoclonal Antibodies

15 Immunofluorescent staining analysis using a FACScan® (Becton Dickinson & Co., Mountain View, CA) was used to identify the monoclonal antibodies which were reactive with the CCR2b receptor. Hybridoma culture supernatants were assayed in a 96-well format using goat anti-mouse IgG FITC as the secondary antibody. CCR2bDEF3.L1/2 cells were used to identify monoclonal antibodies reactive with CCR2b, and untransfected
20 L1/2 cells were used to eliminate monoclonal antibodies reactive with other cell surface proteins.

FACS Staining - Cultured Cells

For the staining of cultured transfectant cell lines 0.5 x 10⁶ cells in 50 μ l were
25 resuspended in PBS + 1% FCS in a 96 well polystyrene V-bottom plate. 50 μ l of primary antibody supernatants or HT medium (negative control) were added, and the samples were incubated at 4°C for 30 min. 100 μ l of PBS were added and the cells were pelleted by centrifugation and washed once with PBS. The pellet was resuspended in 100 μ l PBS + 1% FCS containing FITC-conjugated goat anti-mouse IgG antibody (a 1:250 dilution) and
30 incubated for thirty minutes at 4°C in the dark. The cells were washed twice with PBS, resuspended in PBS, and analyzed by flow cytometry with a FacScan cytometer using the

CellQuest software (Becton-Dickenson) Cells were fixed with PBS/1% formaldehyde if they were not to be analyzed the same day. Monoclonal antibodies 1D9 and 8G2 stain CCR2 transfectants but not CCR1 or CCR5 transfectants (Figure 1).

5 FACS Staining - Whole Blood

100 µl whole blood was mixed with 100 µL of 1D9 antibody hybridoma supernatants or HT medium (negative control) and incubated at 4°C for 30 min. After one wash with PBS, 100 µL FITC-conjugated goat anti-mouse IgG antibody (a 1:250 dilution) was added to each sample and incubated for 30 min. at 4°C in the dark. Samples were then
 10 washed once with PBS if a second color staining is to be done, otherwise washed twice more in PBS. For two color staining 5 µl of mouse serum was added to the cell pellets after the single wash, mixed, and incubated for five minutes at 4°C in the dark. Second primary antibodies (or PBS as a negative control) were added (10 µl anti-CD16, 100 µl 1:200 dilution of anti-IgE) and incubated for thirty minutes at 4°C in the dark. Samples were then washed
 15 one time with PBS and resuspended in 100 µL streptavidin PE (1:200 PBS + 1% BSA) and incubated for fifteen minutes at 4°C in the dark. Erythrocytes were lysed by adding 2 ml of FACS Lysing Buffer to each sample and incubating at room temperature in the dark for fifteen minutes or until samples were clear. The cells were pelleted by centrifugation and all but 200 µl of the supernatant was aspirated. The samples were analyzed by flow cytometry
 20 on a FacScan cytometer using the CellQuest software (Becton-Dickenson). CCR2b is expressed on most monocytes, a subpopulation of lymphocytes and a subset of granulocytes (Figure 2). CCR2b is expressed on an IgE-positive population in peripheral blood (basophils)(Figure 3).

25 MCP-1 Binding Assays

MCP-1 binding was performed in a final volume of 0.1 ml of 50 mM Hepes pH 7.4, 1 mM CaCl₂, 5 mM MgCl₂, 0.02% sodium azide, 0.5% BSA (HBB), containing either 2.5 µg THP-1 membrane protein or 500,000 PBMC and 0.1 nM of [¹²⁵I]-MCP-1. Competition binding experiments were performed by including variable concentrations of
 30 unlabeled MCP-1, 1D9 antibody, or a negative control IgG2a. Nonspecific binding was determined following the addition of a 2500-fold excess of unlabeled MCP-1. Samples were

incubated for 60 minutes at room temperature, and bound and free tracer were separated by filtration through 96-well GF/B filterplates presoaked in 0.3% polyethyleneimine. The filters were washed in HBB further supplemented with 0.5 M NaCl, dried, and the amount of bound radioactivity determined by liquid scintillation counting. mAb 1D9 inhibits [125I]MCP-1 binding to THP-1 cell membranes with an IC₅₀ of about 0.004 µg/ml (approximately 0.02 nM; Figure 4) and to fresh PBMC with an IC₅₀ of 0.04 µg/ml (approximately 0.2 nM; Figure 5).

Chemotaxis of PBMC

Chemotaxis was assayed using a 3 µm pore size 96-well chemotaxis plate (Neuroprobe, Cabin John, MD). PBMC isolated by standard methods using Ficoll-Hypaque density gradient centrifugation were washed with PBS/0.5%BSA and then resuspended in chemotaxis assay media (HBSS/10mM HEPES/0.5% Fatty acid free BSA) to a final concentration of 10 x 10⁶ cells/ml. Cells were preincubated in chemotaxis assay media at room temperature for 20 min. with various concentrations of the anti-CCR2 antibody, 1D9, or nonspecific murine IgG2a. The same dilutions of antibody were mixed with chemokine and 30 µl of the mixture was added to each of the bottom wells of the chemotaxis plate. The bottom wells are covered with the membrane, and 25 µl of the cell and antibody mixture are added to the top of the filter. The plates are incubated at 37°C in 5% CO₂ incubator for approximately 80 min. At the completion of the migration, the membrane is removed and the plate with the bottom wells is incubated -80C for 30 minutes to freeze the contents. The plates are thawed at 37°C for 10 minutes. 6 µl of a 1:400 dilution of CyQuant reagent (Molecular Probes, Eugene, OR) in a lysis buffer provided by the supplier is added to each well, and the cell migration is quantified as indicated by fluorescence intensity determined using a CytoFlour fluorescence plate reader at 485ex/535em. mAb 1D9 inhibits MCP-1-induced chemotaxis, but not RANTES-induced chemotaxis, of fresh PBMC (Figures 6A and 6B). Inhibition of MCP-1-induced chemotaxis of fresh PBMC has been demonstrated with 10 µg/ml (.40 nM).

Example 2Humanisation of monoclonal antibody 1D9

The 1D9 monoclonal antibody is likely to be immunogenic in humans, potentially eliciting a human anti-mouse antibody (HAMA) response. This HAMA response usually results in rapid clearance of the mouse monoclonal antibody from the body, thus limiting any therapeutic effect the 1D9 monoclonal antibody might have. Therefore, in an effort to reduce the immunogenicity of this antibody in humans and to maximize its therapeutic potential, the humanisation of the 1D9 mouse monoclonal antibody was undertaken. The following examples provide a detailed analysis of the 1D9 amino acid sequence data, the building of a molecular model of the murine 1D9 FV domain, and the design strategy for the successful humanization of the mouse antibody. This design strategy resulted in the design of a number of humanized versions of both the kappa light chain variable (VK) region and the heavy chain variable (VH) region. In total, the humanized VH region included up to 16 amino acid changes in the FRs of the selected human VH region. These changes were subdivided between four versions of the humanized VH region. In addition, twelve amino acid changes in the FRs of the selected human VK region were included in the four versions of the humanized VK region which were also designed.

Sequence analysis of the mouse 1D9 kappa light chain variable region

The amino acid sequence of the 1D9 VK region (Figure 7) was compared with other mouse kappa light chain variable regions and also the consensus sequences of the subgroups that the variable regions were subdivided into in the Kabat database (Kabat et al., Sequences of proteins of immunological interest, Fifth edition, U.S. Department of Health and Human Services, U.S. Government Printing Office (1991)). From this analysis the 1D9 VK region was found to most closely match the mouse consensus sequence of mouse kappa subgroup II (Identity = 79.46%, Similarity = 82.14%). When only the FRs of the 1D9 kappa light chain variable region were compared in the mouse subgroup II, percentage identity increased to 87.5%, while percentage similarity increased to 88.75%. In addition, the mouse 1D9 VK region showed good homology to a translation of the 70/3 murine VK germline gene (Figure 13). Taken together, the above evidence clearly proved that the 1D9 sequence was typical of a mouse VK region.

Sequence analysis of the mouse 1D9 heavy chain variable region

A similar analysis of the 1D9 VH region (Figure 8) found that it matched closest to the consensus sequence of the mouse heavy chain subgroup IIIc in the Kabat database (Kabat et al., Sequences of proteins of immunological interest, Fifth edition, U.S. Department of Health and Human Services, U.S. Government Printing Office (1991)). Identity between the mouse heavy chain variable region amino acid sequence of 1D9 and the consensus sequence of mouse subgroup IIIc was measured at 70.94%, while the similarity was calculated to be 76.07%. When only the FRs of the 1D9 VH region was compared to mouse subgroup IIIc, percentage identity increased to 75.86%, while the similarity increased to 80.46%. The mouse 1D9 VH region also showed good homology to a translation of the MLR-RF24BG murine VH germline gene, among others (Figure 14). Thus, the above evidence confirmed that the 1D9 sequence was typical of a mouse VH region.

Molecular modelling of the 1D9 domain

To assist in the design of the humanized variable regions of the 1D9 antibody, a series of molecular models were constructed of the murine 1D9 FV region and the eight CDR grafted variants. This was done using the AbM molecular modeling package supplied and utilized by Oxford Molecular Limited (OML). Antibody x-ray crystallographic structures available from the Brookhaven database were formatted to allow them to be used for modeling with AbM.

The FRs of the 1D9 variable regions were modeled on FRs from similar, structurally-solved immunoglobulin variable regions. While identical amino acid side chains were kept in their original orientation, mismatched side chains were substituted as in the original 1D9 FV. The backbone atoms of the FRs of the Fab Bv04-01 VK region were used for the model of the Fv framework region of 1D9 for both the VK and VH chains (Brookhaven PDB code 1nbv, solved to 2.0D). The sequence of Fab Bv04-01 was a good match for the variable region sequences of murine 1D9 and their humanized variants. The identities between Fab Bv04-01 and the murine 1D9 and humanized sequences ranged from 76% to 78% for VK sequences and from 74% to 84% for VH sequences. Testing of AbM with known structures has shown that FR backbone homology is an important factor in the

quality of any model, since the use of FR structures that poorly match a sequence being modeled can significantly and adversely affect the position and orientation of the CDR loops.

For the backbone structures of CDRs L1, L2, L3, H1 and H2, conformations for all of the models were taken from canonical classes used by AbM without modification, using the classes shown in Figures 9 and 10.

For the backbone structure of the L1 loop, the loop conformations of the murine 1D9 VK region was taken from canonical Class 4 from AbM. This canonical class is based on those described by Chothia and his colleagues (Chothia and Lesk, J. Mol. Biol. 197:901 (1987); Chothia et al., Nature 34:877 (1989); Tramontano et al., J. Mol. Biol. 215:175 (1990); and Chothia et al., J. Mol. Biol. 227:799 (1992)), but they have been modified to take into consideration structures that have become available since the original articles were published. Testing of the performance of AbM predictions for known loop structures has shown that CDR loops which are created in this way are usually modeled very accurately, i.e. to within 1-1.5Å RMS deviation.

The H3 loop in the 1D9 VH region is comparatively short at six residues long. It was modeled using a search for backbone conformations from X-ray structures in the Brookhaven databank. For short loops like this, there are sufficient loop conformations from known X-ray structures to saturate the conformational space available to the loop. Testing of the predictions made by AbM with the structures of new antibodies, where the structure is not included in the databases used by the program, shows that for CDR H3 loops of this size, the accuracy is likely to be at least 2.0Å.

After adjusting the whole of the model for obvious steric clashes it was subjected to energy minimisation, as implemented in MACROMODEL, both to relieve unfavorable atomic contacts and to optimize van der Waals and electrostatic interactions.

Design of the humanised 1D9 VK antibody variants

The first step in the design of the humanised variable regions of the 1D9 antibody was the selection of the human kappa light chain variable region that would serve as the basis for the humanized 1D9 VK, region. As an aid to this process the 1D9 VK region was initially compared to the consensus sequence of the four human kappa light chain variable region subgroups as defined by Kabat (Kabat et al., Sequences of proteins of

immunological interest, Fifth edition, U.S. Department of Health and Human Services, U.S. Government Printing Office (1991)). The mouse 1D9 light chain variable region was most similar to the consensus sequence of human kappa light chain subgroup II, with which it displayed a 76.2% identity over the whole variable region and a 82.5% identity within the FRs alone. When measured with respect to similarity, these values increased to 79.7% overall and 85.0% within the FRs alone. Consequently it generally appeared to match well to human kappa light chain variable region sequences from kappa subgroup II.

The mouse 1D9 VK was then compared to all the recorded examples of individual sequences of human variable regions publicly available. Figure 15 shows the best seventeen matches to the mouse 1D9 VK region which were identified through this analysis. Overall, the search algorithm selected the human VK region antibody 036521 (Rheinnecker et al., *Journal of Immunology*. 157(7):2989-97 (1996)) as the best match to the mouse 1D9 VK region (Figure 16). However, a review of the source paper for this antibody revealed that murine oligonucleotide primers had been used to rescue the genes from the hybridoma. This meant that this antibody was in fact a murine antibody and not human, as suggested by the Kabat database. Thus, the next best match to the murine 1D9 VK region that was selected by the database search was the human VK region from antibody HF-21/28 (Kabat database ID number 005056; Chastagner et al., *Gene*. 101(2):305-6 (1991)). The human sequence had an overall identity to the 1D9 VK region of 79.3% and 85.0% within the FRs alone. When measured with respect to similarity, these values increase to 83.99% overall and 87.5% within the FRs alone. In addition, key FR amino acids were more conservatively preserved in HF-21/28 VK region than in the other candidate human kappa light chain variable regions. Consequently, the HF-21/28 kappa light chain variable region FR was selected as the human acceptor sequence for the humanization of the 1D9 antibody kappa light chain variable region.

Unfortunately, the very last residue in FR4 (at position 107, according to the Kabat numbering system) of the human HF-21/28 VK region was not defined by the Kabat database or the authors who originally isolated this variable region sequence. Therefore, it was decided to insert the most commonly found amino acid at this position in the variable region sequences described by Kabat human kappa light chain subgroup 6-II (Kabat et al., *Sequences of proteins of immunological interest*, Fifth edition, U.S. Department of Health

and Human Services, U.S. Government Printing Office (1991)). Accordingly, lysine was added at position 107 in FR4 based upon an analysis of the Kabat database which found that 85.7% of the sequences in Kabat human kappa light chain subgroup K-II had a lysine at this position. This then became the basis of the first humanized version of the 1D9 kappa light chain (1D9RKA), which essentially comprised the CDR's of the 1D9 VK region and the FRs of HF-21/28 VK region. Figures 19A-19C define the amino acid sequence of this first CDR-grafted version of the humanised 1D9 VK region.

The next step in the design process was to study the amino acid sequences of the human acceptor HF-21/28 VK region FRs to determine if any of these amino acid residues were likely to adversely influence binding to antigen. This could be caused directly through interactions with antigen, or indirectly by altering the confirmation or orientation of the CDR loops. This was a difficult process which was only made possible through the availability of a model of the 1D9 variable regions, i.e., both the VK and the VH regions. Nevertheless, any amino acids in the mouse 1D9 FRs that did appear to affect antigen binding were then considered for conversion in the humanized 1D9 antibody. In deciding which murine residues to conserve the following points were addressed:

It was of great importance that the canonical structures of the hypervariable loops (Chothia and Lesk, J. Mol. Biol. 197:901 (1987); Chothia et al., Nature 34:877 (1989); Tramontano et al., J. Mol. Biol. 215:175 (1990); and Chothia et al., J. Mol. Biol. 227:799 (1992)) were conserved. Consequently, it was crucial to conserve in the humanized 1D9 variable regions all the mouse FR residues that were part of these canonical structures (Figures 9 and 10).

The sequences of the 1D9 antibody variable regions were compared to similar sequences from other mouse antibodies to identify unusual or rare residues which may have indicated an important role in antigen binding. This was then investigated using the mouse model of the 1D9 variable region genes.

A direct analysis of the model was also made to try and predict whether any of the other mouse FR residues not present in the humanized FRs could influence antigen binding in some way.

Comparisons of the individual human acceptor sequences for the kappa light chain and heavy chain variable regions to the consensus sequence of human variable regions subgroups to which the acceptor sequences belonged were also made. The identification of any idiosyncratic amino acids in the human donor sequences was important, as these could have adversely affected antigen binding.

Since the human light and heavy chain variable regions selected would be derived from two different human antibodies, a careful analysis of the interdomain packing residues of both the donor and the acceptor kappa light chain variable regions should be carried out (Chothia et al., J. Mol. Biol. 186:651 (1985)). This was because any mispacking in this region could have had a dramatic affect upon antigen binding, irrespective of the conformation of the CDR loop structures of the humanized 1D9 antibody.

Although there were 12 amino acid differences between the FRs of the donor mouse 1D9 VK region and the acceptor human HF-21/28 VK region, only two mouse residues were considered sufficiently important for binding affinity to preserve them in the humanised FRs. The first of the FR changes that were introduced into 1D9RKB was located at position 36. This residue is a Vernier residue (Foote and Winter, J. Mol. Biol. 224:487 (1992)) and is predicted to be a key structure determining residue for L1 loop structure as defined by Chothia and his coworkers (Chothia and Lesk, J. Mol. Biol. 197:901 (1987); Chothia et al., Nature 34:877 (1989); Tramontano et al., J. Mol. Biol. 215:175 (1990); and Chothia et al., J. Mol. Biol. 227:799 (1992)). Although both residues are hydrophobic, the human Phe is bulkier at this position, and X-ray structures with Leu and Phe at this position show that if Phe is present steric hindrance causes the side chain at 34Asn to point in the opposite direction. Thus, it was considered critical for the successful humanization of the 1D9 kappa light chain.

The second change incorporated into the 1D9RKB humanized version was at residue 37, i.e., Gln37Leu. Although this was a conservative change it occurred in a highly conserved region at the base of CDR1. It was thought that by preserving this murine Leu residue in this version, alongside the murine Leu residue at position 36, the affinity of the humanised antibody could be preserved.

Two other versions of the humanised VK region were also considered for construction to explore the structural and binding affinity consequences of manipulating the FRs of the humanised 1D9 antibody. 1D9RKC was essentially identical to 1D9RKB, except for the mutation Gln100Gly. There is a dramatic difference in molecule bulk between these two residues, and thus this version was made to explore the consequences of this change to the FRs of the reshaped human kappa light chain on antibody structure and overall antibody affinity. 1D9RKD contained the modifications described in 1D9RKC and, in addition, contained the FR change Gln17His. Although Gln and His are similar in size and both are weakly polar, the mouse residue (His) at this position is extremely rare amongst all mouse VK sequences (0.07% overall, but has not been seen in mouse Kabat subgroup II sequences) and has never been seen in any human VK sequences. Conversely, the Gln residue is more commonly seen at this position in both mouse (16.16% overall and 6.12% in mouse Kabat subgroup II sequences) and human (5.00% overall and 39.7% in human Kabat subgroup II sequences) sequences. Thus, the simple rarity of the His at this position suggests that it may be important for binding,

although there is no clear evidence to support this from the molecular modeling data.

A description of the amino acid sequences of all the humanised 1D9 antibody VK region variants proposed above are given in Figure 11.

Design of humanised 1D9 VH antibody variants

Once again, the first step in the design of the humanised VH region of the mouse 1D9 antibody was the selection of the acceptor human heavy chain variable region that would serve as the basis of the humanised 1D9 VH region. When the 1D9 VH region was initially compared to the consensus sequences of the three human heavy chain variable region subgroups it was found to be most similar to the consensus sequence for the human heavy chain subgroup III, with a 69.231% identity overall and a 78.161% identity between

the FRs alone. When measured with respect to similarity, these values increased to 74.359% overall and to 82.759% within the FRs alone.

The mouse 1D9 VH region was then compared to all the recorded examples of individual sequences of human variable regions publicly available. Figures 17A-B show the best 24 matches to the mouse 1D9 VH region which were identified through this analysis. Overall the search algorithm selected the human VH region from antibody 4B4'CL (Kabat data base ID number 000490; Sanz et al., Journal of Immunology. 142:883 (1989)) as the best match to the mouse 1D9 VH region. The VH region of this clone had an overall identity to the 1D9 VH region of 67.2%, a value which increased to 80.95% when the FRs alone were compared (Figures 18A-B). When measured with respect to similarity, these values increased to 69.66% overall and to 84.52% within the FRs alone. Thus, although once again not the most homologous of the potential human acceptor VH sequences, this human FR became the basis of the humanised version of the 1D9 heavy chain.

The next step in the design process was to study the amino acid sequences of the human acceptor 4B4'CL VH region FRs to determine if any of these amino acid residues were likely to adversely affect binding to antigen. Once again the molecular models built by OML were important in this design process, from which a number of amino acids in the murine 1D9 VH region FRs were identified for conversion in the first (1D9RHA) and subsequent versions of the humanised 1D9 antibody (Figure 12 and Figures 20A-C). There were 16 amino acid differences between the FRs of the donor mouse 1D9 and the acceptor human 4B4'CL VH regions, and up to 5 murine residues were considered for conservation in the humanised FRs (Figure 12).

1D9RHA consisted of the CDRs of the murine 1D9 antibody VH region genetically inserted into the FRs of the human 4B4'CL antibody VH region. 1D9RHB was identical to version 1D9RHA apart from two FR1 mutations, Thr28Ser and Ser30Asn. These changes were made because they represented Vernier amino acids as defined by Foote and Winter (J. Mol. Biol. 224:487 (1992)), which were thought to be critical for H1 loop conformation. Residues 27-30 are considered part of the H1 loop itself and so are even more critical to the correct conformation and orientation of this loop, justifying their conservation even more strongly. Thus, these two residues represented the sum of the changes made to the FRs of the human 4B4'CL VH sequence in 1D9RHB. 1D9RHC was identical to version

1D9RHB except that it contained two further changes at positions Gly49Ala and Phe67Tyr. The Gly49Ala was a conservative change. However, residue position 49 has been identified as a Vernier residue (Foote and Winter, J. Mol. Biol. 224:487 (1992)), important for H2 hypervariable loop structure, so it was decided to conserve the murine Ala residue in this version. Residue position 67 was also a Vernier residue position, identifying it as important for maintaining CDR loop conformation. Tyr is very rarely seen in human VH sequences (0.08% overall) and has not previously been found in murine VH regions at this position. Consequently, it must have arisen through somatic mutation. Thus, given its location close to CDR2 according to the molecular model and its Vernier residue status, it was decided to conserve the murine Tyr residue at this position. 1D9RHD was identical to 1D9RHC except for a Thr93Val mutation. This residue had been identified as important as both a VH/VK packing residue (Chothia et al., J. Mol. Biol. 186:651 (1985)). Moreover, its buried position between CDR loops H1 and H3, according to the molecular model, supported the decision to conserve the murine Val residue at this position. A description of the amino acid sequences of all the humanised VH region variants described above are given in Figure 12.

Inhibition of MCP-1 Binding by Humanized Version of 1D9

Figure 25 illustrates the ability of murine mAb 1D9 and a humanized version of mAb 1D9 comprising the 1D9RHAVH heavy chain (Figure 12) and the 1D9RKAVK light chain (Figure 11) to inhibit the binding of [125I]-MCP-1 to whole THP-1 cells. 0.5 X 10⁶ THP-1 cells were incubated in 50 mM HEPES, 1 mM CaCl₂, 5 mM MgCl₂, 0.1% BSA, 0.05% sodium azide (binding buffer) with different dilutions of antibody samples for 10 minutes at 37EC. An equal volume of [125I]-MCP-1 in binding buffer was added to a final concentration of 0.1 nM and incubated for a further 30 minutes at 37EC. Cells were diluted and vortexed with an equal volume of binding buffer with 0.5 M NaCl (wash buffer) and pelleted by centrifugation (benchtop centrifuge, 7000 rpm, 2 minutes). After removal of the supernatant, the pellet was vortexed in 200 µL of wash buffer, spun as before, and the supernatant removed. The cells were resuspended in 100 µL wash buffer and counted on a gamma counter (Cobra, Packard Instruments). Data analysis was performed using Graphpad software.

Example 3Creation of Combined Immunoglobulin DNA Cassette Vectors

A pcDNA3 was used as a backbone vector which contains the gene for resistance to G418 (NEO) to allow for easy selection in research conditions. The SpeI restriction site was eliminated from pcDNA3 by site directed mutagenesis. The EF-1a promoter from the plasmid pcDEF3 (originally pBOS) was inserted into pcDNA3, thus eliminating the CMV promoter.

A BamHI site in the pcDNA3 cloning linker and an Mfe I cloning site within the EF-1a promoter were removed using site directed mutagenesis, and a BamHI site was added 3' in the polyA region. This allowed the combination of the heavy and light chain active regions in a single vector and would allow for the addition of any other selectable marker including the gene cassette DHFR that confers resistance to methyloxymethylamine.

The heavy chain cassettes (including its promoter and polyadenylation region) were subcloned from the various single cassette vectors into their corresponding light chain cassette vectors to create the combined two cassette vectors (Figure 26). The combinations are summarized in Table 1. All the combination vectors had a similar overall structure that is shown in Figure 27.

The pLKTOK58 vector, created by combining pLKTOK55 and pLKTOK57, can be used to produce an antibody that contains a human kappa constant region and human IgG1 constant region in its native form. The pLKTOK59 vector, created by combining pLKTOK56 and pLKTOK57, can be used to produce an antibody that contains a human kappa constant region and a human IgG1-FcRmut constant region contains the mutations at positions L235A and G237A. These mutations inhibit the binding of the constant region to human Fc receptors and inhibit the initiation of ADCC reactions. Such mutation have been described previously in US Patent No.: 5,985,279 and International Publication No.: WO98/06248, which are incorporated herein by reference. The pLKTOK92 and pLKTOK73 vectors, created by combining pLKTOK55 or pLKTOK56 and pLKTOK72, can be used to produce antibodies that contain human lambda constant regions and either human IgG1-WT (pLKTOK92) or human IgG1-FcRmut (pLKTOK73).

The vectors pLKTOK68 and pLKTOK69, created by combining pLKTOK65 or pLKTOK66 and pLKTOK67, can be used to produce antibodies that contain a macaque

kappa constant region and macaque IgG1 constant region. The constant region of pLKTOK68 is IgG1-WT for both cynomologous and rhesus macaques. The constant region of pLKTOK69 is IgG1-FcRmut that contains the L235A and G237A mutations and theoretically should inhibit the binding of the constant region to macaque FC receptors. The kappa constant region is one of the two kappa constant regions expressed by both the cynomologous and rhesus macaques thus should be recognized as native by both monkey species.

The vectors pLKTOK63 and pLKTOK64, created by combining pLKTOK60 or pLKTOK61 and pLKTOK62, can be used to produce antibodies that contain a mouse kappa constant region and mouse C57BL/6 IgG2a constant region. The constant region of pLKTOK63 is the native conformation of IgG2a in mouse and is the allotype most closely matched in structure and function to human IgG1. The constant region of pLKTOK64 contains the murine IgG1 with the mutations at positions L235A and G237A (for Fc region I) and E318A (for Fc region II) to inhibit the binding of the constant region to mouse FC receptors and inhibit the initiation of ADCC reactions (See, Isaacs JD, et al. Therapy with monoclonal antibodies. II. The contribution of Fc gamma receptor binding and the influence of C(H)1 and C(H)3 domains on in vivo effector function. 1998. J Immunol. 161(8):3862-9.)

Table 1. Composition of combined expression vectors

COMBINED VECTOR	HEAVY CHAIN VECTOR	HEAVY CHAIN COMPOSITION	LIGHT CHAIN VECTOR	LIGHT CHAIN COMPOSITION
pLKTOK58	pLKTOK55	nVHL – Human IgG1-WT	pLKTOK57	nVKL – Human C Kappa
pLKTOK59	pLKTOK56	nVHL – Human IgG1-FcRmut	pLKTOK57	nVKL – Human C Kappa
pLKTOK92	pLKTOK55	nVHL – Human IgG1-WT	pLKTOK72	nVLL – Human C Lambda
pLKTOK73	pLKTOK56	nVHL – Human IgG1-FcRmut	pLKTOK72	nVLL – Human C Lambda
pLKTOK68	pLKTOK65	nVHL – Macaque IgG1-WT	pLKTOK67	nVKL – Macaque C Kappa
pLKTOK69	pLKTOK66	nVHL – Macaque IgG1-FcRmut	pLKTOK67	nVKL – Macaque C Kappa
pLKTOK63	pLKTOK60	nVHL – Mouse IgG2a-WT	pLKTOK62	nVKL – Mouse C Kappa
pLKTOK64	pLKTOK61	nVHL – Mouse IgG2a-FcRmut	pLKTOK62	nVKL – Mouse C Kappa

Example 4Creation of DNA Cassette Insert Sequences: Adapting antibody variable regions

5 To test these vectors by creating intact antibodies, the variable regions from the monoclonal antibody 1D9 were PCR adapted to add the desired restriction enzyme sites (MfeI and BlnI for the VH, PpuMI and BsiWI for the human and macaque VK and PpuMI and ClaI for the mouse VK. The 1D9 antibodies have been described previously in International Publication Nos: WO 00/05265 and WO 01/57226, which are incorporated
10 herein by reference. Once adapted, these and any other set of variable regions could be cloned into the various expression vectors.

The primers for the VH region are designed such that the 5' primer includes the 7 codons at the end of the VH leader (including the Mfe I restriction enzyme) and the first 7-9 codons of the hybridoma VH. The 3' primer included 7-9 codons of the hybridoma VH
15 followed by 3 codons of the IgG1 constant region (including the BlnI restriction enzyme). Primers used for adapting variable regions are demonstrated in Table 2, (upper case letters depict identical sequences for all antibodies encoded in vectors; lower case letters are determined by the sequence of the individual antibody).

The primers for the VK region are designed such that the 5' primer includes
20 the 6 codons at the end of the VK leader (including the PpuM I restriction enzyme) and the first 7-9 codons of the hybridoma VK (Table 2). The human 3' primer included 7-9 codons of the hybridoma VH followed by 4 codons of the human kappa constant region (including the BsiWI restriction enzyme). The mouse 3' primer included 7-9 codons of the hybridoma VH followed by 12 codons of the mouse kappa constant region (including the ClaI restriction
25 enzyme) (Table 2). The primers used to adapt the hybridoma 1D9 VH gene were n1D9VH5 and n1D9VH3. The primers used to adapt the hybridoma 1D9 VK gene were n1D9VK5 and n1D9VK3. The adapted 1D9 VH and VK genes were cloned into the vectors in standard two step cloning to create the complete plasmids (Figure 28).

30 Table 2. Primers to adapt 1D9 VH and VK for cloning into all expression vectors.

(Upper case letters are identical for all antibodies. Lower case letters are determined by the sequence of the individual antibody.)

PRIMER	SEQUENCE
n1D9VH5	5' TTACCCAATTGTGTCTGTCCgaggtgcagcttgtagtctg 3' SEQ ID NO:
n1D9VH3	5' GTTTTAGGCTGAGCTgacggtgaccgtggccctgtg 3' SEQ ID NO: 119
n1D9VK5	5' TTCCCAGGGTCCCGTTCCgatgttgatgaccagact 3' SEQ ID NO: 120
n1D9VKhum3	5' AGCCACCGTACGCtttattccagcttggtcc 3' SEQ ID NO: 121
n1D9VKmur3	5'TGGGAATATCGATACAGTTGGTGCAGCATCAGCACGCtttatttc cagcttggtcc 3' SEQ ID NO: 122

Example 5

Testing the System: Determining Relative Rates of Antibody Production

The protein production level, the rate of correct folding and the rate of transportation out of cells determine high production of secreted proteins. A major component of protein production is the function of its promoter.

For the successful folding and transport of antibodies out of cell, two proteins must be produced and properly associate. It is believed that having similar concentrations of the heavy and light chains may assist in the association, folding and transportation of the antibodies. An excess of one antibody chain within a cell can lead to cell death. We believed that creating cassettes in which each antibody chain had its own promoter through polyadenylation site would increase the likely-hood of equivalent protein production of the two chains. This system also uses cDNA inserts that eliminate the need for post-translational modification. The decreased variability may reduce the levels of other RNA species and increase the levels of the desired RNA species.

To test this system and determine optimal promoter combinations, vectors were created with either the CMV or the EF-1a promoters. (See U.S. Patent Nos.: 5,225,348 and 5,266,491; Mizushima S, Nagata S. pEF-BOS, a powerful mammalian expression vector. 1990 Nucleic Acids Res 18(17):5322.) In both cases, restriction sites that would interfere with cloning within these cassettes were removed by site directed mutagenesis. The SpeI cloning site was removed from CMV and the MfeI cloning site was removed from EF-1a. Testing with an EGFP cassette demonstrated these changes did not make any noticeable

changes in the general function of the promoters. A vector was also created with a combination of the mutated CMV promoter with the beta-kinesin IRES to see if that increased antibody production.

The vector pcDNA3 was used as a backbone vector because it contains the gene for resistance to G418 (NEO) to allow for easy selection in research conditions. Into this vector was cloned the mutated CMV promoter, the mutated CMV promoter with the beta-kinesin IRES or the mutated EF-1a promoter.

Changes to aid in cloning were made to the vector backbone including the removal of a MfeI cloning site 5' of the promoter. A BamHI site in the pcDNA3 cloning linker was removed and a BamHI site was added 3' of the polyadenylation region. This flanking BamHI site allows the transfer of the heavy chain cassettes (as a BglII/BamHI fragment) including their promoters and polyadenylation regions into vectors containing the light chain cassettes. As a single BamHI site remains after the cloning, it allows for the later addition of any other selectable marker including the gene cassette for DHFR, which confers resistance to methotrexate.

Three matched vectors were constructed to test the system. Each was a single vector with a light chain cassette (with its promoter and polyadenylation region) followed by a heavy chain cassette (with its promoter and polyadenylation region). All three vectors contain the 1D9 VK functionally attached to the human C Kappa gene and the 1D9 VH functionally attached to the human IgG1-FcRmut gene by the method described in the construction of the inserts. The vectors differed only in their promoter combination. The vector pLKTOK34 had both the light and heavy chain cassettes driven by the mutated CMV promoter. The vector pLKTOK36 had both the light and heavy chain cassettes driven by the mutated CMV promoter combined with the beta-kinesin IRES. The vector pLKTOK38 had both the light and heavy chain cassettes driven by the mutated EF-1a promoter.

Completed vectors were transfected into CHO cells and selected in G418 media. This media results in the death of all cells that do not contain the gene for Neomycin resistance. After 5 days of selection in G418, cells were trypsinized to make a single cell suspension and plated in 96 well plates at the rate of 1, 5 or 10 cells/well. At 5 days of selection, most of the G418 non-resistant cells will be programmed to die. These clones were expanded and tested repeatedly to determine their ability to retain high production levels.

After 10 days in the 96 well plates, the plates are scored visually to select the wells that contain a single CHO clone. At two weeks, these are tested for their ability to produce and secrete intact antibody as measured by an ELISA that coats the wells with a Fab' against human heavy and light and develops with Protein A attached to the enzyme HRP.

In various transfections, 50-70% of the tested single clone wells were producing antibody in significant amounts and usually 70% + would retain production after transfer.

The best clones were tested on their ability to produce antibody over a 5-day period both with and without butyric acid (necessary to amplify CMV production). It was determined that the CMVmut cells produced an average of 0.5 ug/ml without butyric acid treatment and 1.8 ug/ml with butyric acid treatment. The CMVmut/IRES cells produced an average of 0.7 ug/ml without butyric acid treatment and 0.2 ug/ml with butyric acid treatment. The EF-1a cells produced an average of 122.6 ug/ml without butyric acid treatment and 50.2 ug/ml with butyric acid treatment. The results demonstrated that the EF-1a promoter produced 100x the amount of antibody.

It is quite possible that the two chains of the antibody driven by their own promoters and having their own polyadenylation regions allows for the two proteins to be produced at similar rates. Having similar amounts of the two chains probably assists in the correct and rapid association, folding and transportation out of the CHO cells. Functional testing of the antibodies generated demonstrated that the antibodies produced using the present system function in similar fashion to those produced by the original 1D9 hybridoma

While this invention has been particularly shown and described with references to preferred embodiments thereof, it will be understood by those skilled in the art that various changes in form and details may be made therein without departing from the scope of the invention encompassed by the appended claims.